1 canals throughout the undulating terrain (DWR 1999). This high permeability causes 2 water levels in the ponds and canals to rise and fall according to the stage of the Yuba 3 River. Generally, water from the Yuba River enters the Goldfield area from above 4 Daguerre Point Dam, then migrates down-gradient through the Yuba Goldfields. A 5 portion of this migrating water eventually returns to the Yuba River approximately one 6 mile downstream of Daguerre Point Dam via an outlet canal, referred to as Waterway 13, 7 the origin of which is uncertain. This outlet canal helps to drain water out of the 8 Goldfields to the Yuba River, which prevents high water levels from adversely impacting 9 current mining and aggregate operations (DWR 1999).

10 During the fall of 1988 and the winter and spring of 1989, adult fall/late fall-run Chinook 11 salmon and American shad were observed in the area of the Yuba Goldfields (USFWS 12 1990). It was suggested that these fish were attracted into the area via the outfall channel 13 referred to as Waterway 13. In 1989, the Red Bluff Fisheries Assistance Office 14 conducted a fishery investigation in the Yuba Goldfields area near Daguerre Point Dam 15 on the lower Yuba River. Several hundred fall-run Chinook were observed spawning in 16 the open access channel in December 1988. In the 1980s, it was discovered that adult 17 anadromous fish (Chinook salmon, American shad, and steelhead) had migrated into the 18 interconnected ponds and canals of the Yuba Goldfields via the area's outlet canal. 19 USFWS (1990) also observed a pair of spawning late fall-run Chinook salmon during 20 March 1989.

21 Salmon spawning habitat in the Yuba Goldfields was observed in several interconnecting 22 stream channels between ponds, and numerous fall-run Chinook salmon redds were 23 observed (USFWS 1990). From February through April 1989, USFWS (1990) captured 24 241 juvenile Chinook salmon in the Yuba Goldfields ponds with beach seines at five 25 sample sites located in ponds downstream of the spawning area. In May 1989, juvenile 26 sampling was terminated when reduced flows through the ponds prevented access to the 27 sampling sites. The juveniles ranged in size from about 30 to 65 mm, with the average 28 fork length about 40 mm (USFWS 1990). It was suggested that these small individuals 29 would have a poor chance of survival because increasing water temperatures during May

would likely increase predation rates from the numerous adult squawfish and bass
 observed in the ponds (USFWS 1990).

SWRCB (2000) reported that on various occasions CDFW staff also observed from a few 3 4 fish to several hundred adult fall-run Chinook salmon attracted up through the outfall into 5 the Yuba Goldfields in the late 1990s. Attraction of adult fall-run Chinook salmon was 6 of concern because there is a general lack of spawning habitat in the Yuba Goldfields, 7 and water temperatures in the Yuba Goldfields can be unsuitable, especially in the lower 8 ends where water discharges into the lower Yuba River (SWRCB 2000). Additionally, 9 fish habitat within the ponds and canals is not conducive to anadromous fish survival 10 because food supply is limited, predator habitat is extensive, and water quality 11 conditions, especially water temperature, are poor (DWR 1999).

12 There have been several past attempts at taking actions to preclude anadromous 13 salmonids from entering the Yuba Goldfields (SWRCB 2000). In the early 1980s, a large 14 grate was placed on the outfall of Waterway 13 to preclude fish from entering the Yuba 15 Goldfields. However, no one maintained the grate and it was damaged by debris. Thus, 16 adult salmon and steelhead continued to access the Yuba Goldfields. During the January 17 1997 floods, flows through the Yuba Goldfields became so high that they washed out the 18 structure (SWRCB 2000). The entry point remained open for several years. Realizing 19 that adult fish were once again entering the Yuba Goldfields, CDFW worked with a local 20 aggregate company to install a temporary aggregate berm to exclude adult fish, which 21 was effective for several years. However, any time there is high water in the Yuba 22 Goldfields, the barrier can be breached and activities to replace that barrier cannot begin 23 until the summer or late spring (SWRCB 2000).

The USFWS provided funding for an investigation through the AFRP, and engineering design and environmental evaluation of an adult fish barrier in Waterway 13 that would meet the resource needs of CDFW, USFWS, and NMFS, as well as the needs of the Goldfields' owners - Western Aggregates and Cal-Sierra Development was conducted (SWRCB 2000). Design objectives for a fish barrier located in the Yuba Goldfields outlet canal included the following: (1) prevent adult anadromous fish from entering the Yuba Goldfields; (2) not increase water elevations within the Yuba Goldfields; (3)

1 require minimal maintenance; and (4) allow for passage or removal of debris (DWR 2 1999). The primary project objective was to prevent adult anadromous fish from entering 3 the Yuba Goldfields through the outlet canal. The outlet canal is especially important 4 during periods of high flows, when the outlet canal must be able to pass high flows in 5 order to prevent flooding in nearby low-lying areas. It is also important that flows not be 6 If flows during these periods are greatly restricted during non-flood conditions. 7 restricted, water elevations within the Yuba Goldfields rise, adversely affecting Yuba 8 Goldfields mining operations. Consequently, a project needed to be designed to 9 accommodate high flows exiting the Yuba Goldfields. In addition, this project needed to 10 be low maintenance and allow for the passage or removal of debris (DWR 1999). Outlet 11 canal flows during summer and fall months were estimated to range from five to 50 cfs, 12 whereas canal flows during winter and spring months can exceed 1,000 cfs (DWR 1999).

13 In 2002, the BLM signed a Finding of No Significant Impact for the Yuba Goldfields 14 Fish Barrier Replacement Project. The BLM approved the replacement of the original 15 structure in the same location as the previous structure. The construction of a temporary 16 rock embankment was completed in September 2003 (Figure 5-9). In May 2005, heavy 17 rains and subsequent flooding breached the structure at the east (upstream facing) end. 18 AFRP funding was available to repair the "plug" (i.e., temporary aggregate berm) but, 19 because there was no project proponent to do the necessary work, YCWA facilitated the 20 effort but did not accept any responsibility for construction, operation or maintenance (C. 21 Aikens, YCWA, pers. comm. 2011). A "leaky-dike" barrier (Figure 5-10) intended to 22 serve as an exclusion device for upstream migrating adult salmonids was constructed at 23 the outfall of Waterway 13 (AFRP 2010).

Although most of the area encompassing the Yuba Goldfields is located on private land, it has been determined that the rock weir plug on Waterway 13 is located on Corps property. However, the Corps does not have any operations or maintenance responsibilities for the earthen "plug" and Waterway 13, nor has it issued any permits or licenses for it. Thus, operation and maintenance of Waterway 13 is part of the Environmental Baseline, and is not part of the Proposed Action.



1 2 3

Figure 5-9. Yuba Goldfields barrier located at the outfall of Waterway 13 (Source: AFRP 2011).



4

5 Figure 5-10. Location of the Waterway 13 "leaky-dike" barrier prior to it washing out during 6 the spring of 2011 due to high flows through the Yuba Goldfields.

1 During the spring of 2011, high flows (~30,000 cfs) in the lower Yuba River and high 2 flows through the Yuba Goldfields once again caused the "leaky-dike" barrier at the 3 entrance to Waterway 13 to wash out. In response to this recent loss of the "leaky-dike" 4 barrier at Waterway 13, the Corps conducted a real estate investigation and determined 5 that Waterway 13 is located on lands that are under the Corps' jurisdiction. As a separate 6 action unrelated to this ESA consultation, the Corps will work with local stakeholders 7 and resource agencies to identify potential biological concerns associated with Waterway 8 13 and will support the development of measures to repair the barrier. If needed in the 9 future, the Corps will collaborate with the stakeholders involved to develop a shared 10 agreement (e.g., a right-of-way or easement) that would provide access to those parties 11 that would conduct future maintenance activities that may become necessary if and when 12 the fish barrier at Waterway 13 washes out again in the future. However, because these 13 activities would occur in the future, and a project has not been proposed at this time, 14 Waterway 13 activities are not part of the Proposed Action.

15 **5.3.3 Riparian Vegetation**

16 Most of the original plant communities along the lower Yuba River have been 17 significantly altered from pristine conditions (Corps 1977 as cited in CDFG 1991). 18 Although little has been written specifically about the ancestral riparian forests of the 19 lower Yuba River, it is believed that the banks of the lower Yuba River and its adjacent 20 natural levees once were covered by riparian forest of considerable width. It has been 21 suggested that most riverine floodplains in the Central Valley supported riparian 22 vegetation to the 100-year floodplain, and it is likely that the Yuba River was no 23 exception (CDFG 1991).

In its Final Biological and Conference Opinion for the Yuba River Development Project License Amendment (FERC No. 2246), NMFS (2005b) reports that "The deposition of hydraulic mining debris, subsequent dredge mining, and loss/confinement of the active river corridor and floodplain of the lower Yuba River which started in the mid-1800's and continues to a lesser extent today, has eliminated much of the riparian vegetation along the lower Yuba River. In addition, the large quantities of cobble and gravel that 1 remained generally provided poor conditions for re-establishment and growth of riparian 2 vegetation. Construction of Englebright Dam also inhibited regeneration of riparian 3 vegetation by preventing the transport of any new fine sediment, woody debris, and 4 nutrients from upstream sources to the lower river. Subsequently, mature riparian 5 vegetation is sparse and intermittent along the lower Yuba River, leaving much of the 6 bank areas unshaded and lacking in large woody debris."

7 To determine the cumulative change over time in total vegetative cover and riparian 8 vegetation cover in the lower Yuba River, YCWA compared aerial photographs from 9 1937 and 2010. Over this time period, riparian vegetation cover in the Englebright Dam 10 site decreased over time, and the Narrows study site exhibited little detectable change 11 over time. For the remaining study sites distributed throughout the lower Yuba River, 12 riparian vegetation cover increased over time. Dramatic increases in riparian vegetation 13 cover were observed for the Dry Creek and Parks Bar study sites.

14 Riparian habitats support the greatest diversity of wildlife species of any habitat in 15 California, including many species of fish within channel edge habitats (CALFED 2000a 16 as cited in RMT 2013). Furthermore, more extensive and continuous riparian forest 17 canopy on the banks of estuaries and rivers can stabilize channels, provide structure for 18 submerged aquatic habitat, contribute shade, overhead canopy, and instream cover for 19 fish, and reduce water temperatures (CALFED 2000a as cited in RMT 2013).

20 Although fish species do not directly rely on riparian habitat, they are directly and 21 indirectly supported by the habitat services and food sources provided by the highly 22 productive riparian ecosystem. Riparian communities provide habitat and food for 23 species fundamental to the aquatic and terrestrial food web, from insects to top 24 predators. As stated in CALFED and YCWA (2005), riparian vegetation, an important 25 habitat component for anadromous fish, is known to provide: (1) bank stabilization and 26 sediment load reduction; (2) shade that results in lower instream water temperatures; (3) 27 overhead cover; (4) streamside habitat for aquatic and terrestrial insects, which are 28 important food sources for rearing juvenile fishes; (5) a source of instream cover in the 29 form of woody material; and (6) allochthonous nutrient input. Riparian vegetation on

floodplains can provide additional benefits to fish when the floodplain is inundated, by
 providing velocity and predator refugia.

In 2012, YCWA conducted a riparian habitat study in the Yuba River from Englebright Dam to the confluence with the Feather River (YCWA 2013). Field efforts included descriptive observations of woody and riparian vegetation, cottonwood inventory and coring, and a LWM survey. The RMT contracted Watershed Sciences Inc. to use existing LiDAR to produce a map of riparian vegetation stands by type. The resulting data was subject to a field validation and briefly summarized in WSI (2010 as cited in RMT 2013) and the data were also utilized in YCWA (2013).

Based on field observations, YCWA (2013) reported that all reaches supported woody species in various lifestages - mature trees, recruits, and seedlings were observed within all reaches. Where individuals or groups of trees were less vigorous, beaver (*Castor canadensis*) activity was the main cause, although some trees in the Marysville Reach appeared to be damaged by human camping.

15 The structure and composition of riparian vegetation was largely associated with four 16 landforms. Cobble-dominated banks primarily supported bands of willow shrubs with 17 scattered hardwood trees. Areas with saturated soils or sands supported the most 18 complex riparian areas and tended to be associated with backwater ponds. Scarps and 19 levees supported lines of mature cottonwood and other hardwood species, typically with 20 a simple understory of Himalayan blackberry or blue elderberry shrubs. Bedrock 21 dominated reaches had limited riparian complexity and supported mostly willow shrubs 22 and cottonwoods.

The longitudinal distribution of riparian species in the lower Yuba River downstream of the Englebright Dam shows a trend of limited vegetation in the confined, bedrock areas, with increased vegetation in the less-confined, alluvial areas downstream, which is within expected parameters (Naiman et al. 2005 as cited in YCWA 2013). The increase in hardwood diversity and cover downstream of Daguerre Point Dam may be associated with sediment, as reaches above the Daguerre Point Dam have greater scour, while the downstream reaches have more deposition (YCWA 2012a). 1 Cottonwoods are one of the most abundant woody species in the Action Area, and the 2 most likely source of locally-derived large instream woody material due to rapid growth 3 rates and size of individual stems commonly exceeding 2 feet in diameter and 50 feet in 4 length. Cottonwoods exist in all life stages including as mature trees, recruits, or 5 saplings, and as seedlings. Cottonwoods are more abundant in downstream areas of the 6 Action Area relative to upstream. Of the estimated 18,540 cottonwood 7 individuals/stands, 12% are within the bankfull channel (flows of 5,000 cfs or less), and 8 39% are within the floodway inundation zone (flows between 5,000 and 21,100 cfs).

9 The RMT conducted a LiDAR survey of the lower Yuba River from Highway 20 to the 10 confluence, and digitized the patches of vegetation in recent aerial imagery of Timbuctoo 11 Bend and the Englebright Dam Reach (Pasternack 2012). With respect to having 12 sufficient riparian vegetation to provide ecological functionality, the RMT conducted 13 paired hydrodynamic modeling of the lower Yuba River in which one set of models lacks 14 vegetation and the other represents the actual lower Yuba River vegetation pattern and 15 height as best as possible. As shown at the 2011 Lower Yuba River Symposium and in 16 RMT meeting presentations, vegetation was found to significantly affect the hydraulics of 17 the lower Yuba River and, thus, may be deemed present in a significant quantity relative 18 to that functionality (Pasternack 2012).

19 YCWA (2013) assessed the riparian communities in the Yuba River downstream of the 20 Englebright Dam as healthy and recovering from historical disturbance. Historical aerial 21 photograph analysis indicates that vegetation cover has increased over time, with short-22 term decreases associated with stochastic flow events, which are normal for riparian 23 systems, and anthropogenic channel changes. Although the riparian vegetation is healthy 24 (plants have high vigor and are present in all age classes), the vegetation communities 25 tend to be simplistic in structure. Riparian communities are seral, establishing first with 26 simplistic herb and shrub layers, then canopies of hardwood trees, and becoming more 27 complex over time. Indicative of early seral stages, the assessed riparian communities 28 tended to be simplistic in both lateral and horizontal stratification, with limited pockets of 29 diverse and well-stratified riparian forests (YCWA 2013). As an example, bands of 30 willows on the floodplains, with some alder and cottonwood recruits, are early in the

seral process and still capturing sediment or developing soils to support more productive systems. However, these areas on the floodplains may not become more complex, as they are likely to be scoured during peak flow events (YCWA 2013). Areas dominated by cottonwood trees with only herbaceous understories (e.g., those found on levees), are likely a sign of interrupted riparian development, and maintenance of the levees may have prevented the natural stages of the riparian community to develop.

7 5.3.4 Large Woody Material

8 LWM creates both micro- and macro-habitat heterogeneity by forming pools, back eddies 9 Instream object cover provides structure, which promotes hydraulic complexity, diversity 10 and microhabitats for juvenile salmonids, as well as escape cover from predators. The 11 extent and quality of suitable rearing habitat and cover, including SRA, generally has a 12 strong effect on juvenile salmonid production in rivers (Healey 1991 as cited in CALFED 13 and YCWA 2005). LWM also contributes to the contribution of invertebrate food 14 sources, and micro-habitat complexity for juvenile salmonids (NMFS 2007). Snorkeling 15 observations in the lower Yuba River have indicated that juvenile Chinook salmon had a 16 strong preference for near-shore habitats with LWM (JSA 1992).

17 LWM mapping was conducted from the fall 2011 through the fall of 2012 as part of 18 YCWA's FERC relicensing efforts. YCWA also conducted field surveys in the spring of 19 2013 to collect LWM data for pieces found exclusively within bankfull widths. The 20 LWM observed in study sites tended to accumulate in one of three distributions within 21 the active channel: (1) in the bands of willow (Salix sp.) shrubs near the wetted edge; (2) 22 dispersed across open cobble bars; and (3) stranded above normal high-flow indicators 23 (YCWA 2012a). Bands of woody vegetation, dominated by willow shrubs, were present 24 along the cobble bars and floodplains at various distances from the wetted channel. The 25 shrubs acted as a capture point for much of the LWM, creating tall piles of small woody 26 debris and LWM against the upstream side of the vegetation and around the base of the 27 shrubs. On open cobbles of bars in the alluvial reaches, YCWA observed LWM and 28 smaller woody debris deposited at high flow lines (Figure 5-11); this distribution 29 comprised the smallest number of LWM pieces. A great deal of LWM was observed at flood heights, either far from the wetted channel in depressions, in stands of riparian forests, or in areas with reduced floodplains. Piles accumulated on top of boulders or riprap at flood flow levels. The majority of the wood surveyed at flood flow levels was highly degraded (YCWA 2012a). Most pieces of LWM were found to be mobile (not stabilized to resist high flows) and few pieces were observed to have channel forming influences (greater than one square meter) including the capture of other woody debris (**Figure 5-12**).

8 The majority of the LWM located within bankfull areas appeared to have floated in, with 9 less LWM appearing to have fallen from the bank. The largest pieces of LWM were 10 cottonwoods that fell from erosional banks.

11 Pasternack (2012) states that because the lower Yuba River floodway is so wide that on 12 the falling limb of a flood, the LWM gets scattered over a vast area, with disproportionate 13 concentrations racked behind flow obstructions, racked throughout vegetation patches, 14 and lining the water's edge demarking peak flood stages. Pasternack (2012) further states 15 that there is ample roughness along the fringe to catch very large pieces of wood, but the 16 lower Yuba River is so wide and deep during flood conditions that LWM cannot produce 17 log jams relative to the scale of the system. Piles of LWM (Figures 5-13 and 5-14) also 18 were found to accumulate on top of boulders or rip-rap at flood flow levels 19 (YCWA 2013).

20 **5.3.5 Other Environmental Baseline Considerations**

Instream flow requirements are specified for the lower Yuba River at the Smartsville
Gage (RM 23.6), located approximately 2,000 feet downstream from Englebright Dam,
and at the Marysville Gage (RM 6.2). Downstream of the Smartsville Gage, accretions,
local inflow, and runoff contribute, on average, approximately 200 TAF per year to the
lower Yuba River (JSA 2008).



Figure 5-11. LWM and smaller woody debris deposited downstream from Englebright Dam at a high flow line in the Timbuctoo Bend study site, looking downstream on the south side of the lower Yuba River (YCWA 2013).



Figure 5-12. Example of mobile LWM downstream from Englebright Dam at a mid-channel bar looking downstream at the Hallwood study site in the lower Yuba River (YCWA 2013).



Figure 5-13. LWM accumulated downstream from Englebright Dam against the lower portion of the gravel berms that line the north side of the lower Yuba River in the Dry Creek study site at flood flow levels (YCWA 2013).



Figure 5-14. LWM and smaller woody debris accumulated downstream from Englebright Dam on rip-rap at flood flow heights in the Parks Bar study site on the lower Yuba River

8 (YCWA 2013).

1 The hydrology and fluvial geomorphology of the lower Yuba River have been altered 2 through anthropogenic influences. Construction of numerous upstream reservoirs has 3 considerably altered the hydrologic regime of the lower Yuba River. The effects of 4 water storage and subsequent releases for irrigation have been to reduce month-to-month 5 flow variations in the river and have shifted the pattern of peak and minimum flows 6 (DWR and Corps 2003). Upstream dams have reduced the magnitude of more frequently 7 occurring flood flows (i.e. 1.5 to 20 year return period floods) (cbec and McBain & Trush 8 2010). However, larger magnitude, less frequent floods still occur, and cause the lower 9 Yuba River to respond to geomorphic processes.

The two major tributaries to the lower Yuba River are Deer Creek and Dry Creek. Located about 1.2 miles downstream of Englebright Dam, Deer Creek flows into the lower Yuba River at approximately RM 22.7. A significant falls exists approximately 500 feet upstream of the mouth of Deer Creek, which is likely impassable during drier years, but steelhead have been found above the falls during wetter years with high runoff (CDFG 1991). Deer Creek flows are regulated at Lake Wildwood (CALFED and YCWA 2005).

Located about 10.3 miles downstream of Englebright Dam, Dry Creek flows into the
lower Yuba River at RM 13.6, approximately two miles upstream of Daguerre Point Dam
(JSA 2008). The flow in Dry Creek is regulated by BVID's operation of Merle Collins
Reservoir, located on Dry Creek about 8 miles upstream from its confluence with the
Yuba River.

22 **5.3.5.1 Regulatory Requirements**

Flow releases through the powerplants at Englebright Dam are subject to provisions of various permits, licenses and contracts, including water rights permits and licenses administered by the SWRCB, PG&E's FERC License for Project No. 1403, YCWA's FERC License for Project No. 2246, YCWA's 1966 Power Purchase Contract with PG&E, a 1965 contract between YCWA and CDFW concerning instream flows, and a 1966 contract between YCWA and DWR under the Davis-Grunsky Act (NMFS 2007). In 1962 and 1965, YCWA entered into agreements with CDFW to provide the following
minimum instream flows for normal water years for preserving and enhancing the fish
resources in the lower Yuba River downstream of Daguerre Point Dam:

- 4 \Box October through December 400 cfs
- 5 \Box January through June 245 cfs
- 6

 \Box July through September – 70 cfs

Minimum flows required by the agreements were subject to reductions in critical dry years. However, in no event were flows to be reduced to less than 70 cfs. YCWA's FERC license also contains these requirements. In most years, YCWA voluntarily exceeded the 1962 and 1965 agreements' minimum flow requirements. However, when these minimum flows were implemented they often produced water temperatures and habitat conditions that were well outside the optimal preferred ranges for salmonids (NMFS 2007).

On February 23, 1988, the SWRCB received a complaint filed by a coalition of fishery groups referred to as the United Groups regarding fishery protection and water rights issues on the lower Yuba River. In 1992 and 2000, the SWRCB held hearings to receive testimony and other evidence regarding fishery issues in the lower Yuba River and other issues raised in the United Groups complaint. The SWRCB held supplemental hearings in 2003.

20 On July 16, 2003, the SWRCB issued a decision (RD-1644) regarding the protection of 21 fishery resources and other issues relating to diversion and use of water from the lower 22 Yuba River. Among other requirements, RD-1644 specified new minimum flow 23 requirements and flow fluctuation criteria for the lower Yuba River. Although these 24 minimum flow requirements did not provide the level of flow protection recommended 25 by CDFW or NMFS, according to RD-1644 these flows were developed to attempt to 26 enhance habitat for adult attraction and passage, spawning, egg incubation, juvenile 27 rearing, and emigration of Chinook salmon, steelhead, and American shad in the lower 28 Yuba River (NMFS 2007).

Conflicts among fisheries resources, water supply reliability, flood concerns, and surface
 and groundwater management associated with the lower Yuba River resulted in litigation
 between environmental and water supply interests regarding RD-1644. The Yuba Accord
 was developed as an alternative to litigation over the flow requirements specified in RD 1644.

6 LOWER YUBA RIVER ACCORD

7 The Yuba Accord includes three separate but interrelated agreements that protect and 8 enhance fisheries resources in the lower Yuba River, increase local supply reliability, and 9 provide Reclamation and DWR with increased operational flexibility for protection of 10 Delta fisheries resources through the Environmental Water Account (EWA) Program, 11 and provision of supplemental dry-year water supplies to State and Federal water 12 contractors (YCWA et al. 2007). These agreements are:

13 Dever Yuba River Fisheries Agreement (Fisheries Agreement)

14 Description of the Agreements (Conjunctive Use Agreements)

15 Description Description 15 Description Descripti Description Description Description De

16 The development of the Yuba Accord was a collaborative process, which led to a 17 comprehensive settlement of 20 years of litigation over lower Yuba River instream flow 18 requirements and related issues. Stakeholders that participated in the development of the 19 Yuba Accord include NMFS, CDFW, USFWS, YCWA, SYRCL, Trout Unlimited (TU), 20 FOR, and the Bay Institute.

21 The Fisheries Agreement is the cornerstone of the Yuba Accord. The Fisheries 22 Agreement contains new instream flow requirements for the lower Yuba River, 23 developed to increase protection of the river's fisheries resources. In addition to the best available science and data, the interests of the participating State, Federal, and local 24 25 fisheries biologists, fisheries advocates, and policy representatives were considered 26 during development of the Fisheries Agreement. The Fisheries Agreement provides for 27 minimum instream flows during specified periods of the year that are higher than the 28 corresponding flow requirements of RD-1644.

Besides the new minimum instream flows, the Fisheries Agreement also contains
 provisions for a monitoring and evaluation program to oversee the success of the flow
 schedules and a funding mechanism to pay for monitoring and study activities.

4 The Yuba Accord Technical Team tasked with flow schedule development pursued a 5 variety of analytic techniques and tools, and performed numerous evaluations to develop 6 minimum flow requirements, referred to as "flow schedules" for the lower Yuba River. 7 Additionally, the development of a new Yuba Basin water availability index was required 8 to allow a more precise determination of which flow schedule to use in the lower Yuba 9 River under each of several hydrological conditions.

10 Several steps were taken to develop to the Yuba Accord flow schedules:

- 11 (1) Development of a stressor matrix for key fisheries species in the lower
 12 Yuba River
- 13 (2) Focusing on key fish species, but also considering general aquatic habitat
 14 conditions and health in the lower Yuba River
- 15 (3) Defining general fisheries goals (e.g., maintenance, recovery,
 16 enhancement, etc.)
- 17 (4) Defining specific fisheries-related goals of the new flow regime in terms of18 flow, temperature, habitat, etc.
- 19 (5) Developing a comprehensive understanding of the hydrology and range of20 variability in hydrology for the Yuba Basin
- 21 (6) Developing a comprehensive understanding of the operational constraints
 22 (regulatory, contractual, and physical) of the YRDP and lower Yuba River, as
 23 well as an understanding of the flexibilities and inflexibilities of
 24 those constraints
- 25 (7) Developing flow regimes based on specific fisheries-related goals and water
 26 availability (as defined by operational constraints and hydrologic conditions)

The Technical Team recognized that a new flow regime for the lower Yuba River wouldneed to achieve several objectives, including:

| 1 | (1) | Maximize the occurrence of "optimal" flows and minimize the occurrence of |
|----|-----|--|
| 2 | | sub-optimal flows, within the bounds of hydrologic constraints |
| 3 | (2) | Maximize occurrence of appropriate flows for Chinook salmon and steelhead |
| 4 | | immigration spawning, rearing, and emigration |
| 5 | (3) | Provide month-to-month flow sequencing in consideration of Chinook salmon |
| 6 | | and steelhead life history periodicities |
| 7 | (4) | Provide appropriate water temperatures for Chinook salmon and steelhead |
| 8 | | immigration and holding, spawning, embryo incubation, rearing and |
| 9 | | emigration |
| 10 | (5) | Promote a dynamic, resilient, and diverse fish assemblage |
| 11 | (6) | Minimize potential stressors to fish species and lifestages |
| 12 | (7) | Develop flow regimes that consider all freshwater life stages of salmonids and |
| 13 | | allocate flows accordingly |

To build a scientific basis for crafting a flow regime that would meet these objectives, the Technical Team needed a tool to prioritize impacts on and benefits to the lower Yuba River aquatic resources. To meet this need, the Technical Team undertook development of a matrix of the primary "stressors" that affect anadromous salmonids in the lower Yuba River.

19 While the Technical Team recognized the critical importance of having a dynamic and 20 resilient aquatic community, the Technical Team also realized that developing a flow 21 regime that considered the environmental and biotic requirements of each species in the 22 entire aquatic community would not only be exceedingly complex and difficult, but 23 probably also impossible, given the myriad constraints (time, operations, finite water 24 availability, water rights, conflicting requirements of aquatic species, etc.) confronting 25 the process. The Technical Team decided that, to meet its goals, efforts would be 26 focused on addressing "keystone" lower Yuba River species. The Technical Team 27 agreed that a flow regime that supported key fish species such as Central Valley steelhead 28 and Central Valley Chinook salmon would generally benefit other native fish species, 29 recreationally important fish species such as American shad and striped bass, aquatic

1 macroinvertebrates, and other aquatic and riparian resources. The Technical Team also 2 realized that, above all else, the developed flow regime would be evaluated primarily on 3 its perceived value or benefit to State and Federally listed species, namely Central Valley 4 steelhead and Central Valley spring-run Chinook salmon, and also to fall-run Chinook 5 salmon. For this reason, the lower Yuba River stressor prioritization process principally 6 considered steelhead, spring-run Chinook salmon, and fall-run Chinook salmon. Other 7 fish species considered, but ultimately not included in the stressor prioritization process, 8 were American shad, striped bass, and green sturgeon. At the time of development, green 9 sturgeon were neither listed nor proposed for listing under the Federal ESA. The primary 10 purpose of the stressor prioritization process was to provide specific input and rationale 11 for seasonal flow regime development as well as to provide overall guidance for other 12 management and potential restoration actions.

13 For the purpose of developing the lower Yuba River Anadromous Salmonid Stressor 14 Matrix – the ultimate product of the stressor prioritization process – each species' or 15 race's freshwater lifecycle was broken up into six commonly acknowledged lifestages. 16 These lifestages are: (1) adult immigration and holding; (2) spawning and egg incubation; 17 (3) post-emergent fry outmigration (referred to as young-of-year (YOY) downstream 18 movement/outmigration for steelhead); (4) fry rearing; (5) juvenile rearing; and (6) smolt 19 outmigration (referred to as yearling (+) outmigration for steelhead). Each of the 20 lifestages was then assigned a temporal component reflecting the best available 21 knowledge of the timing and duration of that lifestage in the lower Yuba River.

Potential stressors (also referred to as limiting factors) were then identified for each species' or race's lifestage. Because most potential stressors were limited to a particular geographic reach or extent in the lower Yuba River, a geographical component was assigned to each stressor. The following is a listing of all of the potential stressors considered for the purpose of Stressor Matrix development.

| □ Water Temperature | Spawning Substrate Availability |
|--|--|
| □ Flow Fluctuations | □ Angler Impacts |
| Flow Dependent Habitat Availability | Attraction Of Non-Native Chinook Salmon |
| □ Habitat Complexity and Diversity | Overlapping Habitat |
| □ Predation | Physical Passage Impacts |
| Entrainment/Diversion Impacts | □ Lake Wildwood Operations/Deer Creek |
| Physical Passage Impediments | Flow Fluctuations |
| □ Transport/Pulse Flows | □ Motor-powered Watercraft |
| | |

1 These potential stressors were not necessarily considered to be an exhaustive list of all 2 stressors, but were the major perceived stressors, based on information current at that 3 time. In addition, the list of stressors included some elements that were not necessarily 4 considered to be stressors by all Technical Team members. The stressor prioritization 5 process was intended to serve as a tool to provide context for and assistance in the 6 development of the flow schedules.

Geographic and temporal considerations then were assigned to each stressor, further defining the extent of the potential stressor's effect on each species and lifestage. The result was a stressor matrix, which provided the Technical Team with a quantitative context of the relative importance of stressors for each month. The Technical Team members utilized the Stressor Matrix results for each month to help guide flow schedule development.

The first step in developing the flow schedules was the development of an "optimal" flow schedule that was not constrained by water availability limitations. Available information such as the Stressor Matrix results (and the species and lifestage rankings, lifestage periodicities, and geographical considerations developed for the Stressor Matrix), flow-habitat relationships (i.e., WUA) for Chinook salmon and steelhead

□ Poaching

spawning, and an understanding of the lower Yuba River flow-water temperature
 relationship was utilized in this process.

The development of the "optimal" flow schedule resulted in a "high" (Schedule 1) and a "low" (Schedule 2) range of ideal flows. The development of the "high" and "low" range of ideal flows was representative of the variety of opinions among the Technical Team biologists. Through extensive discussion and collaboration, the Technical Team biologists and representatives came to a general agreement that the two flow schedules represented the range of the "optimal" flows.

9 The second step of the flow schedule development process was the development of a 10 "worst case" flow schedule for years with extremely low water availability, targeting 11 hydrologic year classes in the 5% of driest years. This flow schedule, which eventually 12 became Schedule 6, was termed the "survival" flow schedule, because the Technical 13 Team sought to develop a flow regime that would permit survival of the year's cohort 14 during very dry hydrological conditions.

15 Recognizing the year-to-year variations in lower Yuba River water availability, the 16 Technical Team developed three additional flow schedules (Schedules 3, 4, and 5) between the "optimal" flows and the "survival" flows to be used during intermediate 17 18 hydrological conditions. The step size between each successive flow schedule was 19 adjusted to be large enough to cover the ranges of water availability without excessive 20 jumps between flow schedules. The Technical Team considered utilizing more or fewer 21 than a total of six flow schedules; however, it was ultimately determined that six flow 22 schedules could adequately address nearly the entire spectrum of hydrological 23 occurrences.

Ultimately, six flow schedules, plus conference year provisions, were developed to cover the entire range of Yuba River Basin water availabilities. The flow schedules were developed to maximize fisheries benefits during wetter years, and to maintain fisheries benefits to the greatest extent possible for drier years while taking into account other key considerations such as water supply demands, flood control operations, and hydrologic constraints of the system (NMFS 2007). Conference Years are predicted to occur during the 1% driest hydrological conditions. The Yuba Accord contains provisions regarding the minimum flows, reductions in diversions for irrigation and consultations among
 representatives of interested parties and regulatory agencies that will occur during
 Conference Years.

4 The Yuba Accord flow schedules were developed between 2001 and 2004, and 5 formalized in a set of proposed agreements in 2005. In April of 2005, a statement of 6 support for the proposed Fisheries Agreement was signed by YCWA, CDFW, NMFS, 7 USFWS, SYRCL, FOR, TU, and the Bay Institute. NMFS played a vital role in the 8 development, and subsequent implementation, of the Yuba Accord.

9 In January 2006, the parties to the Proposed Yuba Accord signed the 2006 Pilot Program 10 Fisheries Agreement, which contained minimum instream flow requirements for the 11 lower Yuba River for the period of April 1, 2006 through February 28, 2007 (YCWA 12 2006). On April 5, 2006, the SWRCB issued Order WR 2006-0009, which granted 13 YCWA's petition to extend the effective date of the RD-1644 interim instream flow 14 requirements from April 21, 2006 to March 1, 2007. On April 10, 2006, the SWRCB's 15 Division of Water Rights issued WR-2006-0010-DWR, which approved YCWA's 16 petition for the 2006 Pilot Program water transfer. Due to hydrologic conditions in the 17 Delta (e.g., unbalanced conditions), YCWA was not able to transfer water to DWR for 18 use in the EWA Program in 2006. However, the 2006 Pilot Program Fisheries 19 Agreement flow schedules remained in effect through February 28, 2007 (YCWA 2006).

20 In August 2006, YCWA also filed two petitions to temporarily amend its water right 21 permits so that YCWA could implement the 2007 Pilot Program. The first petition (the 22 Extension Petition) requested a change in the effective date of the SWRCB RD-1644 23 long-term instream flow requirements from March 1, 2007 to April 1, 2008. The second 24 petition (the Transfer Petition), filed pursuant to Water Code Section 1725, requested 25 approval of the temporary changes in YCWA's water right permits that were necessary 26 for a one-year water transfer from YCWA to DWR. The SWRCB approved these 27 petitions in February 2007.

The 2006 and 2007 Pilot Programs closely followed the proposed Yuba Accord flow regimes, accounting rules, management framework and other aspects of the Yuba Accord. Additionally, implementation of the 2006 and 2007 Pilot Programs allowed realworld tests of several of the principal elements of the Yuba Accord, including the
 proposed lower Yuba River flow schedules, transfer accounting rules, and compliance
 provisions (YCWA et al. 2007).

In 2008, the SWRCB approved the water-rights petitions necessary to implement the Yuba Accord on a long-term basis. The six flow schedules for specific types of water years are based on hydrologic conditions represented by the North Yuba Index (NYI). The NYI is an indicator of the amount of water available in the North Yuba River at New Bullards Bar Reservoir that is used to achieve the flow schedules on the lower Yuba River through operations of the reservoir. The estimated frequencies of occurrence of year-type designations under the NYI are shown below.

| Flow Schedule | North Yuba Index (TAF) | Percent Occurrence (%) | Cumulative (%) |
|---------------|---------------------------|---------------------------|-------------------|
| 1 | ≥ 1,400 | 56 | 56 |
| 2 | 1,040 – 1,399 | 22 | 78 |
| 3 | 920 – 1,039 | 7 | 85 |
| 4 | 820 – 919 | 5 | 90 |
| 5 | 693 – 819 | 5 | 95 |
| 6 | 500 – 692 | 4 | 99 |
| Conference | < 500 | 1 | 100 |

In addition to the six types of water years for the flow schedules, Conference Years are
predicted to occur at a frequency of 1% or less (during the driest years). Conference
Years are defined as water years for which the NYI is less than 500 TAF.

As part of the Yuba Accord, YCWA operates the YRDP and manages lower Yuba River instream flows according to the revised instream flow requirements, and according to specific flow schedules, numbered 1 through 6 (measured at the Marysville Gage) and lettered A and B (measured at the Smartsville Gage), based on water availability (see **Table 5-1** for Schedules 1 through 6 and **Table 5-2** for Schedules A and B). The specific flow schedule that is in effect at any time is determined by the value of the NYI and the rules described in the Fisheries Agreement.

1 Table 5-1. Yuba Accord lower Yuba River minimum instream flows (cfs) for Schedules 1 2 through 6, measured at the Marysville Gage.

| Schedule ^a | Oct 1-31 | Nov 1-30 | Dec 1-31 | Jan 1-31 | Feb 1-29 | Mar 1-31 | Apr 1-15 | Apr 16-30 | May 1-15 | May 16-31 | Jun 1-15 | Jun 16-30 | Jul 1-31 | Aug 1-31 | Sep 1-30 |
|-----------------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|--------------|-------------|--------------|-------------|--------------|-------------|-------------|-------------|
| 1 | 500 | 500 | 500 | 500 | 500 | 700 | 1,000 | 1,000 | 2,000 | 2,000 | 1,500 | 1,500 | 700 | 600 | 500 |
| 2 | 500 | 500 | 500 | 500 | 500 | 700 | 700 | 800 | 1,000 | 1,000 | 800 | 500 | 500 | 500 | 500 |
| 3 | 500 | 500 | 500 | 500 | 500 | 500 | 700 | 700 | 900 | 900 | 500 | 500 | 500 | 500 | 500 |
| 4 | 400 | 500 | 500 | 500 | 500 | 500 | 600 | 900 | 900 | 600 | 400 | 400 | 400 | 400 | 400 |
| 5 | 400 | 500 | 500 | 500 | 500 | 500 | 500 | 600 | 600 | 400 | 400 | 400 | 400 | 400 | 400 |
| 6 ^{b, c} | 350 | 350 | 350 | 350 | 350 | 350 | 350 | 500 | 500 | 400 | 300 | 150 | 150 | 150 | 350 |
| | | | | | | | | | | | | | | | |

^a For the Yuba Accord Alternative (using the NYI): Schedule 1 years are years with the NYI ≥ 1,400 TAF, Schedule 2 are years with NYI 1,040 to 1,399 TAF, Schedule 3 are years with NYI 920 to 1,039 TAF, Schedule 4 are years with NYI 820 to 919 TAF, Schedule 5 are years with NYI 693 to 819 TAF, Schedule 6 are years with NYI 500 to 692 TAF, and Conference Years are years with NYI < 500 TAF.</p>

² Indicated flows represent the average flow rate at the Marysville Gage for the specified time periods listed above. Actual flows may vary from the indicated flows according to established criteria.

^c Indicated Schedule 6 flows do not include an additional 30 TAF available from groundwater substitution to be allocated according to the criteria established in the Fisheries Agreement.

3 Table 5-2. Yuba Accord lower Yuba River minimum instream flows (cfs) for Schedules A 4 and B, measured at the Smartsville Gage

| Schedule ^a | Oct 1-31 | Nov 1-30 | Dec 1-31 | Jan 1-31 | Feb 1-29 | Mar 1-31 | Apr 1-15 | Apr 16-30 | May 1-15 | May 16-31 | Jun 1-15 | Jun 16-30 | Jul 1-31 | Aug 1-31 | Sep 1-30 |
|---|-------------|-------------|-------------|-------------|-------------|-------------|-------------|--------------|-------------|--------------|-------------|--------------|-------------|-------------|-------------|
| A ^a | 700 | 700 | 700 | 700 | 700 | 700 | 700 | с | с | с | с | с | с | с | 700 |
| B ^b | 600 | 600 | 550 | 550 | 550 | 550 | 600 | с | с | с | с | с | с | с | 500 |
| ^a Schedule A flows are to be used concurrently with Schedules 1, 2, 3, and 4 at Marysville. | | | | | | | | | | | | | | | |
| ^b Schedule B flows are to be used concurrently with Schedules 5 and 6 at Marysville. | | | | | | | | | | | | | | | |
| ^c During the summer months, flow requirements at the downstream Marysville Gage always will control, and thus, Schedule A and Schedule B flows were not developed for the May through August period. Flows at the Smartsville Gage will equal or exceed flows at Marysville. | | | | | | | | | | | | | | | |

5 Implementation of the flow schedules contained in the Yuba Accord has addressed many 6 of the flow-related stressors that existed previously, and represents relatively recent 7 improvement to Environmental Baseline conditions. The NMFS (2009) Draft Recovery 8 Plan states that "For currently occupied habitats below Englebright Dam, it is unlikely 9 that habitats can be restored to pre-dam conditions, but many of the processes and 10 conditions that are necessary to support a viable independent population of spring-run 11 Chinook salmon can be improved with provision of appropriate instream flow regimes, 12 water temperatures, and habitat availability. Continued implementation of the Yuba 13 Accord is expected to address these factors and considerably improve conditions in the lower Yuba River." 14

1 The Yuba Accord had not been approved or implemented on a long-term basis at the time 2 that the 2007 NMFS BO was prepared. The 2007 NMFS BO generally treated effects 3 resulting from flow regime changes on the lower Yuba River as part of the 4 Environmental Baseline, but also discussed flow- and water temperature-related effects 5 on critical habitat as part of the Proposed Action.

6 For this BA, previous regulatory requirements, including previous instream flow 7 requirements and the Yuba Accord instream flows and associated water temperatures that 8 have been implemented since 2006, which have led to the current status of the listed 9 species in the lower Yuba River, are included in the Environmental Baseline. Page Left Blank

1

4.0 Status of Listed Species and Critical 2 Habitat

3 4.1 Physical Features and Habitat Conditions

4 4.1.1 Hydrology

5 Historically, the Yuba River supported large numbers of spring-run Chinook salmon, fall-6 run Chinook salmon, and steelhead. Extensive hydraulic mining in the late 1800s 7 resulted in the massive influx of mining sediments that filled the lower river valleys and profoundly changed the physical character of the lower Yuba River (Moir and Pasternack 8 9 2008). The resulting habitat degradation followed by the construction of a series of 10 impassable debris dams from the early to mid-1900s likely caused major reductions in 11 salmon and steelhead populations in the Yuba River Basin (Mitchell 2010). Loss of 12 access to much of their historic spawning and rearing habitat in the upper basin likely had 13 particularly severe impacts on spring-run Chinook salmon and steelhead populations, 14 which depended on the upper basin for successful summer holding and rearing 15 (Yoshiyama et al. 1998; 2001).

16 The Yuba River suffered perhaps the most significant damage from hydraulic mining of 17 any California river. Approximately 1.5 billion cubic yards of mining debris were 18 washed into the Central Valley from five rivers, with the Yuba River accounting for 40 19 percent of that total (Mount 1995). Gilbert (1917) as cited in Yoshiyama et al. (2001) 20 estimates that "...during the period 1849-1909, 684 million cubic yards of gravel and 21 debris due to hydraulic mining were washed into the Yuba River system – more than 22 triple the volume of earth excavated during the construction of the Panama Canal", and 23 Beak Consultants, Inc. (1989) states "The debris plain ranged from about 700 feet wide 24 and up to 150 feet thick near the edge of the foothills to nearly 3 miles wide and 26 feet 25 tall near Marysville" (Beak Consultants, Inc. 1989). In addition to eliminating much of 26 the riparian vegetation corridor along the lower Yuba River (NMFS 2005b), the hydraulic 27 mining debris probably had devastating impacts on salmonids because the sediments in

1 these debris would have suffocated incubating eggs and pre-emergent fry (NMFS 2001). 2 Even by the 1870s and 1880s, the Yuba River salmon runs had been greatly diminished 3 by hydraulic mining debris effects (Yoshiyama et al. 2001). In addition, because mercury 4 was used to extract gold from mining debris, mercury exists in the Yuba River system, 5 and this mercury can be extremely toxic to salmonids (NMFS 2001). Cyanide also was 6 used in hard-rock mining to recover gold from the finely ground ore (Sumner and Smith 7 1940). Along the South Fork of the Yuba River, it was reported that "An occasional 8 heavy dose of the cyanide would kill of fish and their food, even though a stream might 9 otherwise remain unpolluted." (Sumner and Smith 1939).

10 The hydrology of the Yuba River has been altered by a series of reservoirs and water 11 conveyance facilities that are operated for water supply, hydropower production, and 12 flood control (Mitchell 2010). Three projects export significant amounts of water from 13 the Yuba River watershed. South Feather Water and Power Agency (formerly Oroville-14 Wyandotte Irrigation District) diverts water from Slate Creek (a tributary to the North 15 Yuba River) to the South Fork Feather River via its South Feather Power Project. 16 PG&E's South Yuba Canal diverts water from the South Yuba River, some of which is 17 consumptively used by the Nevada Irrigation District (NID) and some of which is 18 released into the Bear River watershed. These diversions also support NID's Yuba-Bear 19 Hydroelectric Project. PG&E's Drum-Spaulding Project diverts water from the South 20 Yuba watershed, via the Drum Canal, to the Drum Forebay. If that water is used at 21 PG&E's Drum Powerhouse, it is released to the Bear River watershed. If the water is not 22 used there, it is released to Canyon Creek (a tributary of the north fork of the North Fork 23 American River), where it is eventually used for consumptive purposes by Placer County 24 Water Agency and other entities.

The amount of water that these projects collectively export from the Yuba River watershed ranges between 589,000 acre-feet (17.3 percent of unimpaired runoff in wet years) and 267,000 acre-feet (31.1 percent of unimpaired runoff) in critical years¹ (SWRI et al. 2000). The impairment of the runoff in the lower Yuba River resulting from these

¹ Water year types are defined by the Yuba River Index of SWRCB Decision 1644.

diversions is particularly high during the April through September period during
snowmelt runoff, reaching an average of 43.2 percent of the runoff in critical years and
an estimated 50.7 percent during hydrologic conditions like those that occurred in 1931
(SWRI et al. 2000).

Located upstream of the Action Area, New Bullards Bar Reservoir was constructed by 5 6 YCWA on the North Yuba River in the late 1960s, and is the largest water storage 7 reservoir in the watershed. This reservoir is operated for flood control, power generation, 8 irrigation, recreation, and protection and enhancement of fish and wildlife. Since 1970, 9 operation of New Bullards Bar Reservoir has modified the seasonal distribution of flows 10 in the lower Yuba River by reducing spring flows and increasing summer and fall flows. 11 However, the Yuba River below Englebright Dam still experiences a dynamic flood 12 regime because of frequent uncontrolled winter and spring flows (Moir and 13 Pasternack 2008).

14 Although not part of the Action Area for this ESA consultation, New Bullards Bar 15 Reservoir operations are discussed below in recognition that water released from New 16 Bullards Bar Reservoir flows into Englebright Reservoir and water is then released into 17 the lower Yuba River. The magnitude and timing of water releases controlled by 18 YCWA's operation of New Bullards Bar Reservoir influence flow and water temperature 19 conditions in the lower Yuba River.

20 Operations of New Bullards Bar Reservoir can be described in terms of: (1) water 21 management operations (i.e., baseflow operations); (2) storm runoff operations; and (3) 22 flood control operations (NMFS 2009). Baseflow operations describe normal reservoir 23 operations when system flows are controlled through storage regulation. These 24 operations occur outside periods of flood control operations, spilling, bypassing 25 uncontrolled flows into Englebright Reservoir, and outside periods of high unregulated 26 inflows from tributary streams downstream from Englebright Dam (NMFS 2009). Flood 27 control space in New Bullards Bar Reservoir is addressed through a Water Management 28 Group, which was developed by YCWA. During flood control operations, the seasonal 29 flood pool specified in the Corps flood operation manual for New Bullards Bar Reservoir 30 is kept evacuated for flood protection, and to avoid unnecessary flood control releases.

1 Storm runoff operations occur during the storm season (typically between October and 2 May), but reservoir releases may be required to maintain flood control space between 3 September 15 and June 1 (YCWA et al. 2007). The Corps does not regulate the 4 operations of New Bullards Bar Reservoir and Englebright Dam and Reservoir, which 5 influence flow and water temperature conditions downstream in the lower Yuba River.

6 Water from Englebright Dam is released through either the Narrows I Powerhouse or the 7 Narrows II Powerhouse or, if Englebright Reservoir is full, over the top of the dam 8 (FERC 1992). Controlled releases are made through the Narrows I and Narrows II 9 powerhouses at total rates of up to about 4,200 cfs; above that rate, releases are made 10 over the spillway at the top of Englebright Dam and are essentially uncontrolled (JSA 2008). Englebright Dam has no low-level outlet.

12 Narrows I Powerhouse, owned by PG&E, is a 12 MW FERC-licensed facility, with a 13 discharge capacity of approximately 730 cfs and a bypass flow capacity (when the 14 generator is not operating) of 540 cfs. Narrows II, which is part of YCWA's YRDP, is a 15 50 MW FERC-licensed facility, with a discharge capacity of approximately 3,400 cfs and 16 a bypass flow capacity of 3,000 cfs. Annual maintenance requires the Narrows II 17 Powerhouse to be shut down for a two- to three-week period, or longer if major 18 maintenance is performed. Maintenance is typically scheduled for mid-September each 19 year. Outflows from Englebright Reservoir pass through either the Narrows II full-flow 20 bypass or through Narrows I during Narrows II maintenance activities.

YCWA and PG&E coordinate the operations of Narrows I and II for hydropower
efficiency and to maintain relatively stable flows in the lower Yuba River. The Narrows
I Powerhouse typically is used for low-flow reservoir releases (less than 730 cfs), or to
supplement the Narrows II Powerhouse capacity during high flow reservoir releases
(JSA 2008).

26 4.1.1.1 PG&E Narrows I

PG&E built the Narrows I Powerhouse in the 1940s (NMFS 2005a). Several times
during the 1950s, PG&E drew water from storage in Englebright Reservoir to generate
power at the Narrows I Powerhouse during October, when adult Chinook salmon were

1 returning to the Yuba River to spawn (Wooster and Wickwire 1970). PG&E's releases 2 attracted adult Chinook salmon in the lower Yuba River, but most of them were stranded, 3 and subsequently died when PG&E reduced its releases, and there was very little water 4 left in the lower Yuba River (Wooster and Wickwire 1970). In 1960, several parties, 5 including PG&E and CDFW, reached an agreement to prevent similar fish losses in 6 future years. Under that agreement, CDFW agreed to install a temporary barrier across the lower Yuba River's mouth before September 7th to prevent Chinook salmon from 7 8 entering the Yuba River "until October 15, when adequate transportation and spawning 9 flows are provided" (Wooster and Wickwire 1970). While this measure may have helped 10 protect fall-run Chinook salmon, it would not have provided protection for spring-run 11 Chinook salmon, because these fish would have entered the river long before September 7th, and would therefore have been exposed to all of the adverse conditions that occurred 12 13 in the river during the late summer and fall (NMFS 2005a). These practices were halted 14 following the construction of New Bullards Bar Dam and Reservoir, because the new 15 reservoir provided enough water storage to ensure adequate fall flows during most years 16 (NMFS 2005a).

As previously discussed, the Corps does not regulate or control water rights or releases.
Although the Corps does coordinate with PG&E, the Corps does not have the authority to
require Narrows I operations-related changes, nor does the Corps control water
operations in the upper Yuba River Basin or inflows into Englebright Reservoir.

21 4.1.1.2 YCWA Narrows II

The Narrows II Powerhouse, located about 400 feet downstream of Englebright Dam, was constructed in 1970 as part of the Yuba Project (FERC No. 2246). Narrows II includes one power tunnel and penstock, and one powerhouse. The penstock has a maximum capacity of 3,400 cfs.

YCWA's maintenance activities at Narrows II include generator brush replacement, which requires a 6-hour shut down 2 to 3 times per year, and annual maintenance, which typically requires a 2 to 3 week shut down, but may be longer if major maintenance is needed (NMFS 2005a). During annual maintenance prior to 2006, the 650 cfs Narrows II bypass valve usually could not be opened, and Narrows I was used to maintain instream flows in the lower Yuba River. Consequently, in the absence of water spilling over the top of Englebright Dam, flows in the lower Yuba River were reduced to a maximum of 650 cfs for several days to several weeks, depending on the type of maintenance (NMFS 2005a). YCWA schedules annual maintenance activities at Narrows II from late August to mid-September.

6 FLOW FLUCTUATIONS AND POWERHOUSE SHUTDOWNS

In addition to regularly scheduled maintenance outages, low-flow shutdowns (outages) at
the Narrows II Powerhouse used to occur when streamflows in the lower Yuba River
were below 650 cfs. During such times, YCWA's and PG&E's coordinated operation of
Narrows I and Narrows II Powerhouses resulted in releases to the lower Yuba River
being made exclusively by the Narrows I Powerhouse (NMFS 2005a).

12 Short-term emergency outages at the Narrows II Powerhouse typically resulted from 13 electrical transmission line faults (e.g., birds, trees, lightning strikes, storms) or plant 14 malfunctions. Depending on the cause of the outage, the Narrows II Powerhouse release 15 could be reduced to somewhere between 0 and 650 cfs (the capacity of the Narrows II 16 Powerhouse bypass) for a period of minutes to one or more hours. In the past, the 17 frequency of these types of outages ranged from none to several in a year, with an annual 18 average of about two per year.

19 In 2006, YCWA constructed a full-flow bypass on the Narrows II Powerhouse, which 20 allows approximately 3,000 cfs (or 88%), of the 3,400 cfs capacity of the powerhouse to 21 be bypassed around the power generation facilities to maintain river flows during 22 emergencies, maintenance, and accidental shut-downs of the powerhouse (NMFS 2007). 23 This bypass minimizes the possibility that emergencies or other events requiring that the 24 Narrows II Powerhouse be taken offline will cause significant flow fluctuations in the 25 lower Yuba River, and thereby minimizes the possibility that such fluctuations will strand 26 juvenile spring-run Chinook salmon and steelhead, or dewater redds of those species 27 (NMFS 2005a).

Before this bypass was completed, flow reductions resulting from emergency and accidental shutdowns of the Narrows II Powerhouse were a major concern due to adverse flow and water temperature effects on listed spring-run Chinook salmon and steelhead. 1 The ability to manage releases during maintenance and emergency operations was limited 2 by the design of Englebright Dam and the bypass capability of the Narrows II 3 Powerhouse which was previously only able to bypass 650 cfs (or approximately 20%) of 4 the 3,400 cfs capacity of the powerhouse. In the past, uncontrolled flow reductions due 5 to unexpected outages at Narrows II adversely affected spawning redds and fry/juvenile 6 rearing areas (FERC 2001). However, with the completion of the full-flow bypass in 7 2006, adverse effects to listed species due to emergencies, maintenance, and accidental 8 shut-downs of the powerhouse have been virtually eliminated.

9 4.1.2 Fluvial Geomorphology

10 According to Pasternack (2010), no known records of conditions prior to placer gold 11 mining in the mid-nineteenth century are available that describe the hydrologic 12 conditions in the river reach of the canyon where Englebright Dam and Reservoir are 13 located. During the era of placer gold mining, Malay Camp on the northern bank of the 14 lower Yuba River near the confluence of Deer Creek served as a base of operations for 15 miners working Landers Bar, an alluvial deposit in the nearby canyon. The historical 16 records of the existence of this camp and placer-mining site proves that coarse sediment 17 was stored in the canyon prior to hydraulic mining in a large enough quantity to produce 18 emergent alluvial bars (Pasternack 2010).

19 During the period of hydraulic gold mining, vast quantities of sand, gravel, and cobble 20 entered the Yuba River (Gilbert 1917 as cited in Yoshiyama et al. 2001) and deposited 21 throughout the system. This human impact completely transformed the river. Historical 22 photos from 1909 and 1937 document that the canyon was filled with alluvial sediment 23 with an assemblage of river features including riffles (Pasternack et al. 2010). Conditions 24 downstream of the canyon during that period were described by James et al. (2009). 25 Even though Daguerre Point Dam was built on the valley floor to prevent the transport of 26 hydraulic mining debris in 1906, it is too small to block sediment migration during floods 27 (Pasternack 2010).

Following the construction of Englebright Dam, historic photographs show that the amount of alluvium in the entire lower Yuba River, including the canyon, decreased (Pasternack et al. 2010). At the Marysville gaging station, the river incised about 20 feet from 1905-1979, while 0.5 miles downstream of the Highway 20 Bridge it incised about 35 feet over the same period (Beak Consultants, Inc., 1989). Landform adjustments continue to occur - as illustrated by Pasternack (2008), who estimated that about 605,000 yds³ of sediment (primarily gravel and cobble) were exported out of Timbuctoo Bend from 1999 to 2006. Further investigations of landform and sediment-storage changes are on-going.

8 The reported changes conform with the expected, natural response of a river to blockage 9 of downstream sediment passage (e.g. Williams and Wolman 1984). For most rivers, 10 such geomorphic changes represent a harmful human impact on a river, but here, where 11 there is a pre-existing, unnatural condition of the river corridor influenced by mining 12 debris, the dam is actually contributing to the restoration of the river toward its historical 13 geomorphic condition, in the truest meaning of the term – going back to the pre-existing 14 state prior to hydraulic gold mining (Pasternack 2010).

Despite evidence that Timbuctoo Bend is undergoing significant sediment export and river-corridor incision, White et al. (2010) reported that eight riffles persisted in the same locations over the last 26 years, and possibly longer. Most of these persistent riffles are positioned in the locally wide areas in the valley, while intervening pools are located at valley constrictions. Thus, incision and sediment export do not necessarily translate into harmful degradation of fluvial landforms.

21 The lower Yuba River has been subjected to harmful in-channel human activities that 22 further altered it. The greatest impact came from dredgers processing and re-processing 23 most of the alluvium in the river valley in the search for residual gold and to control the 24 river (James et al. 2009). First, there was the formation of the approximately 10,000-acre 25 Yuba Goldfields in the ancestral migration belt. Subsequently, there was the relocation 26 of the river to the Yuba Goldfield's northern edge and its isolation from most of the 27 Goldfields by large "gravel berms" of piled-up dredger spoils. Dredger-spoil gravel 28 berms also exist further upstream in Timbuctoo Bend away from the Yuba Goldfields; 29 these berms provide no flood-control benefit (Pasternack 2010).

1 Although no gravel berms exist in the canyon downstream of Englebright Dam, 2 mechanized gold mining facilitated by bulldozers, beginning in about 1960, completely 3 reworked the alluvial deposits in the vicinity of the confluence with Deer Creek, 4 changing the lower Yuba River geomorphology (Pasternack et al. 2010). Prior to 5 mechanized mining, glide-riffle transitions were gradual, enabling fish to select among a 6 diverse range of local hydraulic conditions. Bulldozer debris constricted the channel significantly, induced abrupt hydraulic transitioning, and caused the main riffle at the 7 8 apex of the bar to degrade into a chute. In addition, mining operations evacuated the 9 majority of alluvium at the mouth of Deer Creek, and the 1997 flood caused angular 10 hillside rocks and "shot rock" debris from the canyon bottom to be deposited on top of 11 the hydraulic-mining alluvium in the canyon (Pasternack 2010).

12 Physical habitat conditions related to salmonids downstream of Englebright Dam have 13 been studied over the years. With respect to the spawning lifestage, Fulton (2008) 14 investigated salmon spawning habitat conditions in the canyon below Englebright Dam 15 and found the conditions to be very poor to nonexistent. No rounded river 16 gravels/cobbles, suitable for spawning, were present in the canyon immediately 17 downstream of Englebright Dam and Sinoro Bar, which is located near the confluence 18 with Deer Creek, until a small amount (500 tons) of gravel was injected artificially by the 19 Corps in November 2007 (see Chapter 2 for additional discussion).

Farther downstream, spawning habitat does not appear to be limited by an inadequate supply of gravel in the lower Yuba River due to ample storage of mining sediments in the banks, bars, and dredger-spoil gravel berms (RMT 2013).

23 4.1.2.1 Englebright Dam Effects

Englebright Dam was not constructed for fish passage and therefore blocks access by anadromous salmonids to the historically utilized habitat located upstream above the dam. Consequently, spring-run Chinook salmon, fall-run Chinook salmon and steelhead in the lower Yuba River are restricted to the 24 miles extending from Englebright Dam to the mouth of the lower Yuba River.

1 Historically, spring-run and fall-run Chinook salmon were reproductively isolated due to 2 spatial and temporal segregation. Under historic natural conditions, spring-run Chinook 3 salmon migrated during spring high-flow conditions into the upper reaches of the Yuba 4 River watershed, held over the summer in relatively deep coldwater pools, and then 5 spawned in the late summer beginning in early to mid-September (Campbell and Moyle 6 1990). Fall-run Chinook salmon entered the lower Yuba River later in the year, were 7 generally unable to reach the upper reaches of the Yuba River watershed due to fall low-8 flow conditions, and are believed to have spawned in areas located farther downstream 9 than those used by spawning spring-run Chinook salmon (NMFS 2007).

10 The existence of Englebright Dam blocks the migration of spring-run fish, resulting in 11 some overlaps in the temporal and spatial distributions of spawning fall-run and spring-12 run Chinook salmon in the lower Yuba River. The resultant reduction in reproductive 13 isolation is believed to have resulted in interbreeding and genetic dilution of the genetics 14 of the much smaller spring-run Chinook salmon population (NMFS 2007). There is also 15 the potential, in areas heavily used by spawning fall-run Chinook salmon, for the later 16 spawning fall-run to superimpose their redds onto previously constructed spring-run 17 redds, thereby disrupting the spring-run redds and reducing the survival of eggs in those 18 redds (NMFS 2007).

Another potential adverse effect resulting from the existence of Englebright Dam is that it requires anadromous salmonids to complete their freshwater lifestages in the lower Yuba River without the benefit of (historically available) smaller tributaries, which can provide some level of refuge in the event of catastrophic events such as chemical spills or massive flood events (NMFS 2007). Major catastrophic events are rare, but have the potential to occur in any given year.

Nonetheless, because of the loss of historical spawning and rearing habitat above Englebright Dam, resultant loss of reproductive isolation and subsequent hybridization with fall-run Chinook salmon, restriction of spatial structure and associated vulnerability to catastrophic events, the existence of Englebright Dam represents a very high stressor to Yuba River spring-run Chinook salmon.

1 4.2 Central Valley Spring-run Chinook Salmon ESU

2 4.2.1 ESA Listing Status

3 On September 16, 1999, NMFS listed the Central Valley ESU of spring-run Chinook 4 salmon (Oncorhynchus tshawytscha) as a "threatened" species (64 FR 50394). On June 5 14, 2004, following a five-year species status review, NMFS proposed that the Central 6 Valley spring-run Chinook salmon remain listed as a threatened species based on the 7 Biological Review Team strong majority opinion that the Central Valley spring-run 8 Chinook ESU is "likely to become endangered within the foreseeable future" due to the 9 greatly reduced distribution of Central Valley spring-run Chinook salmon and hatchery 10 influences on the natural population. On June 28, 2005, NMFS reaffirmed the threatened 11 status of the Central Valley spring-run Chinook salmon ESU, and included the FRFH 12 spring-run Chinook salmon population as part of the Central Valley spring-run Chinook 13 salmon ESU (70 FR 37160).

14 Section 4(c)(2) of the ESA requires that NMFS review the status of listed species under 15 its authority at least every five years and determine whether any species should be 16 removed from the list or have its listing status changed. In August 2011, NMFS 17 completed a second 5-year status review of the Central Valley spring-run Chinook 18 salmon ESU. Prior to making a determination on whether the listing status of the ESU 19 should be uplisted (i.e., threatened to endangered), downlisted, or remain unchanged, 20 NMFS considered: (1) new scientific information that has become available since the 21 2005 status review (Good et al. 2005); (2) an updated biological status summary report 22 (Williams et al. 2011) intended to determine whether or not the biological status of 23 spring-run Chinook salmon has changed since the 2005 status review was conducted 24 (referred to as the "viability report"); (3) the current threats to the species; and (4) 25 relevant ongoing and future conservation measures and programs.

Based on a review of the available information, NMFS (2011a) recommended that the
Central Valley spring-run Chinook salmon ESU remain classified as a threatened species.
NMFS' review also indicates that the biological status of the ESU has declined since the
previous status review in 2005 and, therefore, NMFS recommended that the ESU's status
be reassessed in 2 to 3 years if it does not respond positively to improvements in environmental conditions and management actions. As part of the 5-year review, NMFS also re-evaluated the status of the FRFH stock and concluded that it still should be considered part of the Central Valley spring-run Chinook salmon ESU.

5 In addition to Federal regulations, the California Endangered Species Act (CESA, Fish 6 and Game Code Sections 2050 to 2089) establishes various requirements and protections 7 regarding species listed as threatened or endangered under state law. California's Fish 8 and Game Commission is responsible for maintaining lists of threatened and endangered 9 species under CESA. Spring-run Chinook salmon in the Sacramento River Basin, 10 including the lower Yuba River, was listed as a threatened species under CESA on 11 February 2, 1999.

12 **4.2.2 Critical Habitat Designation**

13 Critical habitat was designated for the Central Valley spring-run Chinook salmon ESU on 14 September 2, 2005 (70 FR 52488), and includes stream reaches of the Feather and Yuba 15 rivers, Big Chico, Butte, Deer, Mill, Battle, Antelope, and Clear creeks, the Sacramento 16 River, and portions of the northern Delta (NMFS 2009a). On the lower Yuba River, 17 critical habitat is designated from the confluence with the Feather River upstream to 18 Englebright Dam. This critical habitat includes the stream channels in the designated stream reaches and their lateral extents, as defined by the ordinary high-water line. In 19 20 areas where the ordinary high-water line has not been defined, the lateral extent will be 21 defined by the bankfull elevation (defined as the level at which water begins to leave the 22 channel and move into the floodplain; it is reached at a discharge that generally has a 23 recurrence interval of 1 to 2 years on the annual flood series; Bain and Stevenson 1999; 24 70 FR 52488, September 2, 2005).

25 **4.2.2.1 Primary Constituent Elements**

26 In designating critical habitat, NMFS (2009a) considers the following requirements of the

- 27 species: (1) space for individual and population growth, and for normal behavior; (2)
- 28 food, water, air, light, minerals, or other nutritional or physiological requirements; (3)

1 cover or shelter; (4) sites for breeding, reproduction, or rearing offspring; and, generally, 2 (5) habitats that are protected from disturbance or are representative of the historic 3 geographical and ecological distributions of a species [see 50 CFR 424.12(b)]. In 4 addition to these factors, NMFS also focuses on the key physical and biological features 5 within the designated area that are essential to the conservation of the species and that 6 may require special management considerations or protection. Specifically, primary constituent elements (PCEs) of critical habitat are those physical and biological features 7 8 essential to the conservation of a species for which its designated or proposed critical 9 habitat is based on.

Within the range of the spring-run Chinook salmon ESU, the PCEs of the designated critical habitat include freshwater spawning sites, freshwater rearing sites, freshwater migration corridors, estuarine areas, and nearshore and offshore marine areas. The following summary descriptions of the current conditions of the freshwater PCEs for the Central Valley spring-run Chinook salmon ESU were taken from NMFS (2009a), with the exception of new or updated information regarding current habitat conditions.

16 Freshwater Spawning Habitat

17 Freshwater spawning sites are areas with appropriate water quantity, water quality and 18 substrate for successful spawning, egg incubation, and larval development. Spring-run 19 Chinook salmon have been reported to spawn in the mainstem Sacramento River between 20 Red Bluff Diversion Dam (RBDD) and Keswick Dam, although little spawning activity 21 has been reported in recent years. Spring-run Chinook salmon primarily spawn in 22 Sacramento River tributaries such as Mill, Deer, and Butte creeks. Operations of Shasta 23 and Keswick dams on the mainstem Sacramento River are confounded by the need to 24 provide water of suitable temperature for adult winter-run Chinook salmon migration, 25 holding, spawning and incubation, as well as for spring-run Chinook salmon embryo 26 incubation in the mainstem Sacramento River.

27 FRESHWATER REARING HABITAT

Freshwater rearing sites are areas with: (1) water quantity and floodplain connectivity to form and maintain physical habitat conditions and support juvenile growth and mobility;

1 (2) water quality and forage supporting juvenile development; and (3) habitat complexity 2 characterized by natural cover such as shade, submerged and overhanging LWM, log 3 jams and beaver dams, aquatic vegetation, large rocks and boulders, side channels, and 4 undercut banks. Both spawning areas and migratory corridors comprise rearing habitat 5 for juveniles, which feed and grow before and during their outmigration. Rearing habitat 6 condition is strongly affected by habitat complexity, food supply, and the presence of 7 predators of juvenile salmonids. The channelized, leveed, and rip-rapped river reaches 8 and sloughs that are common in the Sacramento River system typically have low habitat 9 complexity, relatively low production of food organisms, and offer little protection from 10 either fish or avian predators. However, some complex, productive habitats with 11 floodplains remain in the system (e.g., Sacramento River reaches with setback levees 12 (i.e., primarily located upstream of the City of Colusa)) and flood bypasses (i.e., Yolo and 13 Sutter bypasses). Juvenile lifestages of salmonids are dependent on the function of this 14 habitat for successful survival and recruitment.

15 Freshwater Migration Corridors

16 Freshwater migration corridors provide upstream passage for adults to upstream 17 spawning areas, and downstream passage of outmigrant juveniles to estuarine and marine 18 areas. Migratory corridors are downstream of the spawning areas and include the lower 19 reaches of the spawning tributaries, the mainstem of the Sacramento River and the Delta.

20 Migratory habitat condition is strongly affected by the presence of barriers, which can 21 include dams (i.e., hydropower, flood control, and irrigation flashboard dams), 22 unscreened or poorly screened diversions, degraded water quality, or behavioral 23 impediments to migration. RBDD, completed in 1964, features a series of 11 gates that, 24 when lowered, provide for gravity diversion of irrigation water from the Sacramento 25 River into the Tehama-Colusa and Corning Canals for potential delivery to the 26 Sacramento Valley National Wildlife Refuge and to approximately 140,000 acres of 27 irrigable lands along the Interstate 5 corridor between Red Bluff and Dunnigan, 28 California (Reclamation 2008b). The RBDD has been a serious impediment to upstream 29 and downstream fish migration, and a significant portion of the Sacramento River 30 spawning habitat for Chinook salmon and steelhead occurs upstream of the dam. Until recently, the RBDD created an upstream migratory barrier in the mainstem Sacramento River during its May 15 through September 15 "gates in" configuration. In response to the NMFS (2009) BO, the RBDD gates were permanently raised in September 2011 and thus, fish passage conditions have likely improved at the RBDD. The Red Bluff Fish Passage Improvement Project, which included construction of a pumping plant to allow for diversion of water from the Sacramento River without closing the RBDD gates, was completed in 2012 (Tehama-Colusa Canal Authority 2012).

8 Both the Sacramento River flow, and many juvenile spring-run Chinook salmon, enter 9 the Delta Cross Channel (when the gates are open) and Georgiana Slough, and 10 subsequently the central Delta, especially during periods of increased water export 11 pumping from the Delta. Mortality of juvenile salmon entering the central Delta is higher 12 than for those continuing downstream in the Sacramento River. This difference in 13 mortality could be caused by a combination of factors, including: the longer migration 14 route through the central Delta to the western Delta; exposure to higher water 15 temperatures; higher predation rates; exposure to seasonal agricultural diversions; water 16 quality impairments due to agricultural and municipal discharges; and a more complex 17 channel configuration that makes it more difficult for salmon to successfully migrate to 18 the western Delta and the ocean. In addition, the State and Federal pumps and associated 19 fish facilities increase mortality of juvenile spring-run Chinook salmon through various 20 means, including entrainment into the State and Federal canals, and salvage operations.

21 ESTUARINE HABITAT AREAS

The current condition of the estuarine habitat in the Delta has been substantially degraded from historic conditions. Over 90% of the fringing fresh, brackish, and salt marshes have been lost due to human activities. This loss of the fringing marshes reduces the availability of forage species and eliminates the cycling of nutrients from the marsh vegetation into the water column of the adjoining waterways.

The channels of the Delta have been modified by the raising of levees and armoring of the levee banks with riprap, which has decreased habitat complexity by reducing the incorporation of woody material and vegetative material into the nearshore area, minimizing and reducing local variations in water depth and velocities, and simplifying
 the community structure of the nearshore environment.

Heavy urbanization and industrial actions have lowered water quality and introduced
persistent contaminants to the sediments surrounding points of discharge (i.e., refineries
in Suisun and San Pablo bays, creosote factories in Stockton, etc.)

6 Delta hydraulics have been modified as a result of federal CVP and state SWP actions. 7 Within the central and southern Delta, net water movement is towards the pumping 8 facilities, altering the migratory cues for emigrating fish in these regions. Spring-run 9 Chinook salmon smolts are drawn to the central and south Delta as they outmigrate, and 10 are subjected to the indirect effects (e.g., predation, contaminants) and direct effects (e.g., 11 salvage, loss) in the Delta and the CVP and SWP fish facilities.

12 The area of salinity transition, the low salinity zone (LSZ), is an area of high 13 productivity. Historically, this zone fluctuated in its location in relation to the outflow of 14 water from the Delta and moved westwards with high Delta inflow (i.e., floods and 15 spring runoff) and eastwards with reduced summer and fall flows. This variability in the 16 salinity transition zone has been substantially reduced by the operations of the 17 CVP/SWP. The CVP/SWP long-term water diversions also have contributed to 18 reductions in the phytoplankton and zooplankton populations in the Delta, as well as to 19 alterations in nutrient cycling within the Delta ecosystem.

20 NEARSHORE COASTAL MARINE AND OFFSHORE MARINE AREAS

Spring-run Chinook salmon reside in the Pacific Ocean from one to four years. The first few months of a salmon's ocean life has been identified as the period of critical climatic influences on survival which, in turn, suggests that coastal and estuarine environments are key areas of biophysical interaction (NMFS 2009). Juvenile salmon grow rapidly as they feed in the highly productive currents along the continental shelf (Barnhart 1986).

Most climate factors affect the entire West Coast complex of salmonids. This is particularly true in their marine phase, because the California populations are believed to range fairly broadly along the coast and intermingle, and climate impacts in the ocean occur over large spatial scales (Schwing and Lindley 2009). Salmon and steelhead residing in coastal areas where upwelling is the dominant process are more sensitive to
 climate-driven changes in the strength and timing of upwelling (NMFS 2009).

3 Oceanic and climate conditions such as sea surface temperatures, air temperatures, 4 strength of upwelling, El Niño events, salinity, ocean currents, wind speed, and primary 5 and secondary productivity affect all facets of the physical, biological and chemical 6 processes in the marine environment. Some of the conditions associated with El Niño 7 events include warmer water temperatures, weak upwelling, low primary productivity 8 (which leads to decreased zooplankton biomass), decreased southward transport of 9 subarctic water, and increased sea levels (Pearcy 1997 as cited in NMFS 2009). Strong 10 upwelling is probably beneficial because it causes greater transport of smolts offshore, 11 beyond major concentrations of inshore predators (Pearcy 1997 as cited in NMFS 2009).

12 The California Current Ecosystem (CCE) is designated by NMFS as one of eight large 13 marine ecosystems within the United States Exclusive Economic Zone. The California 14 Current begins at the northern tip of Vancouver Island, Canada and ends somewhere 15 between Punta Eugenia and the tip of Baja California, Mexico (NMFS 2009). The 16 northern end of the current is dominated by strong seasonal variability in winds, 17 temperature, upwelling, plankton production and the spawning times of many fishes, 18 whereas the southern end of the current has much less seasonal variability (NMFS 2009). 19 The primary issue for the CCE is the onset and length of the upwelling season, that is 20 when upwelling begins and ends (i.e., the "spring" and "fall" transitions). The biological 21 transition date provides an estimate of when seasonal cycles of significant plankton and 22 euphausiid production are initiated (NMFS 2009).

4.2.3 Summary of Past and Ongoing Fisheries Studies on the Lower Yuba River

As stated in YCWA (2010), the Yuba River downstream of Englebright Dam is one of the more thoroughly studied rivers in the Central Valley of California. A description of existing information regarding salmonid populations in the lower Yuba River downstream of Englebright Dam is contained in Attachment 1 to YCWA (2010), which is provided in **Appendix E** of this BA. Appendix E summarizes the available literature for

1 spring-run Chinook salmon where specifically identified, Chinook salmon in general 2 where runs are not specifically identified, and O. mykiss. Much of the referenced 3 information discusses both runs of Chinook salmon and O. mykiss, and therefore is 4 presented in its entirety in Appendix E. The appendix describes available field studies 5 and data collection reports, other relevant documents, and ongoing data collection, 6 monitoring and evaluation activities including the Yuba River Accord Monitoring and 7 Evaluation Program (M&E Program) and other data collection and monitoring programs. 8 Appendix E summarily describes 21 available field studies and data collection reports, 20 9 other relevant documents (e.g., plans, policies, historical accounts and regulatory 10 compliance), 14 ongoing data collection, monitoring and evaluation activities for the 11 M&E Program, and 4 other data collection and monitoring programs.

12 **4.2.4** Historical Abundance and Distribution

13 Spring-run Chinook salmon were once the most abundant run of salmon in the Central 14 Valley (Campbell and Moyle 1990) and were found in both the Sacramento and San 15 Joaquin drainages. The Central Valley drainage as a whole is estimated to have 16 supported annual runs of spring-run Chinook salmon as large as 600,000 fish between the late 1880s and 1940s (CDFG 1998). More than 500,000 spring-run Chinook salmon 17 18 were reportedly caught in the Sacramento-San Joaquin commercial fishery in 1883 alone 19 (Yoshiyama et al. 1998). Before the construction of Friant Dam (completed in 1942), 20 nearly 50,000 adults were counted in the San Joaquin River (Fry 1961). The San Joaquin 21 populations were essentially extirpated by the 1940s, with only small remnants of the run 22 that persisted through the 1950s in the Merced River (Hallock and Van Woert 1959; 23 Yoshiyama et al. 1998).

Annual run sizes of spring-run Chinook salmon are reported in "GrandTab", a database administered by CDFW for the Central Valley that includes reported run size estimates from 1960 through 2012, although mainstem Sacramento River estimates are not available for years before 1969 (CDFW 2013). The Central Valley spring-run Chinook salmon ESU has displayed broad fluctuations in adult abundance. Estimates of spring-run Chinook salmon in the Sacramento River and its tributaries (not including the lower Yuba and Feather rivers because GrandTab does not distinguish between fall-run and
 spring-run Chinook salmon in-river spawners, and not including the FRFH) have ranged
 from 1,404 in 1993 to 25,890 in 1982.

4 The average abundance for the Sacramento River and its tributaries (excluding the lower 5 Yuba and Feather rivers – see above) was 11,646 for the period extending from 1970 6 through 1979, 14,240 for the period 1980 through 1989, 5,825 for the period 1990 7 through 1999, and 14,055 for the period 2000 through 2009. Since 1995, spring-run 8 Chinook salmon annual run size estimates have been dominated by Butte Creek returns. 9 Since carcass survey estimates have been available in Butte Creek in 2001 through 2012, 10 Butte Creek returns have averaged 10,874 fish. The estimated spring-run Chinook 11 salmon run size was 18,511 for 2012, of which Butte Creek returns (based on the carcass 12 survey) accounted for 16,140 fish (CDFW 2013).

13 Historically, spring-run Chinook salmon occurred in the headwaters of all major river 14 systems in the Central Valley where natural barriers to migration were absent, and 15 occupied the middle and upper elevation reaches (1,000 to 6,000 feet) of most streams 16 and rivers with sufficient habitat for over summering adults (Clark 1929). Excluding the 17 lower stream reaches that were used as adult migration corridors (and, to a lesser degree, 18 for juvenile rearing), it has been estimated that at least 72% of the original Chinook 19 salmon spawning and holding habitat in the Central Valley drainage is no longer 20 available due to the construction of non-passable dams (Yoshiyama et al. 2001). Adult 21 migrations to the upper reaches of the Sacramento, Feather, and Yuba rivers were 22 eliminated with the construction of major dams during the 1940s, 1950s and 1960s. 23 Naturally spawning populations of spring-run Chinook salmon have been reported to be 24 restricted to accessible reaches of the upper Sacramento River, Antelope Creek, Battle 25 Creek, Beegum Creek, Big Chico Creek, Butte Creek, Clear Creek, Deer Creek, Mill 26 Creek, Feather River, and the Yuba River (CDFG 1998).

Historically, the Yuba River watershed reportedly was one of the most productive
habitats for runs of Chinook salmon and steelhead (Yoshiyama et al. 1996). Although it
is not possible to estimate the numbers of spawning fish from historical data, CDFG

1 (1993) suggested that the Yuba River "historically supported up to 15% of the annual run

2 of fall-run Chinook salmon in the Sacramento River system" (Yoshiyama et al. 1996).

3 By the late 1800s, anadromous fish populations were experiencing significant declines, 4 primarily because of mining activities and resultant extreme sedimentation following 5 flood events (McEwan 2001; Yoshiyama et al. 2001). As an example, the flood of 1861– 6 1862 buried much of the bottomlands along the lower Yuba River under sand deposits 7 averaging two to seven feet deep (Kelley 1989). By 1876 the channel of the lower Yuba 8 River reportedly had become completely filled, and what remained of the adjoining 9 agricultural lands was covered with sand and gravel (Kelley 1989; CDFG 1993) — a 10 marked deterioration of the river as salmon habitat (Yoshiyama et al. 2001).

11 To control flooding and the downstream movement of sediment, construction of several 12 man-made instream structures on the Yuba River occurred during the early 1900s. A structure referred to as Barrier No. 1, built in 1904 and 1905, was located 1 mile below 13 14 Parks Bar Bridge near Smartsville and was destroyed by flood waters in March 1907 15 (Sumner and Smith 1939). This barrier probably hindered salmon upstream movement 16 (Sumner and Smith 1939). In 1906, the California Debris Commission, a partnership 17 between the Federal Government and the State of California, constructed Daguerre Point 18 Dam, specifically to hold back mining debris. In 1910, the Yuba River was diverted over 19 the new dam. This approximately 24-foot high dam retained the debris, but made it 20 difficult for spawning fish to migrate upstream, although salmon reportedly did surmount 21 the dam in occasional years because they were reportedly observed in large numbers in 22 the North Yuba River at Bullards Bar during the early 1920s (Yoshiyama et al. 2001). 23 Two fishways, one for low water and the other for high water, were constructed at 24 Daguerre Point Dam prior to the floods of 1927-1928 (Clark 1929), when the fish ladders were destroyed, and were not replaced until 1938, leaving a 10-year period when 25 26 upstream fish passage at Daguerre Point Dam was blocked (CDFG 1991). A fish ladder 27 was constructed at the south end of Daguerre Point Dam in 1938 and was generally 28 ineffective (CDFG 1991), but during the fall of 1938, "several salmon were reported 29 seen below the Colgate Head Dam on the North Fork of the Yuba, 35 miles above 30 Daguerre Point Dam." (Sumner and Smith 1939).

Upstream of Daguerre Point Dam, the 260-foot-high Englebright Dam was authorized in
 1935 to hold back hydraulic mining debris, and was constructed in 1941 by the California
 Debris Commission. Englebright Dam was not authorized to provide fish passage,
 therefore it has no fish ladders and blocks anadromous fish access to all areas upstream of
 the dam (Eilers 2008; PG&E 2008; DWR 2009). The dam restricts anadromous fish to
 the lower 24 miles of the Yuba River.

7 There is limited information on the historical population size of spring-run Chinook 8 salmon in the Yuba River. Historical accounts indicate that "large numbers" of Chinook 9 salmon may have been present as far upstream as Downieville on the North Fork Yuba 10 River (Yoshiyama et al. 1996). Due to their presence high in the watershed, Yoshiyama 11 et al. (1996) concluded that these fish were spring-run Chinook salmon.

For the Middle Fork Yuba River, Yoshiyama et al. (2001) concluded that direct information was lacking on historic abundance and distribution of salmon, and they conservatively considered the 10-foot falls located 1.5 miles above the mouth of the Middle Fork Yuba River was the upstream limit of salmon distribution.

16 Yoshiyama et al. (2001) report that little is known of the original distribution of salmon 17 in the South Fork Yuba River where the Chinook salmon population was severely 18 depressed and upstream access was obstructed by dams when CDFW began surveys in 19 the 1930s. Sumner and Smith (1939) stated that the "South Fork of the Yuba is not 20 considered an angling stream in its 24 miles below the mouth of Poorman Creek, where 21 slickens* (pulverized rock) from the Spanish Mine turns the river a muddy grey." They 22 also reported that in "Poorman Creek, cyanide poisoning may have done more harm than 23 the slickens... It was evident that some strong poison was entering the stream with the 24 tailings. An occasional heavy dose of cyanide would kill off fish and fish food..." 25 Yoshiyama et al. (2001) consider the cascade, with at least a 12-foot drop, located 0.5 26 mile below the juncture of Humbug Creek, which was as essentially the historical 27 upstream limit of salmon during most years of natural streamflows.

Clark (1929) reported that the salmon spawning grounds extended from the mouth of the
lower Yuba River upstream to the town of Smartsville, but that very few salmon
(evidently spring-run) went farther upstream past that point. Sumner and Smith (1940)

1 report that salmon ascended in considerable numbers up to Bullard's Bar Dam on the 2 North Fork Yuba River while it was being constructed (1921-1924). In their 1938 survey 3 of Yuba River salmon populations, Sumner and Smith (1940) stated that the height of the 4 dams in the Yuba River blocked all potential salmon and steelhead runs upstream of the 5 barriers (Sumner and Smith 1940). However, Sumner and Smith (1940) describe the 6 ladders as "a rather ineffectual fishway... That few fish have been able to use it...is 7 testified to by the almost universal belief among local residents that at present no fish 8 ever come above the dam." In addition, the fall-run Chinook salmon run was reportedly 9 destroyed at least temporarily, and many miles of streams rendered unfit for trout 10 (Sumner and Smith 1939).

In 1951, two functional fish ladders were installed by the State of California and it was
stated that "With ladders at both ends, the fish have no difficulty negotiating this barrier
at any water stage." (CDFG 1953).

14 CDFG (1991) reports that a small spring-run Chinook salmon population historically 15 occurred in the lower Yuba River but the run virtually disappeared by 1959, presumably 16 due to the effects of water diversion and hydraulic developments on the river (Fry 1961). 17 As of 1991, a remnant spring-run Chinook salmon population reportedly persisted in the 18 lower Yuba River downstream of Englebright Dam, maintained by fish produced in the 19 lower Yuba River, fish straying from the Feather River, or fish previously and 20 infrequently stocked from the FRFH (CDFG 1991).

In the 1990s, relatively small numbers of Chinook salmon that exhibit spring-run phenotypic characteristics were observed in the lower Yuba River (CDFG 1998). Although precise escapement estimates are not available, the USFWS testified at the SWRCB lower Yuba River hearing that "...a population of about 1,000 adult spring-run Chinook salmon now exists in the lower Yuba River" (San Francisco Bay RWQCB 2006 as cited in NMFS 2009).

4.2.5 General Life History and Habitat Requirements

This section presents a general overview of lifestage-specific information (e.g., adult immigration and holding, adult spawning, embryo incubation, juvenile rearing and 1 outmigration) for the Central Valley spring-run Chinook salmon ESU. Then, this section 2 specifically focuses and provides information on lifestage specific temporal and spatial 3 distributions for spring-run Chinook salmon in the lower Yuba River. Recently, the 4 RMT developed representative temporal distributions for specific spring-run Chinook 5 salmon lifestages through review of previously conducted studies, as well as recent and 6 currently ongoing data collection activities of the M&E Program (Table 4-1). The 7 resultant lifestage periodicities encompass the majority of activity for a particular 8 lifestage, and are not intended to be inclusive of every individual in the population (RMT 9 2010; RMT 2013).

Four distinct runs of Chinook salmon spawn in the Sacramento-San Joaquin River system, with each run named for the season when the majority of the run enters freshwater as adults. The primary characteristic distinguishing spring-run Chinook salmon from the other runs of Chinook salmon is that adult spring-run Chinook salmon enter their natal streams during the spring, and hold in areas downstream of spawning grounds during the summer months until their eggs fully develop and become ready for spawning.

| Lifestage | Ja | n | Fe | b | Mar | A | pr | Ma | ay | Jı | ın | Ju | 1 | Aug | Se | ep | 0 | ct | No | v | De |)C |
|----------------------------------|----|---|----|---|-----|---|----|----|----|----|----|----|---|-----|----|----|---|----|----|---|----|----|
| Spring-run Chinook Salmon | | | | | | | | | | | | | | | | | | | | | | |
| Adult Immigration and Holding | | | | | | | | | | | | | | | | | | | | | | |
| Spawning | | | | | | | | | | | | | | | | | | | | | | |
| Embryo Incubation | | | | | | | | | | | | | | | | | | | | | | |
| Fry Rearing | | | | | | | | | | | | | | | | | | | | | | |
| Juvenile Rearing | | | | | | | | | | | | | | | | | | | | | | |
| Juvenile Downstream Movement | | | | | | | | | | | | | | | | | | | | | | |
| Smolt (Yearling+) Emigration | | | | | | | | | | | | | | | | | | | | | | |

17Table 4-1. Lifestage-specific periodicities for spring-run Chinook salmon in the lower Yuba18River (Source: RMT 2013).

1 4.2.5.1 Adult Immigration and Holding

Adult spring-run Chinook salmon immigration and holding in California's Central Valley has been reported to occur from mid-February through September (CDFG 1998; Lindley et al. 2004). Spring-run Chinook salmon are known to use the Sacramento River primarily as a migratory corridor to holding and spawning areas located in upstream tributaries. For the mainstem Sacramento River, all of the potential spring-run Chinook salmon holding habitat is located upstream from the Red Bluff Diversion Dam and downstream of Keswick Dam (CDFG 1998).

9 Suitable water temperatures for adult upstream migration reportedly range between 57°F 10 and 67°F (NMFS 1997). In addition to suitable water temperatures, adequate flows are 11 required to provide migrating adults with olfactory and other cues needed to locate their 12 spawning reaches (CDFG 1998). The primary characteristic distinguishing spring-run 13 Chinook salmon from the other runs of Chinook salmon is that adult spring-run Chinook 14 salmon hold in areas downstream of spawning grounds during the summer months until 15 their eggs fully develop and become ready for spawning. NMFS (1997) states, 16 "Generally, the maximum temperature for adults holding, while eggs are maturing, is 17 about 59-60°F, but adults holding at 55-56°F have substantially better egg viability."

18 For the lower Yuba River, adult spring-run Chinook salmon immigration and holding has 19 previously been reported to primarily occur from March through October (Vogel and 20 Marine 1991; YCWA et al. 2007), with upstream migration generally peaking in May 21 (SWRI 2002). The RMT's examination of preliminary data obtained since the VAKI 22 Riverwatcher infrared and videographic sampling system has been operated (2003 -23 present) found variable temporal modalities of Chinook salmon ascending the fish 24 ladders at Daguerre Point Dam. The RMT (2013) identified the spring-run Chinook 25 immigration and holding period as extending salmon adult from April 26 through September.

Previously, it has been reported that spring-run Chinook salmon in the lower Yuba River
hold over during the summer in the deep pools and cool water downstream of the
Narrows I and Narrows II powerhouses, or further downstream in the Narrows Reach
(CDFG 1991; SWRCB 2003), where water depths can exceed 40 feet (YCWA et al.

1 2007). Congregations of adult Chinook salmon (approximately 30 to 100 fish) have been 2 observed in the outlet pool at the base of the Narrows II Powerhouse, generally during 3 late August or September when the powerhouse is shut down for maintenance. During 4 this time period, the pool becomes clear enough to see the fish (M. Tucker, NMFS, pers. 5 comm. 2003; S. Onken, YCWA, pers. comm. 2004). While it is difficult to visually 6 distinguish spring-run from fall-run Chinook salmon in this situation, the fact that these 7 fish are congregated this far up the river at this time of year indicates that some of them 8 are likely to be spring-run Chinook salmon (NMFS 2007).

9 Past characterizations of spring-run Chinook salmon distributions from available 10 literature on the lower Yuba River have provided some anecdotal references to behavioral 11 run details (such as migration timing and areas of holding and spawning), but the 12 referenced information has not provided or referenced the basis for these descriptions. 13 Spring-run Chinook salmon have been reported to migrate immediately to areas upstream 14 of the Highway 20 Bridge after entering the lower Yuba River from March through 15 October (Vogel and Marine 1991; YCWA et al. 2007), and then over-summer in deep 16 pools located downstream of the Narrows 1 and 2 powerhouses, or further downstream in 17 the Narrows Reach through the reported spawning period of September through 18 November (CDFG 1991; SWRCB 2003).

19 The RMT's (2013) examination of preliminary data obtained since the VAKI 20 Riverwatcher infrared and videographic sampling system has been operated (2003 – 21 present) found variable temporal modalities of Chinook salmon ascending the fish 22 ladders at Daguerre Point Dam. The RMT's 3-year acoustic telemetry study of adult 23 spring-run Chinook salmon tagged downstream of Daguerre Point Dam during the 24 phenotypic adult upstream migration period has provided new information to better 25 understand adult spring-run Chinook salmon temporal and spatial distributions in the 26 lower Yuba River. The results from the Vaki Riverwatcher monitoring, and particularly 27 from the acoustic telemetry study found past characterizations of temporal and spatial 28 distributions to be largely unsupported, as phenotypic adult spring-run Chinook salmon 29 were observed to exhibit a much more diverse pattern of movement, and holding 30 locations in the lower Yuba River were more expansive than has been previously 31 reported (RMT 2013).

Although some of the acoustically-tagged spring-run Chinook salmon were observed to adhere to other previously reported characterizations, observations from the telemetry study also identified that a large longitudinal extent of the lower Yuba River was occupied by the tagged phenotypic adult spring-run Chinook salmon during immigration and holding periods (**Figure 4-1**). Figure 4-1 displays all individual fish detections obtained during the RMT's mobile acoustic tracking surveys conducted from May 2009 until November 2011 (RMT 2013).

8 Also, temporal migrations to areas upstream of Daguerre Point Dam occurred over an 9 extended period of time (Figure 4-2). The tagged phenotypic adult spring-run Chinook 10 salmon in the lower Yuba River actually migrated upstream of Daguerre Point Dam from 11 May through September, and utilized a broad expanse of the lower Yuba River during the 12 summer holding period, including areas as far downstream as Simpson Lane Bridge (i.e., 13 ~RM 3.2), and as far upstream as the area just below Englebright Dam. A longitudinal 14 analysis of acoustic tag detection data indicated that distributions were non-random, and 15 that the tagged spring-run Chinook salmon were selecting locations for holding.



16

- Figure 4-1. Spatial distribution of all individual acoustically-tagged adult phenotypic spring-run Chinook salmon (SRCS) detections obtained from the mobile tracking surveys
- 19 conducted during 2009, 2010 and 2011 (Source: RMT 2013).



1

Figure 4-2. Spatial and temporal distribution of all individual acoustically-tagged adult
 phenotypic spring-run Chinook salmon detected from the mobile tracking surveys
 conducted during 2009, 2010 and 2011 in the lower Yuba River (Source: RMT 2013).

5 The area of the river between Daguerre Point Dam and the Highway 20 Bridge was 6 largely used as a migratory corridor by the tagged adult spring-run Chinook salmon 7 during all three years of the study (RMT 2013). Telemetry data in this area demonstrated 8 relatively brief periods of occupation, characterized by sequential upstream detections as 9 individually-tagged fish migrated through this area. By contrast, frequent and sustained 10 detections were observed from the Highway 20 Bridge upstream to Englebright Dam 11 (RMT 2013).

12 Examination of individual detection data indicated that tagged phenotypic adult spring-13 run Chinook salmon that moved upstream of Daguerre Point Dam had generally passed 14 through the Daguerre Point Dam fish ladders by the end of September during all three 15 years (RMT 2013). Acoustic tag detection data were used to discern tagged spring-run 16 Chinook salmon residing in holding areas during June, July and August, and shifting to spawning areas during September into early October. This observation was repeated 17 18 during all three years of the study, and in all occupied reaches. Telemetry data 19 demonstrated that the majority of tagged phenotypic adult spring-run Chinook salmon 20 that ascended the ladders at Daguerre Point Dam also continued to move farther upstream

to the Timbuctoo, Narrows, and Englebright Dam reaches during September, coincident
 with the initiation of spawning activity (RMT 2013).

3 YCWA (2013) used the RMT's 2009-2011 acoustic tagging study data to evaluate 4 movements of the individual acoustically-tagged spring-run Chinook salmon and 5 potential relationships between changes in flow. Visual examination of the time series 6 plots of daily locations of individual acoustically-tagged Chinook salmon and mean daily 7 flows at the Smartsville Gage showed highly variable behavior among individuals on a 8 daily basis within and among years. However, several general patterns of fish movement 9 in relationship to flow are apparent.

11 Abrupt upstream movement coinciding with a decrease in flow

12 Abrupt downstream movement coinciding with a decrease in flow

13 Abrupt upstream movement occurring after an increase in flow

YCWA (2013) found that most of the individual movements of acoustically-tagged spring-run Chinook salmon potentially associated with a change in Smartsville flow were abrupt upstream movements occurring concurrently with a noticeable decrease in flow. Additional notable observations included some individuals that abruptly moved upstream in the days following a reduction in flow.

19 Observed movements of individual spring-run Chinook salmon identified during 2009 20 generally occurred within the time period from about mid-May to early September, and 21 generally occurred over a period ranging from one to nine days. Most of the observed 22 movements identified during 2010 occurred during early to mid-June, with a few 23 movements occurring during August, and generally occurred over a period ranging from 24 about one to seven days. The identified movements during 2011 generally occurred 25 during late August into early September, and generally occurred over a period ranging 26 from about one to five days. Because spring-running Chinook salmon immigrated into 27 the lower Yuba River later in 2011 than during 2009 and 2010, and were not captured 28 and acoustically-tagged until July, no potential relationships between fish movement and 29 flow reductions during the spring months could be evaluated for 2011.

1 More than half (40 out of 60) of the identified movements of Chinook salmon over the 2 three years that were potentially associated with a concurrent change in flow consisted of 3 upstream movements coinciding with a large decrease in flow (measured at the 4 Smartsville Gage). Most of the identified upstream movements occurring coincident to a 5 decrease in flow occurred when flow decreased substantially during a 1 to 2 week period 6 in late August to early September and/or during a 1 to 2 week period during May or June, depending on the year. In other words, the most common potential relationship 7 8 identified between spring-run Chinook salmon movement and flow was an abrupt and 9 continued movement upstream to the upper reaches during a large reduction in mean 10 daily Smartsville flow (38 to 68% reduction in flow) occurring over about 1 to 2 weeks.

11 4.2.5.2 Adult Spawning

In the Central Valley, spawning has been reported to primarily occur from September to November, with spawning peaking in mid- September (DWR 2004c; Moyle 2002; Vogel and Marine 1991). Within the ESU, spring-run Chinook salmon spawn in accessible reaches of the upper Sacramento River, Antelope Creek, Battle Creek, Beegum Creek, Big Chico Creek, Butte Creek, Clear Creek, Deer Creek, Mill Creek, Feather River, and the Yuba River (CDFG 1998).

18 All of the potential spring-run Chinook salmon spawning habitat in the mainstem 19 Sacramento River is located upstream from the Red Bluff Diversion Dam and 20 downstream of Keswick Dam (CDFG 1998). It has been reported that in some years high 21 water temperatures would prevent spring-run Chinook salmon egg and embryo survival 22 (USFWS 1990 as cited in CDFG 1998). During years of low storage in Shasta Reservoir 23 and under low flow releases, water temperatures exceed 56°F downstream of Keswick 24 Dam during critical months for spring-run Chinook salmon spawning and egg incubation 25 (YCWA et al. 2007).

In general, Central Valley spring-run Chinook salmon have been reported to spawn at the tails of holding pools (Moyle 2002; NMFS 2007). Redd sites are apparently chosen in part by the presence of subsurface flow. Chinook salmon usually seek a mixture of gravel and small cobbles with low silt content to build their redds. Characteristics of spawning habitats that are directly related to flow include water depth and velocity. Chinook 1 salmon spawning reportedly occurs in water velocities ranging from 1.2 feet/sec to 3.5
2 feet/sec, and spawning typically occurs at water depths greater than 0.5 feet (YCWA
3 et al. 2007).

4 For the lower Yuba River, the spring-run Chinook salmon spawning period has been 5 reported to extend from September through November (CDFG 1991; YCWA et al. 2007). 6 Limited reconnaissance-level redd surveys conducted by CDFW since 2000 during late 7 August and September have detected spawning activities beginning during the first or 8 second week of September. They have not detected a bimodal distribution of spawning 9 activities (i.e., a distinct spring-run spawning period followed by a distinct fall-run 10 Chinook salmon spawning period), and instead have detected a slow build-up of 11 spawning activities starting in early September and transitioning into the main fall-run 12 spawning period.

13 The RMT's (2013) examination of the 2009, 2010 and 2011 acoustically-tagged spring-14 run Chinook salmon data revealed a consistent pattern in fish movement. In general, 15 acoustically-tagged spring-run Chinook salmon exhibited an extended holding period, 16 followed by a rapid movement into upstream areas (upper Timbuctoo Reach, Narrows 17 Reach, and Englebright Reach) during September. Then, a period encompassing 18 approximately one week was observed when fish held at one specific location, followed 19 by rapid downstream movement. The approximate one-week period appeared to be 20 indicative of spawning events, which ended by the first week in October. These 21 observations, combined with early redd detections and initial carcasses appearing in the 22 carcass surveys (see below), suggest that the spring-run Chinook salmon spawning period 23 in the lower Yuba River may be of shorter duration than previously reported, extending 24 from September 1 through mid-October (RMT 2013).

The earliest spawning (presumed to be spring-run Chinook salmon) generally occurs in the upper reaches of the highest quality spawning habitat (i.e., below the Narrows pool) and progressively moves downstream throughout the fall-run Chinook salmon spawning season (NMFS 2007). Spring-run Chinook salmon spawning in the lower Yuba River is believed to occur upstream of Daguerre Point Dam. USFWS (2007) collected data from 168 Chinook salmon redds in the lower Yuba River on September 16-17, 2002 and 1 September 23-26, 2002, considered to be spring-run Chinook salmon redds. The redds 2 were all located above Daguerre Point Dam. During the pilot redd survey conducted 3 from the fall of 2008 through spring of 2009, the RMT (2010a) report that the vast 4 majority (96%) of fresh Chinook salmon redds constructed by the first week of October 5 2008, potentially representing spring-run Chinook salmon, were observed upstream of 6 Daguerre Point Dam. Similar distributions were observed during the 2010 and 2011 redd 7 surveys, when weekly redd surveys were conducted. About 97 and 96% of the fresh 8 Chinook salmon redds constructed by the first week of October were observed upstream 9 of Daguerre Point Dam during 2009 and 2010, respectively (RMT 2013).

10 4.2.5.3 Embryo Incubation

11 The spring-run Chinook salmon embryo incubation period encompasses the time period 12 from egg deposition through hatching, as well as the additional time while alevins remain 13 in the gravel while absorbing their yolk sacs prior to emergence.

The length of time for spring-run Chinook salmon embryos to develop depends largely on water temperatures. In well-oxygenated intragravel environs where water temperatures range from about 41°F to 55.4°F embryos hatch in 40 to 60 days and remain in the gravel as alevins for another 4 to 6 weeks, usually after the yolk sac is fully absorbed (NMFS 2009). In Butte and Big Chico creeks, emergence occurs from November through January, and in the colder waters of Mill and Deer creeks, emergence typically occurs from January through as late as May (Moyle 2002).

In the lower Yuba River, the RMT (2013) concluded that spring-run Chinook salmon
embryo incubation period generally extends from September through December.

23 4.2.5.4 Juvenile Rearing and Outmigration

After emerging, Chinook salmon fry tend to seek shallow, nearshore habitat with slow water velocities and move to progressively deeper, faster water as they grow. However, fry may disperse downstream, especially if high-flow events correspond with emergence (Moyle 2002). Spring-run juveniles may emigrate as fry soon after emergence, rear in their natal streams for several months prior to emigration as young-of-the-year, or remain in their natal streams for extended periods and emigrate as yearlings. Information regarding the duration of rearing and timing of emigration of spring-run Chinook salmon
 in the Central Valley is summarized in NMFS (2009), much of which is presented herein.

Upon emergence from the gravel, juvenile spring-run Chinook salmon may reside in freshwater for 12 to 16 months, but some migrate to the ocean as young-of-the-year fish in the winter or spring months within eight months of hatching (CALFED 2000). The average size of fry migrants (approximately 40 mm between December and April in Mill, Butte and Deer creeks) reflects a prolonged emergence of fry from the gravel (Lindley et al. 2004).

9 The timing of juvenile emigration from the spawning and rearing grounds varies among 10 the tributaries of origin, and can occur during the period extending from October through 11 April (Vogel and Marine 1991). Studies in Butte Creek (Ward et al. 2003) found the 12 majority of spring-run migrants to be fry, moving downstream primarily during 13 December, January and February, and that these movements appeared to be influenced by 14 flow. Small numbers of spring-run juveniles remained in Butte Creek to rear and migrate 15 later in the spring. Some juveniles continue to rear in Butte Creek through the summer 16 and emigrate as yearlings from October to February, with peak yearling emigration occurring in November and December (CDFG 1998). Juvenile emigration patterns in 17 18 Mill and Deer creeks are very similar to patterns observed in Butte Creek, with the 19 exception that Mill and Deer creek juveniles typically exhibit a later young-of-the-year 20 migration and an earlier yearling migration (Lindley et al. 2004). In contrast, data 21 collected on the Feather River suggests that the bulk of juvenile emigration occurs during 22 November and December (Painter et al. 1977). Seesholtz et al. (2003) speculate that 23 because juvenile rearing habitat in the Low Flow Channel of the Feather River is limited, 24 juveniles may be forced to emigrate from the area early due to competition for resources.

In general, juvenile Chinook salmon have been collected by electrofishing and observed by snorkeling throughout the lower Yuba River, but with higher abundances above Daguerre Point Dam (Beak 1989; CDFG 1991; Kozlowski 2004). This may be due to larger numbers of spawners, greater amounts of more complex, high-quality cover, and lower densities of predators such as striped bass and American shad, which reportedly are restricted to areas below the dam (YCWA et al. 2007). During juvenile rearing and outmigration, salmonids prefer stream margin habitats with sufficient depths and
velocities to provide suitable cover and foraging opportunities. Juvenile Chinook salmon
reportedly utilize river channel depths ranging from 0.9 feet to 2.0 feet, and most
frequently are in water with velocities ranging from 0 feet/sec to 1.3 feet/sec (Raleigh
et al. 1986).

6 Juvenile snorkeling surveys conducted in the lower Yuba River during 2012 indicate that 7 juvenile Chinook salmon in the lower Yuba River initially prefer slower, shallower 8 habitat, and move into faster and deeper water as they grow. RMT (2013) reported that 9 the vast majority of observations of juvenile Chinook salmon in the lower Yuba River 10 occurred in water velocities and depths indicative of slackwater and slow glide 11 mesohabitats. Juvenile Chinook salmon are known to prefer slower water habitats than 12 many other members of Oncorhynchus (Quinn 2005), and have been previously reported 13 to actively seek out slow backwaters, pools, or floodplain habitat for rearing (Sommer et 14 al. 2001; Jeffres et al. 2008). The snorkeling data collected by the RMT during 2012 are 15 generally consistent with other data available for multiple rivers (Bjornn and Reiser 16 1991). Juvenile Chinook salmon in the 30-50 mm size class tended to occupy shallower 17 habitats than larger (and presumably older) individuals, which is consistent with other 18 observations of salmonids (e.g., Bjornn and Reiser 1991). Similarly, juvenile Chinook 19 salmon showed a clear preference for faster water (up to an average of about 1.8 ft/s) as 20 they grew, consistent with trends found with salmonids in other rivers (Bjornn and 21 Reiser 1991).

22 Based upon review of available information, the RMT (2010b) recently identified the 23 spring-run Chinook salmon fry rearing period as extending from mid-November through 24 March, the juvenile rearing period extending year-round, and the young-of-year (YOY) 25 emigration period extending from November through mid-July. Associated with the 26 previously described shortened duration of spring-run Chinook salmon spawning, the fry 27 rearing period is estimated to extend from mid-November through mid-February (RMT 28 2013). Updated characterization of the juvenile (YOY) emigration (i.e., downstream 29 movement) period extends from mid-November through June (RMT 2013).

In the lower Yuba River, CDFW has conducted juvenile salmonid outmigration monitoring by operating rotary screw traps (RSTs) near Hallwood Boulevard, located approximately 6 RM upstream from the city of Marysville. CDFW's RST monitoring efforts generally extended from fall (October or November) through winter, and either into spring (June) or through the summer (September) annually from 1999 to 2006. The RMT took over operation of the year-round RST effort in the fall of 2006, and continued operations through August 2009 (RMT 2013).

8 Analyses of CDFW RST data indicate that most Chinook salmon juveniles move 9 downstream past the Hallwood Boulevard location prior to May of each year. For the 5 10 years of data included in the analyses, 97.5 to 99.2% of the total numbers of juvenile 11 Chinook salmon were captured by May 1 of each year. The percentage of the total 12 juvenile Chinook salmon catch moving downstream past the Hallwood Boulevard 13 location each year ranged from 0.4 to 1.3% during May, and 0 to 1.2% during June 14 (YCWA et al. 2007). During the 2007/2008 sampling period, 95% of all juvenile 15 Chinook salmon were captured by June 2, 2008 (Campos and Massa 2010a). Analysis of 16 the fitted distribution of weekly juvenile Chinook salmon catch at the Hallwood 17 Boulevard RST site from survey year 1999 through 2008 revealed that most emigration 18 occurred from late-December through late-April in each survey year (RMT 2013). 19 Approximately 95% of the observed catch across all years based on the fitted distribution 20 occurred by April 30 (RMT 2013).

21 Overall, most (about 84%) of the juvenile Chinook salmon were captured at the 22 Hallwood Boulevard RSTs soon after emergence from November through February, with 23 relatively small numbers continuing to be captured through June. Although not 24 numerous, captures of (oversummer) holdover juvenile Chinook salmon ranging from 25 about 70 to 140 mm FL, primarily occurred from October through January with a few 26 individuals captured into March (Massa 2005; Massa and McKibbin 2005). These fish 27 likely reared in the river over the previous summer, representing an extended juvenile 28 rearing strategy characteristic of spring-run Chinook salmon. During the 2007/2008 29 sampling period, 33 Chinook salmon that met this criterion were observed at the 30 Hallwood Boulevard RST site from mid-December through January. Juvenile Chinook 31 salmon captured during the fall and early winter (October-January) larger than 70 mm are likely exhibiting an extended rearing strategy in the lower Yuba River (Campos and
 Massa 2010a).

3 For the sampling periods extending from 2001 to 2005, CDFW identified specific runs 4 based on sub-samples of lengths of all juvenile Chinook salmon captured in the RSTs by 5 using the length-at-time tables developed by Fisher (1992), as modified by S. Greene 6 (DWR 2003b). Although the veracity of utilization of the length-at-time tables for 7 determining the run type of Chinook salmon in the Yuba River has not been ascertained, 8 based on the examination of run-specific determinations, in the lower Yuba River the vast 9 majority (approximately 94%) of spring-run Chinook salmon were captured as post-10 emergent fry during November and December, with a relatively small percentage (nearly 11 6%) of individuals remaining in the lower Yuba River and captured as YOY from 12 January through March. Only 0.6% of the juvenile Chinook salmon identified as spring-13 run was captured during April, and only 0.1% during May, and none were captured during June (YCWA et al. 2007). The above summary of juvenile Chinook salmon 14 15 emigration monitoring studies in the Yuba River is most consistent with the temporal 16 trends of spring-run Chinook salmon outmigration reported for Butte and Big Chico 17 creeks (YCWA et al. 2007).

18 4.2.5.5 Smolt Emigration

19 For the Central Valley, it has been reported that while some spring-run Chinook salmon 20 emigrate from natal streams soon after emergence during the winter and early-spring 21 (NMFS 2004a), some may spend as long as 18 months in freshwater and move 22 downstream as smolts during the first high flows of the winter, which typically occur 23 from November through January (CDFG 1998; USFWS 1995). In the Sacramento River 24 drainage, spring-run Chinook salmon smolt emigration reportedly occurs from October 25 through March (CDFG 1998). In Butte Creek, some juvenile spring-run Chinook salmon 26 rear through the summer and emigrate as yearlings from October to February, with peak 27 yearling emigration occurring in November and December (CDFG 1998). In the Feather 28 River, some spring-run Chinook salmon smolts reportedly emigrate from the Feather 29 River system from October through June (B. Cavallo, DWR, pers. comm. 2004).

1 Although it has been previously suggested that spring-run Chinook salmon smolt 2 emigration generally occurs from November through June in the lower Yuba River 3 (CALFED and YCWA 2005; CDFG 1998; SWRI 2002), recent (1999-2005), CDFW 4 monitoring data indicate that the vast majority of spring-run Chinook salmon emigrate as 5 post-emergent fry during November and December. There were some captures of (over-6 summer) holdover juvenile Chinook salmon ranging from about 70 to 140 mm FL, which 7 primarily occurred from October through January with a few individuals captured into 8 March (Massa 2005; Massa and McKibbin 2005). These fish likely reared in the river 9 over the previous summer, representing an extended juvenile rearing strategy 10 characteristic of spring-run Chinook salmon. During the 2007/2008 sampling period, 33 11 Chinook salmon that met this criterion were observed at the Hallwood Boulevard RST site from mid-December through January. Juvenile Chinook salmon captured during the 12 13 fall and early winter (October-January) larger than 70 mm are likely exhibiting an 14 extended rearing strategy in the lower Yuba River (Campos and Massa 2010a).

Based upon review of available information, the RMT (2013) recently identified the
spring-run Chinook salmon smolt (yearling+) outmigration period as extending from
October through mid-May.

18 **4.2.5.6** Lifestage-Specific Water Temperature Suitabilities

19 During November 2010, the RMT prepared a technical memorandum (RMT 2010b) to 20 review the appropriateness of the water temperature regime associated with implementation of the Yuba Accord using previously available data and information, 21 22 updated in consideration of recent and ongoing monitoring activities conducted by the 23 RMT since the pilot programs were initiated in 2006. The RMT's objectives for that 24 memorandum were to review and update the lifestage periodicities of target species in the 25 lower Yuba River, identify the appropriate thermal regime for target fish species taking 26 into account individual species and lifestage water temperature requirements, identify water temperature index values, assess the probability of occurrence that those water 27 28 temperature index values would be achieved with implementation of the Yuba Accord, 29 and to evaluate whether alternative water temperature regimes are warranted.

1 Since November 2010, additional water temperature monitoring and life history 2 investigations of anadromous salmonids in the lower Yuba River have been conducted by 3 the RMT. An update to the water temperature suitability evaluation in RMT (2010) was 4 recently conducted by RMT (2013). The water temperature suitability evaluation 5 conducted for this BA incorporates additional water temperature monitoring data from 6 what was presented in RMT (2013).

Through review of previously conducted studies, as well as recent and currently ongoing
data collection activities of the M&E Program, the RMT (2013) developed the following
representative lifestage-specific periodicities and primary locations for water temperature
suitability evaluations. The locations used for water temperature evaluations correspond
to Smartsville, Daguerre Point Dam, and Marysville.

- 12 Adult Immigration and Holding (April through September) Smartsville,
 13 Daguerre Point Dam, and Marysville
- 14 D Spawning (September through mid-October) Smartsville
- 15 🗖 Embryo Incubation (September through December) Smartsville
- Id Juvenile Rearing and Outmigration (Year-round) Daguerre Point Dam and
 Marysville

18 General Smolt (Yearling+) Emigration (October through mid-May) – Daguerre Point Dam 19 and Marysville

20 Lifestage-specific water temperature index values used as evaluation guidelines for 21 spring-run Chinook salmon were developed based on the information described in 22 Attachment A to RMT (2010b), as well as additional updated information provided in 23 Bratovich et al. (2012). These documents present the results of literature reviews that 24 were conducted to: (1) interpret the literature on the effects of water temperature on the 25 various lifestages of Chinook salmon and steelhead; (2) consider the effects of short-term 26 and long-term exposure to constant or fluctuating temperatures; and (3) establish water 27 temperature index (WTI) values to be used as guidelines for evaluation. Specifically, the 28 RMT (2013) evaluation adopted the approach established by Bratovich et al. (2012) 29 which uses the lifestage and species-specific upper tolerance WTI values. These WTI

values were not meant to be significance thresholds, but instead provide a mechanism by which to compare the suitability of the water temperature regimes associated with implementation of the Yuba Accord. Spring-run Chinook salmon lifestage-specific WTI values are provided in **Table 4-2**. The lifestages and periodicities presented in Table 4-2 differ from those presented in Table 4-1 due to specific lifestages that have the same or distinct upper tolerable WTI values.

| Lifestage | Upper Tolerance WTI | Jan | | Feb | | Mar | | Apr | | Мау | | Jun | | Jul | | Aug | | Sep | | Oct | | Nov | | Dec | |
|---|---------------------------|-----|---|-----|---|-----|---|-----|---|-----|--|-----|---|-----|--|-----|--|-----|---|-----|---|-----|---|-----|---|
| Adult Migration | 68°F | | | | | | | | | | | | | | | | | | | | | | | | |
| Adult Holding | 65°F | | | | | | | | | | | | | | | | | | | | | | | | |
| Spawning | 58°F | | | | | | | | | | | | | | | | | | | | | | | | |
| Embryo Incubation | 58°F | | | | | | | | | | | | | | | | | | | | | | | | |
| Juvenile Rearing and Downstream Movement | 65°F | | — | — | — | — | ٦ | | — | | | _ | - | ٦ | | | | — | _ | - | - | — | — | — | _ |
| Smolt (Yearling+) Emigration | 68°F | | | | | | | | _ | | | | | | | | | | | - | | | | | |

7 Table 4-2. Spring-run Chinook salmon lifestage-specific upper tolerance WTI values.

8 Recent water temperature monitoring data in the lower Yuba River are available for the 9 period extending from 2006 into June 2013, during which time operations have complied 10 with the Yuba Accord. In general, the lowest water temperatures in the lower Yuba 11 River are observed during January and February, and water temperatures steadily 12 increase until mid-June or July, remain at relatively high values through September 13 and steadily decrease thereafter. The coldest water temperatures are observed upstream at 14 the Smartsville Gage, intermediate water temperatures occur at Daguerre Point Dam, and 15 the warmest temperatures are observed downstream at the Marysville Gage for most 16 months of the year. The least amount of spatial variation in water temperature is observed 17 during late fall through winter months (i.e., late November through February), when 18 water temperatures are similar at the three monitoring locations.

Figure 4-3 displays daily water temperature monitoring results from October 2006 through late June 2013 at the Smartsville, Daguerre Point Dam, and Marysville water temperature gages, superimposed with spring-run Chinook salmon lifestage-specific



1 2

2 Figure 4-3. Monitored lower Yuba River water temperatures and spring-run Chinook 3 salmon upper tolerance WTI values.

4 upper tolerance WTI values. Water temperatures at all three gages during the period 5 evaluated are always below the upper tolerance WTI values for smolt (yearling+) 6 outmigration, juvenile rearing and outmigration, and adult immigration and holding. The 7 upper tolerance spawning and embryo incubation WTI value is never exceeded at 8 Smartsville, which is the only location evaluated for spring-run Chinook salmon 9 spawning and embryo incubation.

10 **4.2.6** Limiting Factors, Threats and Stressors

11 Limiting factors and threats supporting the listing of the Central Valley spring-run 12 Chinook salmon ESU are presented in two documents. The first is titled "Factors for 13 Decline: A Supplement to the Notice of Determination for West Coast Steelhead" (NMFS 14 1996). That report concluded that all of the factors identified in section 4(a)(1) of the 15 ESA have played roles in the decline of steelhead and other salmonids, including 16 Chinook salmon. The report identifies destruction and modification of habitat, 17 overutilization of fish for commercial and recreational purposes, and natural and human-18 made factors as being the primary reasons for the declines of west coast steelhead and 19 other salmonids including Chinook salmon. The second document is a supplement to the

1 document referred to above. This document is titled "Factors Contributing to the

- 2 Decline of West Coast Chinook Salmon: An Addendum to the 1996 West Coast Steelhead
- 3 Factors for Decline Report'' (NMFS 1998a).

4 At the ESU level, more recent descriptions of limiting factors, threats and stressors are 5 provided in the CVP/SWP OCAP BA (Reclamation 2008), the CVP/SWP OCAP BO 6 (NMFS 2009a), and the Public Draft Recovery Plan for the Evolutionarily Significant 7 Units of Sacramento River Winter-run Chinook Salmon and Central Valley Spring-run 8 Chinook Salmon and the Distinct Population Segment of Central Valley Steelhead 9 (NMFS Draft Recovery Plan) (NMFS 2009). In addition to the ESU-level discussions, 10 limiting factors, threats and stressors specifically addressing spring-run Chinook salmon 11 in the lower Yuba River are discussed in the NMFS Draft Recovery Plan (NMFS 2009). 12 These documents are incorporated by reference into this BA, and brief summaries of 13 limiting factors, threats and stressors to spring-run Chinook salmon at the ESU level, and 14 in the lower Yuba River specifically, are provided below. These brief summaries provide 15 additional detail, explanation or clarification of limiting factors, threats and stressors in 16 the lower Yuba River.

17 **4.2.6.1 ESU**

18 According to the NMFS Draft Recovery Plan (NMFS 2009), threats to Central Valley 19 spring-run Chinook salmon are in three broad categories: (1) loss of historical spawning 20 habitat; (2) degradation of remaining habitat; and (3) threats to the genetic integrity of the wild spawning populations from the FRFH spring-run Chinook salmon production 21 program. As stated in the NMFS (2009), the Central Valley spring-run Chinook salmon 22 23 ESU continues to be threatened by habitat loss, degradation and modification, small 24 hydropower dams and water diversions that reduce or eliminate instream flows during 25 migration, unscreened or inadequately screened water diversions, excessively high water 26 temperatures, and predation by non-native species. The potential effects of long-term 27 climate change also may adversely affect spring-run Chinook salmon and their recovery. 28 The 2009 NMFS OCAP BO (2009a), summarized below, identified the factors that have 29 lead to the current status of the species to be habitat blockage, water development and 30 diversion dams, water conveyance and flood control, land use activities, water quality,

hatchery operations and practices, over-utilization (e.g., ocean commercial and sport
 harvest, inland sport harvest), disease and predation, environmental variation (e.g.,
 natural environmental cycles, ocean productivity, global climate change), and non-native
 invasive species.

5 <u>HABITAT BLOCKAGE</u>

6 Hydropower, flood control, and water supply dams of the CVP, SWP, and other 7 municipal and private entities have permanently blocked or hindered salmonid access to 8 historical spawning and rearing grounds. As a result of migrational barriers, spring-run 9 Chinook salmon (as well as winter-run Chinook salmon and steelhead) populations have 10 been confined to lower elevation mainstems that historically only were used by these 11 species for migration and rearing. Population abundances have declined in these streams 12 due to decreased quantity, quality, and spatial distribution of spawning and rearing 13 habitat (Lindley et al. 2009). Higher temperatures at these lower elevations during late-14 summer and fall are also a major stressor to adult and juvenile salmonids.

15 Juvenile downstream migration patterns have been altered by the presence of dams. 16 Juvenile spring-run Chinook salmon (as well as winter-run) on the mainstem Sacramento 17 River generally outmigrate earlier than they did historically because they are hatched 18 considerably farther downstream and now have less distance to travel. Therefore, smolts 19 in the Sacramento River under present conditions must rear for a longer period of time in 20 order to reach sizes comparable to those of smolts that historically reared in upstream 21 reaches above the dams. However, for several months of the year, habitat conditions in 22 the mainstem Sacramento River do not provide the necessary features for listed 23 anadromous fish species, especially for an extended period of time.

24 WATER DEVELOPMENT

The diversion and storage of natural flows by dams and diversion structures on Central Valley waterways have altered the natural hydrologic cycles on which juvenile and adult salmonids historically based their migration patterns upon (NMFS 2009a). As much as 60% of the natural historical inflow to Central Valley watersheds and the Delta has been diverted for human uses. Dams have contributed to lower flows, higher water temperatures, lower dissolved oxygen (DO) levels, and decreased recruitment of gravel
 and LWM. More uniform flows year round have resulted in diminished natural channel
 formation, altered food web processes, and slower regeneration of riparian vegetation.

4 Water diversions for irrigated agriculture, municipal and industrial use, and managed 5 wetlands exist throughout the Central Valley. Thousands of small and medium-size 6 water diversions exist along the Sacramento River, its tributaries and the Delta. Although 7 efforts have been made in recent years to screen some of these diversions, many remain 8 unscreened. Depending on the size, location, and season of operation, these unscreened 9 diversions have the potential to entrain many lifestages of aquatic species, including 10 juvenile salmonids.

11 The Anderson-Cottonwood Irrigation District (ACID) operates a diversion dam across 12 the Sacramento River about 5 miles downstream of Keswick Dam, which is one of the 13 three largest diversions on the Sacramento River. Operated from April through October, 14 the installation and removal of the diversion dam flashboards requires close coordination 15 between Reclamation and ACID. Because substantial reductions (limited to 15% in a 16 24-hour period and 2.5% in any 1 hour) in Keswick Dam releases are necessary to install 17 or remove the flashboards, the ACID diversion dam operations have the potential to 18 impact various lifestages of Chinook salmon (e.g., redd dewatering, juvenile stranding 19 and exposure to elevated water temperatures). Redd dewatering primarily affects spring-20 and fall-run Chinook salmon during October. Although flow reductions are usually of a 21 short-term duration (i.e., lasting less than 8 hours), these short-term flow reductions may 22 cause mortality through desiccation of incubating eggs and loss of stranded juveniles.

23 Located 59 miles downstream of Keswick Dam, RBDD is owned and operated by 24 Reclamation. Historically, RBDD impeded adult salmonid passage throughout its May 25 15 through September 15 "gates in" period. Although there are fish ladders at the right 26 and left banks, and a temporary ladder in the middle of the dam, they were not very 27 efficient at passing fish because it was difficult for fish to locate the entrances to the 28 ladders. Water released from RBDD flows through a small opening under each of the 11 29 gates in the dam cause turbulent flows that confused fish and keep them from finding the 30 ladders. The effects resulting from upstream migrational delays at RBDD ranged from

1 delayed but eventually successful spawning, to pre-spawn mortality and the complete loss 2 of spawning potential in that fraction of the population. The fish ladders are not designed 3 to allow a sufficient amount of flow through them to attract adult salmonids, and previous 4 studies have shown that salmon could be delayed up to 20 days in passing the dam. 5 These delays had the potential to reduce the fitness of adults that expend their energy 6 reserves fighting the flows beneath the gates, and increase the chance of pre-spawn 7 mortality. Passage delays of a few days up to a week were believed to prevent timely 8 movement of adult spring-run Chinook salmon upstream to enter the lower reaches of 9 Sacramento River tributaries (e.g., Cottonwood Creek, Cow Creek) above the RBDD, 10 which dry up or warm up during the spring. These passage delays prevented adult 11 spring-run Chinook salmon from accessing summer holding pools in the upper reaches of 12 these tributaries. As previously discussed, the RBDD gates were permanently raised in 13 September 2011 and, thus, many of the historical migration-related stressors associated 14 with this location have likely been eliminated due to the improved fish passage conditions. 15

16 Outmigrant juvenile salmonids in the Delta have been subjected to adverse environmental 17 conditions created by water export operations at the CVP and SWP facilities. 18 Specifically, juvenile salmonid survival has been reduced by: (1) water diversions from 19 the mainstem Sacramento River into the Central Delta through the Delta Cross Channel 20 (DCC); (2) upstream or reverse flows of water in the lower San Joaquin River and 21 southern Delta waterways; (3) entrainment at the CVP/SWP export facilities and 22 associated problems at Clifton Court Forebay; and (4) increased exposure to introduced, 23 non-native predators such as striped bass (Morone saxatilis), largemouth bass 24 (Micropterus salmoides), and sunfishes (Centrarchidae spp.) within the waterways of 25 the Delta.

26 WATER CONVEYANCE AND FLOOD CONTROL

More than 1,600 miles of levee construction in the Central Valley has constricted river channels, disconnected floodplains from active river channels, reduced riparian habitat, and reduced natural channel function, particularly in lower reaches of the Sacramento River and the Delta (NMFS 2009a). The development of the water conveyance system in the Delta also has resulted in the construction of armored, rip-rapped levees on more than
1,100 miles of channels and diversions to increase channel elevations and flow capacity
of the channels (Mount 1995 as cited in NMFS 2009a).

4 Levee development in the Central Valley has affected anadromous salmonid spawning 5 habitat, freshwater rearing habitat, freshwater migration corridors, and estuarine habitats. 6 Many of the levees use angular rock (riprap) to armor the banks from erosive forces. The 7 effects of channelization and rip-rapping include the alteration of river hydraulics and 8 vegetative cover along the banks as a result of changes in bank configuration and 9 structural features (Stillwater Sciences 2006 as cited in NMFS 2009a). These changes 10 affect the quantity and quality of nearshore habitat for juvenile salmonids and have been 11 thoroughly studied (USFWS 2000; Schmetterling et al. 2001 as cited in NMFS 2009a; 12 Garland et al. 2002). Simple slopes protected with rock revetment generally create 13 nearshore hydraulic conditions characterized by greater depths and faster, more 14 homogeneous water velocities than those that occur along natural banks. Higher water 15 velocities typically inhibit deposition and retention of sediment and woody debris. These 16 changes generally reduce the range of habitat conditions typically found along natural 17 shorelines, especially by eliminating the shallow, slow-velocity river margins used by 18 juvenile fish as refuge and to escape from fast currents, deep water, and predators 19 (Stillwater Sciences 2006 as cited in NMFS 2009a). In addition, the armoring and 20 revetment of stream banks tend to narrow rivers, reducing the amount of habitat per unit 21 channel length (Sweeney et al. 2004). As a result of river narrowing, benthic habitat 22 decreases and the number of macroinvertebrates (e.g., stoneflies, mayflies) per unit 23 channel length decreases, affecting salmonid food supply.

24 LWM is a functionally important component of many streams (NMFS 1996). LWM 25 influences stream morphology by affecting channel pattern, position, and geometry, as 26 well as pool formation (Keller and Swanson 1979; Bilby 1984; Robison and Beschta 27 1990). Reduction of wood in the stream channel, either from past or present activities, 28 generally reduces pool quantity and quality, alters stream shading which can affect water 29 temperature regimes and nutrient input, and can eliminate critical stream habitat needed 30 for both vertebrate and invertebrate populations. Removal of vegetation also can 31 destabilize marginally stable slopes by increasing the subsurface water load, lowering 1 root strength, and altering water flow patterns in the slope. During the 1960s and early 2 1970s, it was common practice among California fishery management agencies to 3 remove LWM thought to be a barrier to fish migration (NMFS 1996). However, it is now 4 recognized that too much LWM was removed from streams in past decades, resulting in a 5 loss of salmonid habitat. The large scale removal of LWM prior to 1980 is believed to 6 have had major, long-term adverse effects on juvenile salmonid rearing habitat in 7 northern California (NMFS 1996). Aquatic habitat areas that were subjected to the 8 removal of LWM are still limited in the recovery of salmonid stocks, and NMFS (2009) 9 expects that this limitation could persist for 50 to 100 years.

10 LAND USE ACTIVITIES

Land use activities continue to have large-scale impacts on salmonid habitat in the Central Valley. According to Lindley et al. (2009), "Degradation and simplification of freshwater and estuary habitats over a century and a half of development have changed the Central Valley Chinook salmon complex from a highly diverse collection of numerous wild populations to one dominated by fall Chinook salmon from four large hatcheries."

17 riparian forest, with bands of vegetation extending outward for 4 or 5 miles (California 18 Resources Agency 1989). Starting with the gold rush, vast riparian forests were cleared 19 for building materials, fuel, and to open land for farming along the banks of the river. The 20 clearing of the riparian forests also removed a vital source of snags and driftwood in the 21 Sacramento River Basin. The removal of in-river snags and obstructions for navigational 22 safety has further reduced the presence of LWM in the Sacramento River and the Delta 23 (see LWM discussion above). The degradation and fragmentation of riparian habitat 24 continued with extensive flood control and bank protection projects, together with the 25 conversion of the fertile riparian lands to agriculture. By 1979, riparian habitat along the 26 Sacramento River diminished to about 2% (i.e., 11,000 to 12,000 acres) of historic levels 27 (McGill and Price 1987).

Land use activities associated with road construction, urban development, logging, mining, agriculture, and recreation have significantly altered fish habitat quantity and quality through the alteration of streambank and channel morphology, alteration of

1 ambient water temperatures, degradation of water quality, elimination of spawning and 2 rearing habitat, fragmentation of available habitats, elimination of downstream 3 recruitment of LWM, and removal of riparian vegetation, resulting in increased 4 streambank erosion (Meehan 1991 as cited in NMFS 2009a). Urban stormwater and 5 agricultural runoff may be contaminated with herbicides and pesticides, petroleum 6 products, sediment, etc. Agricultural practices in the Central Valley have eliminated 7 large trees and logs and other woody debris that would otherwise be recruited into the 8 stream channel (NMFS 1998a).

9 Increased sedimentation resulting from agricultural and urban practices is one of the 10 primary causes of salmonid habitat degradation in the Central Valley (NMFS 1996). 11 Sedimentation can adversely affect salmonids during all freshwater lifestages by clogging 12 or abrading gill surfaces, adhering to eggs, hampering fry emergence (Phillips and 13 Campbell 1961 as cited in NMFS 2009a), burying eggs or alevins, scouring and filling in 14 pools and riffles, reducing primary productivity and photosynthesis activity (Cordone and 15 Kelley 1961), and affecting intergravel permeability and DO levels. Excessive 16 sedimentation over time can cause substrates to become embedded, which reduces 17 successful salmonid spawning and egg and fry survival (Waters 1995 as cited in 18 NMFS 2009a).

19 River channel dredging to enhance inland maritime trade and to provide raw material for 20 levee construction also has altered the natural hydrology and function of the Central 21 Valley rivers. Since the mid-1800s, the Corps and others have straightened and 22 artificially deepened river channels to enhance shipping commerce, consequently 23 reducing the natural river meander and the formation of pool and riffle segments. In the 24 early 1900s, the Sacramento Flood Control Project ushered in large scale Corps actions 25 for reclamation and flood control purposes along the Sacramento River and in the Delta. 26 The creation of levees and the deep shipping channels reduced the natural tendency of the 27 Sacramento River to create floodplains along its banks during seasonal inundation 28 periods (e.g., spring snow melt). The annual inundations provided necessary juvenile 29 rearing and foraging habitat that became available in conjunction with seasonal flooding 30 processes. The armored riprapped levee banks and active maintenance actions of 31 Reclamation Districts precluded the establishment of ecologically important riparian

vegetation, introduction of valuable LWM from these riparian corridors, and the
 productive intertidal mudflats characteristic of the undisturbed Delta habitat.

3 Since the 1850s, reclamation of wetlands for urban and agricultural development has 4 resulted in the cumulative loss of tidal marsh habitat downstream (79%) and upstream 5 (94%) of Chipps Island (Conomos et al. 1985; Nichols et al. 1986; Wright and Phillips 6 1988 as cited in NMFS 2009a; Monroe et al. 1992 as cited in NMFS 2009a; Goals 7 Project 1999). Little of the extensive tracts of wetland marshes that existed prior to 1850 8 along the Central Valley river systems and within the natural flood basins exist today. 9 Most wetland and marsh areas have been "reclaimed" for agricultural purposes, leaving 10 only small remnant patches of available habitat. In the Delta, juvenile salmonids are 11 exposed to increased water temperatures during the late spring and summer due to the 12 loss of riparian shading and thermal inputs from municipal, industrial, and agricultural 13 discharges. Studies by DWR on water quality in the Delta over the last 30 years show a 14 steady decline in food resources available for juvenile salmonids, as well as an increase 15 in the clarity of the water due to a reduction in phytoplankton and zooplankton. These 16 conditions are believed to have contributed to increased juvenile Chinook salmon and 17 steelhead mortality as fish move through the Delta.

18 **WATER QUALITY**

19 Over the past 150 years, the water quality of the Delta has been adversely affected by 20 increased water temperatures, decreased DO levels, and increased turbidity and 21 contaminant loads, which have degraded the quality of the aquatic habitat for the rearing 22 and migration of salmonids. Historic and ongoing point and nonpoint source discharges 23 impact surface waters, and portions of major rivers and the Delta are impaired, to some 24 degree, by discharges from agriculture, mines, urban areas and industries (California 25 RWQCB 1998). Pollutants include effluents from wastewater treatment plants and 26 chemical discharges (e.g., dioxin from San Francisco Bay petroleum refineries) (McEwan 27 and Jackson 1996). Agricultural drain water, another possible source of contaminants, 28 can contribute up to 30% of the total inflow into the Sacramento River during drier 29 conditions (Reclamation 2008a).
1 According to NMFS (2009a), the California RWQCB (1998; 2001) has identified the 2 Delta as impaired waterbody having elevated levels of chlorpyrifos, an 3 dichlorodiphenyltrichlor (i.e. DDT), diazinon, mercury, Group A pesticides (e.g., aldrin, 4 dieldrin, chlordane, endrin, heptachlor, heptachlor epoxide, hexachlorocyclohexanes 5 (including lindane), endosulfan and toxaphene), organic enrichment, as well as low DO. 6 In general, water degradation or contamination can lead to either acute toxicity, resulting 7 in death when concentrations are sufficiently elevated, or more typically, when 8 concentrations are lower, to chronic or sublethal effects that reduce the physical health of 9 the organism, and lessens its survival over an extended period of time. Mortality may 10 become a secondary effect due to compromised physiology or behavioral changes that 11 lessen the organism's ability to carry out its normal activities. For listed species, these 12 effects may occur directly to the listed fish or to its prey base, which reduces the forage 13 base available to the listed species.

In the aquatic environment, most anthropogenic chemicals and waste materials, including 14 15 toxic organic and inorganic chemicals eventually accumulate in sediment (Ingersoll 1995 16 as cited in NMFS 2009a). Direct exposure to contaminated sediments may cause 17 deleterious effects if a fish swims through a plume of the re-suspended sediments or rests 18 on contaminated substrate and absorbs the toxic compounds via dermal contact, 19 ingestion, or uptake across the gills. Although sediment contaminant levels can be 20 significantly higher than the overlying water column concentrations (EPA 1994), the 21 more likely means of exposure is through the food chain when fish feed on organisms 22 that are contaminated with toxic compounds. Prey species become contaminated either 23 by feeding on the detritus associated with the sediments or dwelling in the sediment 24 itself. Therefore, the degree of exposure to the salmonids depends on their trophic level 25 and the amount of contaminated forage base consumed. Salmonid biological responses to 26 contaminated sediments are similar to those resulting from waterborne exposures once a 27 contaminant has entered the body of the fish.

28 HATCHERY OPERATIONS AND PRACTICES

CDFW is currently operating 10 salmon and steelhead hatchery facilities in California.
Eight of these 10 facilities (i.e., Iron Gate, Trinity River, Warm Springs, Feather River,

Nimbus, Mokelumne River, and Merced River Hatcheries and the Coyote Valley Fish
 Facility) were constructed below dams on major rivers as mitigation for loss of access to
 anadromous fish habitat upstream of the dams. The Thermalito Annex, which is not
 located below a dam, supports the mitigation and enhancement programs that include
 Chinook and coho salmon for the FRFH.

6 Five hatcheries currently produce Chinook salmon in the Central Valley, and four of 7 these also produce steelhead. Releasing large numbers of hatchery fish can pose a threat 8 to wild Chinook salmon and steelhead stocks through genetic impacts, competition for 9 food and other resources between hatchery and wild fish, predation of hatchery fish on 10 wild fish, and increased fishing pressure on wild stocks as a result of hatchery production 11 (Waples 1991). The genetic impacts of artificial propagation programs in the Central 12 Valley are primarily caused by straying of hatchery fish and the subsequent interbreeding 13 of hatchery fish with wild fish. In the Central Valley, practices such as transferring eggs 14 between hatcheries and trucking smolts to distant sites for release contribute to elevated 15 straying levels (USDOI 1999, as cited in NMFS 2009a).

16 Hatchery practices as well as spatial and temporal overlaps of habitat use and spawning 17 activity between spring- and fall-run Chinook salmon have led to the hybridization and 18 homogenization of some subpopulations (CDFG 1998). As early as the 1960s, Slater 19 (1963) observed that spring-run and early fall-run were competing for spawning sites in 20 the Sacramento River below Keswick Dam, and speculated that the two runs may have 21 hybridized. Spring-run Chinook salmon from the FRFH have been documented as 22 straying throughout the Central Valley for many years (CDFG 1998), and may have 23 contributed to hybridization. In the Feather River, the lack of physical separation has led 24 to hybridization of spring- and fall-run Chinook salmon.

The relatively low number of spawners needed to sustain a hatchery population can result in high harvest-to-escapements ratios in waters where fishing regulations are set according to hatchery population. This can lead to over-exploitation and reduction in the size of wild populations existing in the same system as hatchery populations due to incidental by-catch (McEwan 2001). Hatcheries also can have some positive effects on salmonid populations. Spring-run Chinook salmon produced in the FRFH are considered part of the spring-run Chinook salmon ESU. Artificial propagation has been shown to be effective in bolstering the numbers of naturally spawning fish in the short term under specific scenarios. Artificial propagation programs can also aid in conserving genetic resources and guarding against catastrophic loss of naturally spawned populations at critically low abundance levels (IMST 2001, as cited in NMFS 2004).

8 **OVERUTILIZATION**

9 OCEAN COMMERCIAL AND SPORT HARVEST

10 Extensive ocean recreational and commercial troll fisheries for Chinook salmon exist along the Northern and Central California coast, and an inland recreational fishery exists 11 12 in the Central Valley for Chinook salmon and steelhead. The Central Valley Index (CVI) 13 is an annual index of abundance of all Central Valley Chinook salmon stocks combined, 14 and is defined as the calendar year sum of ocean fishery Chinook harvests in the area 15 south of Point Arena, California (where 85% of Central Valley Chinook salmon are 16 caught), plus the Central Valley adult Chinook spawning escapement (Lindley et al. 17 2009). Since 1991, the PFMC's Salmon Technical Team (comprised of scientists from 18 NMFS, USFWS, and state fisheries agencies from OR, WA, and CA) has used a linear 19 regression of the CVI on the previous year's Central Valley age-2 return to forecast the 20 CVI (BDCP 2009). The CVI harvest rate index is an annual index of the ocean harvest 21 rate on all Central Valley Chinook stocks combined, and is defined as the ocean harvest 22 landed south of Point Arena, California, divided by the CVI (Lindley et al. 2009).

23 There are no Pacific Coast Salmon Fisheries Management Plan (FMP) objectives in place 24 specifically regulating the harvest of spring-run Chinook salmon, except that the FMP 25 will manage ocean fisheries consistent with NMFS ESA consultation standards (BDCP 26 2009). The current FMP harvest constraints on winter-run Chinook salmon serve as 27 proxy for Central Valley spring-run Chinook salmon (BDCP 2009). Spring-run Chinook 28 salmon CVI harvest rate index ranged from 0.55 to nearly 0.80 between 1970 and 1995, 29 when harvest rates were adjusted for the protection of winter-run Chinook salmon 30 (NMFS 2003). The decline in the CVI harvest rate index to 0.27 in 2001 as a result of high fall-run Chinook salmon escapement also resulted in reductions to the authorized
 harvest of spring-run Chinook salmon (NMFS 2003).

3 FRFH spring-run Chinook salmon provide indices of harvest of natural spring-run. 4 Maturing age-3 and age-4 spring-run Chinook salmon are vulnerable to the early portion 5 of the recreational and commercial season, whereas fall-run Chinook salmon are exposed 6 to an entire harvest season (BDCP 2009). Inferences drawn from coded-wire tag 7 recoveries indicate that 44% of the spring-run Chinook salmon are taken prior to May 1, 8 the start of the commercial fishing season (BDCP 2009). Ocean fisheries have affected 9 the age structure of spring-run Chinook salmon through targeting large fish for many 10 years and reducing the numbers of 4- and 5-year-old fish (CDFG 1998). As a result of 11 very low returns to the Central Valley in 2007, there was a complete closure of the 12 commercial and recreational ocean Chinook salmon fishery in 2008 and 2009. Due to 13 improved ocean salmon numbers, a severely restricted commercial season and short 14 recreational season opened in 2010 (Bacher 2011). On April 13, 2011, the Pacific 15 Fishery Management Council (PFMC) adopted a set of ocean salmon seasons that 16 provides both recreational and commercial opportunities during the 2011 fishing season. 17 PFMC (2011) reports that "Greatly improved abundance of Sacramento River fall-run 18 Chinook salmon will fuel the first substantial ocean salmon fisheries off California and 19 Oregon since 2007. Fisheries south of Cape Falcon are supported by Sacramento River 20 fall Chinook. In 2008 and 2009, poor Sacramento returns led to the largest ocean salmon 21 fishery closure on record. The abundance forecast of Sacramento River fall Chinook in 22 2011 is 730,000, far above the number needed for optimum spawning this fall (122,000-23 180,000 fish)."

24 INLAND SPORT HARVEST

Historically in California, almost half of the river sport fishing effort has occurred in the
Sacramento-San Joaquin River system, particularly upstream from the city of Sacramento
(Emmett et al. 1991). In-river recreational fisheries historically have taken spring-run
Chinook salmon throughout the species' range. During the summer, adult spring-run
Chinook salmon are targeted by anglers when the fish congregate and hold in large pools.
Poaching also occurs at fish ladders, and other areas where adults congregate. However,

the significance of poaching on the adult population is unknown (NMFS 2009a).
 Specific regulations for the protection of spring-run Chinook salmon in Mill, Deer, Butte,
 and Big Chico creeks and the lower Yuba River have been added to the CDFW
 regulations.

5 DISEASE AND PREDATION

6 Salmonids are exposed to numerous bacterial, protozoan, viral, and parasitic organisms in 7 spawning and rearing areas, hatcheries, migratory routes, and the marine environment 8 (NMFS 1996, 1996a, 1998a), and infectious disease is one of many factors that influence 9 adult and juvenile salmonid survival. Specific diseases such as bacterial kidney disease, 10 Ceratomyxosis shasta, columnaris, furunculosis, infectious hematopoietic necrosis, 11 redmouth and black spot disease, whirling disease, and erythrocytic inclusion body 12 syndrome are known, among others, to affect Chinook salmon and steelhead (NMFS 13 1996; 1996a; 1998a). Little current or historical information exists to quantify changes in 14 infection levels and mortality rates attributable to these diseases; however, studies have 15 shown that wild fish tend to be less susceptible to pathogens than are hatchery-reared fish 16 (NMFS 2009a). Nevertheless, wild salmonids may contract diseases that are spread 17 through the water column (i.e., waterborne pathogens) as well as through interbreeding 18 with infected hatchery fish. The stress of being released into the wild from a controlled 19 hatchery environment frequently causes latent infections to convert into a more 20 pathological state, and increases the potential of transmission from hatchery reared fish to 21 wild stocks within the same waters.

22 As described in NMFS (2005a), accelerated predation is also a significant factor affecting 23 critical habitat for spring-run Chinook salmon. Although predation is a natural 24 component of spring-run Chinook salmon life ecology, the rate of predation likely has 25 greatly increased through the introduction of non-native predatory species such as striped 26 bass (Marone saxatilis) and largemouth bass (Micrapterus salmaides), and through the 27 alteration of natural flow regimes and the development of structures that attract predators, 28 including dams, bank revetment, bridges, diversions, piers, and wharfs (Stevens 1961; 29 Vogel et al. 1988 as cited in NMFS 2009; Garcia 1989 as cited in Reclamation 2008; 30 Decato 1978 as cited in Reclamation 2008). The USFWS found that more predatory fish

1 were found at rock revetment bank protection sites between Chico Landing and Red 2 Bluff than at sites with naturally eroding banks (Michny and Hampton 1984). On the 3 mainstem Sacramento River, high rates of predation are known to occur at RBDD, ACID, 4 GCID, and at south Delta water diversion structures (CDFG 1998). From October 1976 5 to November 1993, CDFW conducted ten mark/recapture experiments at the SWP's 6 Clifton Court Forebay to estimate prescreen losses using hatchery-reared juvenile 7 Chinook salmon. Pre-screen losses ranged from 69 to 99%. Predation from striped bass is thought to be the primary cause of the loss (CDFG 1998; Gingras 1997). 8

9 Predation on juvenile salmonids has increased as a result of water development activities, 10 which have created ideal habitats for predators and non-native invasive species. As 11 juvenile salmonids pass the Sacramento River system dams, fish are subject to conditions 12 that can disorient them, making them highly susceptible to predation by fish or birds. 13 Striped bass and Sacramento pikeminnow (Ptychocheilus grandis), a species native to the 14 Sacramento River Basin that co-evolved with anadromous salmonids, congregate below 15 dams and prey on juvenile salmon in the tail waters. Tucker et al. (1998) reported that: 16 (1) striped bass exhibit a strong preference for juvenile salmonids; (2) during the summer 17 months, juvenile salmonids increased to 66% of the total weight of Sacramento 18 pikeminnow stomach contents; and (3) the percent frequency of occurrence for juvenile 19 salmonids nearly equaled other fish species in the stomach contents of the predatory fish. 20 Additionally, Tucker et al. (2003) showed the temporal distribution for these two 21 predatory species in the RBDD area were directly related to RBDD operations (i.e., 22 predators congregated when the dam gates were in, and dispersed when the dam gates 23 were removed).

24 Other locations in the Central Valley where predation is of concern include flood 25 bypasses, post-release sites for salmonids salvaged at the CVP and SWP Fish Facilities, 26 and the Suisun Marsh Salinity Control Gates (SMSCG). The dominant predator species 27 at the SMSCG was striped bass, and the remains of juvenile Chinook salmon were 28 identified in their stomach contents (Edwards et al. 1996; Tillman et al. 1996; NMFS 29 1997a). Striped bass and pikeminnow predation on salmon at salvage release sites in the 30 Delta and lower Sacramento River has been documented (Orsi 1967; Pickard et al. 1982). 31 However, accurate predation rates at these sites are difficult to determine. From October 1976 to November 1993, CDFW conducted 10 mark/recapture studies at the SWP's
 Clifton Court Forebay to estimate pre-screen losses using hatchery-reared juvenile
 Chinook salmon. Pre-screen losses ranged from 69 to 99%, and predation by striped bass
 is thought to be the primary cause of the loss (Gingras 1997). More recent studies by
 DWR (2008) have verified this level of predation also exists for steelhead smolts within
 Clifton Court Forebay, indicating that these predators were efficient at removing
 salmonids over a wide range of body sizes.

8 Avian predation on fish contributes to the loss of migrating juvenile salmonids (NMFS 9 2009a). Fish-eating birds (e.g., great blue herons, black-crowned night herons, gulls, 10 osprey) in the Central Valley have high metabolic rates and require large quantities of 11 food relative to their body size. Mammals can also be an important source of predation 12 on salmonids within the California Central Valley. These animals, especially river otters, 13 are capable of removing large numbers of salmon and trout from the aquatic habitat 14 (Dolloff 1993 as cited in NMFS 2009a). Mammals have the potential to consume large numbers of salmonids, but generally scavenge post-spawned salmon. In the marine 15 16 environment, Southern Resident killer whales target Chinook salmon as their preferred 17 prey (96% of prey consumed during spring, summer and fall, from long-term study of 18 resident killer whale diet; Ford and Ellis 2006).

19 <u>Environmental Variation</u>

20 The scientific basis for understanding the processes and sources of climate variability has 21 grown significantly in recent years, and our ability to forecast human and natural 22 contributions to climate change has improved dramatically. With consensus on the 23 reality of climate change now established (Oreskes 2004; IPCC 2007), the scientific, 24 political, and public priorities are evolving toward determining its ecosystem impacts, 25 and developing strategies for adapting to those impacts. Global climate change is playing 26 an increasingly important role in scientific and policy debates related to effective water 27 management. The most considerable impacts of climate change on water resources in the 28 United States are believed to occur in the mid-latitudes of the West, where the runoff 29 cycle is largely determined by snow accumulation and subsequent melt patterns. 30 Evidence is continuing to accumulate to indicate global climate change will have a marked effect on water resources in California. Numerous peer-reviewed scientific articles on climate and water issues in California have been published to date, with many more in preparation, addressing a range of considerations from proposed improvements in the downscaling of general circulation models to understanding how reservoir operations might be adapted to new conditions (Kiparsky and Gleick 2003).

6 NMFS (2009) states that the potential effects of long-term climate change may adversely 7 affect spring-run Chinook salmon and steelhead, and the recovery of both species. 8 Current climate change information suggests that the Central Valley climate will become 9 warmer, a challenging prospect for Chinook salmon and steelhead – both of which are 10 coldwater fish at the southern end of their distribution. According to NMFS (2009a), 11 early marine survival for juvenile salmon is a critical phase in their survival and 12 The correlation between various environmental indices that development into adults. 13 track ocean conditions and salmon productivity in the Pacific Ocean, both on a broad and 14 local scale, provides an indication of how climate-related factors influence salmon 15 survival in the ocean. Consistent with the approach taken in recent NMFS BOs (NMFS 16 2011; NMFS 2010; NMFS 2010a; NMFS 2010b), the discussion below describes the 17 potential climate-related threats anticipated to affect the status of listed species, including 18 inter-annual climatic variations (e.g. El Niño and La Niña), the Wells Ocean Productivity 19 Index, and longer term cycles in ocean conditions pertinent to salmonid survival (e.g., 20 Pacific Decadal Oscillation).

21 NATURAL ENVIRONMENTAL CYCLES

22 Natural climate variability in freshwater and marine environments has the potential to 23 substantially affect salmonid abundance, particularly during early lifestages (NMFS 24 2008). Sources of variability include inter-annual climatic variations (e.g., El Niño and 25 La Niña), longer-term cycles in ocean conditions (e.g., Pacific Decadal Oscillation, 26 Mantua et al. 1997), and ongoing global climate change. Climate variability can affect 27 ocean productivity in the marine environment, as well as water storage (e.g., snow pack) 28 and in-stream flow in the freshwater environment. Early lifestage growth and survival of 29 salmon can be negatively affected when climate variability results in conditions that 30 hinder ocean productivity (e.g., Scheuerell and Williams 2005) and water storage (e.g.,

Independent Scientific Advisory Board 2007) in marine and freshwater systems,
 respectively.

3 Fisheries scientists have shown that ocean climate varies strongly at decadal scales (e.g., 4 Beamish 1993; Beamish and Bouillon 1993; Graham 1994; Miller et al. 1994; Hare and 5 Francis 1995; Mantua et al. 1997; Mueter et al. 2002). In particular, the identification of 6 the Pacific Decadal Oscillation (PDO) (Mantua et al. 1997) has led to the belief that 7 decadal-scale variation may be cyclical, and thus predictable (Lindley et al. 2007). 8 Evidence also suggests that marine survival among salmonids fluctuates in response to 9 20- to 30-year cycles of climatic conditions and ocean productivity (Hare et al. 1999 as 10 cited in NMFS 2009a; Mantua and Hare 2002). In addition, large-scale climatic regime 11 shifts, such as the El Niño condition, appear to change productivity levels over large 12 expanses of the Pacific Ocean. A further confounding effect is the fluctuation between 13 drought and wet conditions in the basins of the American west. During the first part of 14 the 1990s, much of the Pacific Coast was subject to a series of very dry years, which 15 reduced inflows to watersheds up and down the west coast.

16 "El Niño" is an environmental condition often cited as a cause for the decline of West 17 Coast salmonids (NMFS 1996). El Niño is an unusual warming of the Pacific Ocean off 18 South America and is caused by atmospheric changes in the tropical Pacific Ocean (El 19 Niño Southern Oscillation [ENSO]) resulting in reductions or reversals of the normal 20 trade wind circulation patterns. El Niño ocean conditions are characterized by anomalous 21 warm sea surface temperatures and changes to coastal currents and upwelling patterns. 22 Principal ecosystem alterations include decreased primary and secondary productivity in 23 affected regions and changes in prey and predator species distributions. Cold-water 24 species are displaced towards higher latitudes or move into deeper, cooler water, and 25 their habitat niches are occupied by species tolerant of warmer water that move upwards 26 from the lower latitudes with the warm water tongue.

A key factor affecting many West Coast stocks has been a general 30-year decline in ocean productivity. The mechanism whereby stocks are affected is not well understood, partially because the pattern of response to these changing ocean conditions has differed among stocks, presumably due to differences in their ocean timing and distribution. It is

1 presumed that survival of Chinook salmon in the ocean is driven largely by events 2 occurring between ocean entry and recruitment to a sub-adult lifestage. The freshwater 3 life history traits and habitat requirements of juvenile winter-run and fall-run Chinook 4 salmon are similar. Therefore, the unusual and poor ocean conditions that caused the 5 drastic decline in returning fall-run Chinook salmon populations coast-wide in 2007 6 (Varanasi and Bartoo 2008) are suspected to have also caused the observed decrease in the winter-run Chinook salmon spawning population in 2007 (Oppenheim 2008 as cited 7 8 in NMFS 2009a). Lindley et al. (2009) reviewed the possible causes for the decline in 9 Sacramento River fall-run Chinook salmon in 2007 and 2008 for which reliable data were 10 available. They concluded that a broad body of evidence suggested that anomalous 11 conditions in the coastal ocean in 2005 and 2006 resulted in unusually poor survival of 12 the 2004 and 2005 broods of fall-run Chinook salmon. However, Lindley et al. (2009) 13 recognize that the rapid and likely temporary deterioration in ocean conditions acted on 14 top of a long-term, steady degradation of the freshwater and estuarine environment.

As suggested by Rudnick and Davis (2003) and Hsieh et al. (2005), apparent regime shifts need not be cyclical or predictable, but rather may be the expression of a stochastic process. If this interpretation is correct, then we should expect future ocean climate conditions to be different than those observed over the past few decades (Lindley et al. 2007).

Lindley et al. (2007) further state that Central Valley salmonid ESUs and DPSs are capable of surviving the kinds of climate extremes observed over the past few thousand years if they have functional habitats, because these lineages are on order of a thousand years old or older. There is growing concern, however, that the future climate will be unlike that seen before, due to global warming in response to anthropogenic greenhouse gas emissions (Lindley et al. 2007).

26 OCEAN PRODUCTIVITY

The time when juvenile salmonids enter the marine environment marks a critical point in their life history. Studies have shown the greatest rates of growth and energy accumulation for Chinook salmon occur during the first 1 to 3 months after they enter the ocean (Francis and Mantua 2003 as cited in NMFS 2009a; MacFarlane et al. 2008 as

1 cited in NMFS 2009a). Emigration periods and ocean entry can vary substantially among, 2 and even within, runs in the Central Valley. Winter-run Chinook salmon typically rear in 3 freshwater for 5 to 9 months and exhibit a peak emigration period in March and April. 4 Spring-run Chinook salmon emigration is more variable and can occur in December or 5 January (soon after emergence as fry), or from October through March (after rearing for a 6 year or more in freshwater; Reclamation 2008). In contrast to Chinook salmon, steelhead 7 tend to rear in freshwater environments longer (anywhere from 1 to 3 years) and their 8 period of ocean entry can span many months. Juvenile steelhead presence at Chipps 9 Island has been documented between at least October and July (Reclamation 2008). 10 While still acknowledging this variability in emigration patterns, a general statement can 11 be made that Chinook salmon typically rear in freshwater environments for less than a 12 year and enter the marine environment as sub-yearlings in late spring to early summer 13 (NMFS 2009a). Similarly, although steelhead life histories are more elastic, they 14 typically enter the ocean in approximately the same time frame. The general timing 15 pattern of ocean entry is commonly attributed to evolutionary adaptations that allow 16 salmonids to take advantage of highly productive ocean conditions that typically occur off the California coast beginning in spring and extending into the fall (MacFarlane et al. 17 18 2008 as cited in NMFS 2009a). Therefore, the conditions that juvenile salmonids encounter when they enter the ocean can play an important role in their early marine 19 20 survival and eventual development into adults.

21 Variations in salmon marine survival correspond with periods of cold and warm ocean 22 conditions, with cold regimes being generally favorable for salmon survival and warm 23 regimes unfavorable (Behrenfeld et al. 2006; Wells et al. 2006). Peterson et al. (2006) 24 provide evidence that growth and survival rates of salmon in the California Current 25 System (CCS) off the Pacific Northwest can be linked to fluctuations in ocean conditions. 26 The CCS extends up to 1000 km offshore from Oregon to Baja California and 27 encompasses a southward meandering surface current, a pole-ward undercurrent and 28 surface countercurrents that exhibit high biological productivity, diverse regional 29 characteristics, and intricate eddy motions that have mystified oceanographers 30 for decades.

1 An evaluation of conditions in the CCS since the late 1970s reveals that a generally 2 warm, unproductive regime persisted until the late 1990s. This regime was followed by a 3 period of high variability that began with colder, more productive conditions lasting from 4 1999 to 2002. In general, salmon populations increased substantially during this period. 5 However, the brief cold cycle was immediately succeeded by a 4-year period of 6 predominantly warm ocean conditions beginning in late 2002, which appeared to negatively impact salmon populations in the CCS (Peterson et al. 2006). These regime 7 shifts follow a more or less linear pattern beginning with the amount and timing of 8 9 nutrients provided by upwelling and passing "up" the food chain from plankton to forage 10 fish and eventually, salmon. There are also indications that these same regime shifts 11 affect the migration patterns of larger animals that prey on salmon (e.g., Pacific hake, sea 12 birds) resulting in a "top-down" effect as well (Peterson et al. 2006).

13 Peterson et al. (2006) evaluated three sets of ecosystem indicators to identify ecological 14 properties associated with warm and cold ocean conditions and determine how those 15 conditions can affect salmon survival. The three sets of ecosystem indicators include: (1) 16 large-scale oceanic and atmospheric conditions [specifically, the PDO and the 17 Multivariate ENSO Index]; (2) local observations of physical and biological ocean 18 conditions off northern Oregon (e.g., upwelling, water temperature, plankton species 19 compositions, etc.); and (3) biological sampling of juvenile salmon, plankton, forage fish, 20 and Pacific hake (which prey on salmon). When used collectively, this information can 21 provide a general assessment of ocean conditions in the northern California Current that 22 pertain to multi-year warm or cold phases. It can also be used to develop a qualitative 23 evaluation for a particular year of the effect these ocean conditions have on juvenile 24 salmon when they enter the marine environment and the potential impact to returning 25 adults in subsequent years (NMFS 2009a).

The generally warmer ocean conditions in the California Current that began to prevail in late 2002 have resulted in coastal ocean temperatures remaining 1°C to 2°C above normal through 2005. A review of the previously mentioned indicators for 2005 revealed that almost all ecosystem indices were characteristic of poor ocean conditions and reduced salmon survival (NMFS 2009a). For instance, in addition to the high sea surface temperatures, the spring transition, which marks the beginning of the upwelling season

1 and typically occurs between March and June, was very late, postponing upwelling until 2 mid-July. In addition, the plankton species present during that time were the smaller 3 organisms with lower lipid contents associated with warmer water, as opposed to the 4 larger, lipid-rich organisms believed to be essential for salmon growth and survival 5 throughout the winter. The number of juvenile salmon collected during trawl surveys was 6 also lower than any other year previously sampled since 1998 (Peterson et al. 2006). 7 Furthermore, although conditions in 2006 appeared to have improved somewhat over 8 those observed in 2005 (e.g., sea surface temperature was cooler, the spring transition 9 occurred earlier, and coastal upwelling was more pronounced), not all parameters were 10 necessarily "good." In fact, many of the indicators were either "intermediate" (e.g., PDO, 11 juvenile Chinook salmon presence in trawl surveys) or "poor" (e.g., copepod 12 biodiversity, Peterson et al. 2006).

13 Peterson et al. (2006) shows the transition to colder ocean conditions, which began in 14 2007 and persisted through 2008. For juvenile salmon that entered the ocean in 2008, 15 ocean indicators suggested a highly favorable marine environment (NMFS 2009a). After 16 remaining neutral through much of 2007, PDO values became negative (indicating a cold 17 California Current) in late 2007 and remained negative through at least August 2008, 18 when sea surface temperatures also remained cold. Because coastal upwelling was 19 initiated early and the larger, energy-rich, coldwater plankton species were present in 20 large numbers during 2007 and 2008, ocean conditions in the broader California Current 21 appear to have been favorable for salmon survival in 2007 and to a greater extent in 2008. 22 These ecosystem indicators can be used to provide an understanding of ocean conditions, 23 and their relative impact on marine survival of juvenile salmon, throughout the broader, 24 northern portion of the California Current. However, they may not provide an accurate 25 assessment of the conditions observed on a more local scale off the California coast.

Wells et al. (2008) developed a multivariate environmental index that can be used to assess ocean productivity on a finer scale for the central California region. This index (also referred to as the Wells Ocean Productivity Index) has also tracked the Northern Oscillation Index, which can be used to understand general ocean conditions in the North Pacific Ocean. The divergence of these two indices in 2005 and 2006 provided evidence that ocean conditions were worse off the California coast than they were in the broader

1 North Pacific region. The Wells et al. (2008) index incorporates 13 oceanographic 2 variables and indices and has correlated well with the productivity of zooplankton, 3 juvenile shortbelly rockfish, and common murre production along the California coast 4 (MacFarlane et al. 2008 as cited in NMFS 2009a). In addition to its use as an indicator of 5 general ocean productivity, the index may also relate to salmon dynamics due to their 6 heavy reliance on krill and rockfish as prey items during early and later lifestages. For instance, not only did the extremely low index values in 2005 and 2006 correlate well 7 8 with the extremely low productivity of salmon off the central California coast in those 9 years, but the index also appears to have correlated well with maturation and mortality 10 rates of adult salmon from 1990-2006 in that region (Wells and Mohr 2008 as cited in 11 NMFS 2009a).

12 Available information suggests ocean conditions in 2007 and 2008 improved 13 substantially over those observed in 2005 and 2006. The spring transition, which marks 14 the beginning of the upwelling season and typically occurs between March and June, was 15 earlier in 2007 and 2008, relative to 2005 and 2006. An early spring transition is often 16 indicative of greater productivity throughout the spring and summer seasons (Wells and 17 Mohr 2008, Peterson et al. 2006). Coastal upwelling, the process by which cool, nutrient 18 rich waters are brought to the surface (perhaps the most important parameter with respect 19 to plankton productivity), was also above average in 2007 and 2008. Moreover, coastal 20 sea surface temperature and sea level height (representative of the strength of the 21 California current and southern transport) values were also characteristic of improved 22 ocean productivity (Wells and Mohr 2008). Thus, contrary to the poor ocean conditions 23 observed in the spring of 2005 and 2006, the Wells et al. (2008) index parameters 24 indicate spring ocean conditions have been generally favorable for salmon survival off 25 California in 2007 and 2008.

In contrast to the relatively "good" ocean conditions that occurred in the spring, the Wells et al. (2008) index values for the summer of 2007 and 2008 were poor in general, and similar to those observed in 2005 and 2006. Summer sea surface temperature followed a similar pattern in both 2007 and 2008, starting out cool in June, and then rising to well above average in July before dropping back down to average in August (Wells and Mohr 2008). The strong upwelling values observed in the spring of 2007 and 2008 were not

1 maintained throughout the summer, and instead dropped to either at or below those 2 observed in 2005 and 2006. Finally, sea level height and spring curl values (a 3 mathematical representation of the vertical component of wind shear which represents the 4 rotation of the vector field), which are negatively correlated with ocean productivity, 5 were both poor (Wells and Mohr 2008). Therefore, during the spring of 2007 and 2008, 6 ocean conditions off California were indicative of a productive marine environment 7 favorable for ocean salmon survival (and much improved over 2005 and 2006). However, 8 those conditions did not persist throughout the year, as Wells et al. (2008) index values 9 observed in the summer of 2007 and 2008 were similar to those experienced in the 10 summer of 2005 and 2006, two years marked by extremely low productivity of salmon 11 off the central California coast.

12 Changes in the state of the California Current since spring 2009 reflected a transition 13 from cool La Nina conditions into and through a short-lived relatively weak El Nino 14 event (Bjorkstedt et al. 2010). Weaker than normal upwelling and several extended 15 relaxation events contributed to warming over much of the California Current during 16 summer 2009, especially in the north. Moderation of La Niña conditions in the California 17 Current coincided with the development of El Niño conditions in the equatorial Pacific, 18 yet manifested well in advance of any evidence for direct effects of El Niño on the 19 California Current. Responses to El Niño in fall 2009 and winter 2009-2010 appear to 20 have varied substantially with latitude - conditions off southern California returned to 21 near climatological values with the decline of La Niña, and did not indicate any 22 subsequent response to El Niño, yet the northern California Current warmed substantially 23 following the decline of La Niña and was strongly affected by intense downwelling 24 during winter 2009–2010. The 2009–2010 El Niño diminished rapidly in early 2010, and 25 upwelling off central and southern California resumed unusually early and strongly for a 26 spring following an El Niño, but recovery from El Niño in early 2010 appears to be less 27 robust in the northern California Current. Thus, despite dynamic changes in the overall 28 state of the California Current, 2009–2010 continued the recent pattern of strong regional 29 variability across the California Current (Bjorkstedt et al. 2010).

Responses to this climate sequence exhibited some consistent patterns across the
 California Current, but regional differences noted in recent State of the California Current

reports appear to have persisted along the west coast of North America (Goericke et al. 2007; McClatchie et al. 2009). The transition from La Nina conditions appears to have unfolded well in advance of the arrival of direct effects of El Nino in the California Current in late 2009. Cool conditions related to the 2007–2008 La Nina abated in summer 2009, and, in general terms, hydrographic and ecological conditions from southern California north approached climatological values during summer 2009 (Bjorkstedt et al. 2010).

8 Warmer than usual conditions had already developed off Baja California in 2008 and 9 persisted into the current year, but showed similar directional responses to climate 10 variability as did regions to the north (Bjorkstedt et al. 2010). Overall, changes in the 11 state of the California Current during 2009 coincided with the decay of La Nina 12 conditions in the tropical Pacific Ocean. In the context of the general pattern of transition 13 from La Nina to El Nino, differences between the northern and southern regions of the 14 California Current are readily apparent. Off southern California, the general trend was for 15 mean hydrographic, chemical, and biological properties of the system to return to long-16 term average conditions during summer 2009. In contrast, the northern California Current 17 experienced anomalous warming of coastal waters and associated ecosystem responses, 18 presumably as a consequence of anomalously weak and intermittent upwelling during 19 2009. Likewise, regional differences and similarities are apparent from late fall 2009 20 through spring 2010, the period during which El Nino conditions propagated into the 21 California Current and subsequently diminished. Off southern California, the arrival of El 22 Nino was clearly indicated by anomalously high sea level, but responses to El Nino were 23 limited to changes in isopynchal depth—presumably related to the passage of poleward-24 propagating Kelvin waves and their lingering consequences (Bjorkstedt et al. 2010).

Coastal waters off Oregon and northern California were affected by unusually strong downwelling during winter 2009–2010. In neither case, however, was there any evidence for intrusion of unusual water masses such as had been observed during the strong 1997– 1998 El Nino. Relatively strong positive anomalies in temperature and salinity off southern Baja California suggest that the 2009–2010 El Nino influenced the southern extent of the California Current, but these changes appear to have been a consequence of local circulation patterns rather than anomalous poleward flows (Bjorkstedt et al. 2010).

1 Copepod assemblages observed at mid-shelf stations off northern California and Oregon 2 continued to show marked seasonal variation, with high abundances developing over the 3 summer and into the fall and subsequently declining over the winter (Bjorkstedt et al. 4 2010). Total abundance of copepods over the shelf appears to have been lower or later in 5 developing in summer 2009 than in 2008 in sampled areas of the northern California 6 Current. Patterns in assemblage structure, as indicated by the abundance of species 7 particular biogeographic affinities (e.g., southern (warm) v. northern (cold), neritic v. 8 oceanic; Hooff and Peterson 2006), show a substantial degree of coherence since 2008, 9 particularly at stations north of Cape Mendocino. Compared to winter 2009, the 10 composition of copepod assemblages off Oregon and northern California shifted strongly 11 towards being dominated by southern and oceanic species by winter 2010. Southern taxa 12 were abundant off Bodega Bay in late 2008, coincident with warm temperatures, but 13 largely disappeared from mid-shelf waters in early 2009, possibly as a consequence of 14 intense transport. Although warm water and reduced flows were observed in summer 15 2009 off Bodega, total copepod abundance did not reach high abundances and southern 16 taxa did not assume a dominant place in the assemblage until winter 2010 (Bjorkstedt et al. 2010). 17

Catches of juvenile salmonids in pelagic surface trawl surveys were unusually low during September 2009 (Bjorkstedt et al. 2010). The fewest juvenile coho salmon (*Oncorhynchus kisutch*; 2 compared to maximum catch of 158 in 1999) and sub-yearling Chinook salmon (*O. tschawytschwa*; 2 versus 465 in 2001) were caught since the beginning of the time series in 1998. Overall spring 2009 appeared to be relatively good for salmon marine survival but oceanographic conditions appear to have deteriorated for salmon by late summer 2009 (Bjorkstedt et al. 2010).

In 2008 and 2009, poor Sacramento returns, primarily supported by Sacramento River fall-run Chinook salmon, led to the largest fishery closure on record. In 2009, adult spawning escapement for Sacramento River fall Chinook failed to meet the escapement goal (122,000-180,000 adults) for the third year in a row, leading to the formal declaration of an overfishing concern (although fishing is not considered one of the major causes of the stock's decline). The forecast for the index of ocean abundance in 2010 was 245,500 adults, which provided adequate numbers for limited fisheries (PFMC 2011).

1 Ecosystem observations offer further suggestion of regional variation in responses to El 2 Nino, but it must be noted that such comparisons are limited by disparity in available data 3 sets (Bjorkstedt et al. 2010). Off southern California, estimates of nutrient concentrations, 4 chlorophyll a standing stock, primary productivity, and zooplankton displacement 5 volumes returned to "normal" levels, and did not show evidence for any decline associated with El Nino. In contrast, anomalies in chlorophyll a concentration shifted 6 7 from positive to negative off Baja California, especially north of Point Eugenia, despite 8 the lack of concomitantly strong changes in hydrographic conditions. Responses at higher 9 trophic levels are much more difficult to connect to simple indices of climate variability, 10 but provide insight to the potential magnitude of ecosystem responses to conditions 11 leading into spring 2009 and the consequences of the 2009-2010 El Nino relative to 12 previous El Ninos. Positive shifts in indices of abundance for the juvenile groundfish 13 assemblage off central California and breeding success of Cassin's Auklet in 2009 are 14 consistent with the persistence of cool conditions into spring 2009. Interestingly, the 15 pelagic juvenile groundfish assemblage did not appear to collapse in 2010, suggesting 16 that El Nino conditions did not substantially diminish productivity available to these taxa 17 during critical lifestages during winter and early spring. In contrast, juvenile salmonids at 18 sea in the northern region of the California Current appear to have fared poorly during the 19 warmer than usual conditions of summer and fall 2009. Changes in the copepod 20 assemblage off Oregon were consistent with warmer conditions that do not favor salmon 21 production (Peterson and Schwing 2003; Peterson et al. 2010).

22 In summary, the significant changes in the state of the California Current during 2009 23 and early 2010 appear to have been more closely associated with diminishment of La 24 Nina conditions than direct effects of El Nino (Bjorkstedt et al. 2010). The signature of 25 the 2009–2010 El Nino throughout much of the California Current was substantially 26 weaker than that of the strong 1997–1998 El Nino when influxes of more tropical waters 27 were observed throughout the California Current. While the 2009-2010 El Nino is 28 perhaps most comparable to the mild 2002–2003 El Nino, direct comparisons between 29 the two events are confounded by the interaction of the 2002–2003 El Nino with a 30 coincident intrusion of subarctic water that affected much of the California Current 31 (Venrick et al. 2003). The more dramatic changes observed during 2009-2010 in the

northern California Current might reflect responses to atmospheric forcing favoring
coastal warming absent countervailing subarctic influences. Because a transition to
moderate La Nina conditions was forecast for summer 2010, the past year might
represent a temporary interruption of an otherwise cool period in the California Current
(Bjorkstedt et al. 2010).

6 NMFS (2009a) suggests that early marine survival for juvenile salmon is a critical phase 7 in their survival and development into adults. The correlation between various environmental indices that track ocean conditions and salmon productivity in the Pacific 8 9 Ocean, both on a broad and local scale, provides an indication of the role they play in 10 salmon survival in the ocean. Moreover, when discussing the potential extinctions of 11 salmon populations, Francis and Mantua (2003) state that climate patterns would not 12 likely be the sole cause but could certainly increase the risk of extinction when combined 13 with other factors, especially in ecosystems under stress from humans. Thus, the efforts 14 to try and gain a greater understanding of the role ocean conditions play in salmon 15 productivity will continue to provide valuable information that can be incorporated into 16 the management of these species and should continue to be pursued. However, the highly 17 variable nature of these environmental factors makes it very difficult, if not impossible, to 18 accurately predict what they will be like in the future. Because the potential for poor 19 ocean conditions exists in any given year, and because there is no way for salmon 20 managers to control these factors, any deleterious effects endured by salmonids in the 21 freshwater environment can only exacerbate the problem of an inhospitable marine 22 environment (NMFS 2009a).

23 GLOBAL CLIMATE CHANGE

Warming over this century is projected to be considerably greater than over the last century (Thomas et al. 2009). Since 1900, the global average temperature has risen by about 1.5°F. By about 2100, it is projected to rise between 2°F and 10.5°F, but could increase up to 11.5°F (Thomas et al. 2009; California Climate Change Center 2006). In the United States, the average temperature has risen by a comparable amount and is very likely to rise more than the global average over this century, with some variation according to location. Regarding climate change impacts already being observed, the

1 Sierra Nevada Alliance (2008) reports that seven of the largest Sierra glaciers have 2 retreated by 30 to 70% in the past 100 years. Changes observed over the past several 3 decades also have shown that the earth is warming, and scientific evidence suggests that 4 increasing greenhouse gas emissions are changing the earth's climate (Moser et al. 2009). 5 Accumulating greenhouse gas concentrations in the earth's atmosphere have been linked 6 to global warming, and projected future trends of increasing atmospheric greenhouse gas 7 concentrations suggest global warming will continue (National Research Council 2001). 8 Several factors will determine future temperature increases. Increases at the lower end of 9 this range are more likely if global heat-trapping gas emissions are substantially reduced. 10 If emissions continue to rise at or near current rates, temperature increases are more 11 likely to be near the upper end of the range (NMFS 2009).

12 Global climate change has the potential to impact numerous environmental resources in 13 California through potential, though uncertain, impacts related to future air temperatures 14 and precipitation patterns, and the resulting implications to stream runoff rate and timing, 15 water temperatures, reservoir operations, and sea levels. Although current models are 16 broadly consistent in predicting increases in probable global air temperatures and 17 increasing levels of greenhouse gasses resulting from human activities, there are 18 considerable uncertainties about precipitation estimates. For example, many regional 19 modeling analyses conducted for the western United States indicate that overall 20 precipitation will increase, but uncertainties remain due to differences among larger-scale 21 General Circulation Models (GCMs) (Kiparsky and Gleick 2003). Some researchers 22 believe that climate warming might push the storm track on the West Coast further north, 23 which would result in drier conditions in California. At the same time, relatively newer 24 GCMs, including those used in the National Water Assessment, predict increases in 25 California precipitation (DWR 2005). Similarly, two popular climate models, including 26 HadCM2 developed by the U.K. Hadley Center and PCM developed by the U.S. National 27 Center for Atmospheric Research, also predict very different future scenarios. The 28 HadCM2 predicts wetter conditions while the PCM predicts drier conditions (Brekke 29 et al. 2004).

30 While much variation exists in projections related to future precipitation patterns, all 31 available climate models predict a warming trend resulting from the influence of rising

1 levels of greenhouse gasses in the atmosphere (Barnett et al. 2005). The potential effects 2 of a warmer climate on the seasonality of runoff from snowmelt in the Central Valley 3 have been well-studied and results suggest that melt runoff will likely shift from spring 4 and summer to earlier periods in the water year (Vanrheenen et al. 2001). Presently, 5 snow accumulation in the Sierra Nevada acts as a natural reservoir for California by 6 delaying runoff from winter months when precipitation is high (Kiparsky and Gleick 7 2003). However, compared to present water resources development, Null et al. (2010) 8 report that watersheds in the Northern Sierra Nevada are most vulnerable to decreased 9 mean annual flow, southern-central watersheds are most susceptible to runoff timing 10 changes, and the central portion of the range is most affected by longer periods with low 11 flow conditions. Despite the uncertainties about future changes in precipitation rates, it is 12 generally believed that higher temperatures will lead to changes in snowfall and 13 snowmelt dynamics. Higher atmospheric temperatures will likely increase the ratio of 14 rain to snow, shorten and delay the onset of the snowfall season, and accelerate the rate of 15 spring snowmelt, which would lead to more rapid and earlier seasonal runoff relative to 16 current conditions (Kiparsky and Gleick 2003). Studies suggest that the spring stream flow maximum could occur about one month earlier by 2050 (Barnett et al. 2005). 17

18 If air temperatures in California rise significantly, it will become increasingly difficult to 19 maintain appropriate water temperatures in order to manage coldwater fisheries, 20 including salmonids. A reduction in snowmelt and increased evaporation could lead to 21 decreases in reservoir levels and, perhaps more importantly, coldwater pool reserves 22 (California Energy Commission 2003). As a result, increasing air temperatures, 23 particularly during the summer, lead to rising water temperatures in rivers and streams, 24 which increase stress on coldwater fish. Projected temperatures for the 2020s and 2040s 25 under a higher emissions scenario suggest that the habitat for these fish is likely to 26 decrease dramatically (Mote et al. 2008; Salathé 2005; Keleher and Rahel 1996; 27 McCullough et al. 2001). Reduced summer flows and warmer water temperatures will 28 create less favorable instream habitat conditions for coldwater fish species.

In the Central Valley, by 2100 mean summer temperatures may increase by 2 to 8°C, precipitation will likely shift to more rain and less snow, with significant declines in total precipitation possible, and hydrographs will likely change, especially in the southern 1 Sierra Nevada mountains (NMFS 2009). Thus, climate change poses an additional risk to 2 the survival of salmonids in the Central Valley. As with their ocean phase, Chinook 3 salmon and steelhead will be more thermally stressed by stream warming at the southern 4 ends of their ranges (e.g., Central Valley Domain). For example, warming at the lower 5 end of the predicted range (about 2°C) may allow spring-run Chinook salmon to persist 6 in some streams, while making some currently utilized habitat inhospitable (Lindley et al. 7 2007). At the upper end of the range of predicted warming, very little spring-run Chinook 8 salmon habitat is expected to remain suitable (Lindley et al. 2007).

9 Under the expected warming of around 5°C, substantial amounts of habitat would be lost, 10 with significant amounts of habitat remaining primarily in the Feather and Yuba rivers, 11 and remnants of habitat in the upper Sacramento, McCloud, and Pit rivers, Battle and 12 Mill creeks, and the Stanislaus River (Lindley et al. 2007). Under the less likely but still 13 possible scenario of an 8°C warming, spring-run Chinook salmon habitat would be found 14 only in the upper-most reaches of the north fork Feather River, Battle Creek, and Mill 15 Creek. This simple analysis suggests that Central Valley salmonids are vulnerable to 16 warming, but more research is needed to evaluate the details of how warming would 17 influence individual populations and subbasins.

As summarized by Lindley et al. (2007), climate change may pose new threats to Central Valley salmonids by reducing the quantity and quality of freshwater habitat. Under the worst case scenario, spring-run Chinook salmon may be driven extinct by warming in this century, while the best-case scenario may allow them to persist in some streams, although prediction of the future status of Central Valley salmonids associated with long-term climate change is fraught with uncertainty.

By contrast to the conditions for other Central Valley floor rivers, climate change may not be likely to have such impacts on salmonids in the lower Yuba River downstream of Englebright Reservoir (YCWA 2010a). Presently, the lower Yuba River is one of the few Central Valley tributaries that consistently has suitable water temperatures for salmonids throughout the year. Lower Yuba River water temperatures generally remain below 58°F year-round at the Smartsville Gage (downstream of Englebright Dam), and below 60°F year-round at Daguerre Point Dam (YCWA et al. 2007). At Marysville, water temperatures generally remain below 60°F from October through May, and below 65°F
 from June through September (YCWA et al. 2007).

3 According to YCWA (2010), because of specific physical and hydrologic factors, the 4 lower Yuba River is expected to continue to provide the most suitable water temperature 5 conditions for anadromous salmonids of all Central Valley floor rivers, even if there are 6 long-term climate changes. This is because New Bullards Bar Reservoir is a deep, steep-7 sloped reservoir with ample coldwater pool reserves. Throughout the period of operations 8 of New Bullards Bar Reservoir (1969 through present), which encompasses the most 9 extreme critically dry year on record (1977), the coldwater pool in New Bullards Bar 10 Reservoir never was depleted. Since 1993, coldwater pool availability in New Bullards 11 Bar Reservoir has been sufficient to accommodate year-round utilization of the 12 reservoir's lower level outlets to provide cold water to the lower Yuba River. Even if 13 climate conditions change, New Bullards Bar Reservoir still will have a very substantial 14 coldwater pool each year that will continue to be available to provide sustained, relatively 15 cold flows of water into the lower Yuba River during the late spring, summer and fall of 16 each year (YCWA 2010).

17 OCEAN ACIDIFICATION

Ocean acidification has been called a "sister" or co-equal problem to climate change because it is caused by the same human-caused production of large amounts of CO₂. Its impacts are additional to, and may exacerbate, the effects of climate change (Alaska Marine Conservation Council 2011).

22 Seawater pH is a critical variable in marine systems. Today's surface ocean water is 23 slightly alkaline, with a pH ranging from 7.5 to 8.5 and it is saturated with calcium 24 carbonate, a very important organic molecule for organisms like corals, mollusks and 25 crustaceans that make shells. As CO_2 reacts with the seawater, it lowers the pH and 26 releases hydrogen ions. These ions bind strongly with carbonate, preventing it from 27 forming the important calcium carbonate molecules. If the pH of the global oceans drops 28 0.4 by the end of the century as predicted, the levels of calcium carbonate available for 29 use by marine organisms will decrease by 50% (Alaska Marine Conservation 30 Council 2011).

Ocean acidification is likely to alter the biodiversity of the world's marine ecosystems and may affect the total productivity of the oceans. Previously it was thought that these changes would take centuries, but new findings indicate that an increasingly acidic environment could cause problems in high-latitude marine ecosystems within just a few decades (Alaska Marine Conservation Council 2011).

6 Currently, the oceans' surface water layers have sufficient amounts of calcium carbonate 7 for organisms to use (known as saturated conditions). This calcium carbonate rich layer is 8 deeper in warmer regions and closer to the surface in colder regions. Because calcium 9 carbonate is less stable in colder waters, marine life in the polar oceans will be affected 10 by calcium carbonate loss first. A study published in Nature by 27 U.S. and international 11 scientists stated, "Some polar and sub-polar waters will become under-saturated [at 12 twice the pre-industrial level of CO₂, 560 ppm], probably within the next 50 years" (Orr et 13 al. 2005). Under-saturated refers to conditions in which the seawater has some calcium 14 carbonate remaining, but it does not have enough available for the organisms to build 15 strong shells (Alaska Marine Conservation Council 2011).

Research has shown that lowered ocean pH will affect the processes by which animals such as corals, mollusks and crustaceans make their support structures. Because these organisms depend on calcium carbonate, increasing acidity threatens their survival. At higher levels of acidity (lower pH levels), any organism that forms a shell through calcification — from clams to pteropods — could be adversely affected. These species use the naturally occurring carbonate minerals calcite and aragonite for the calcification process.

23 Pteropods are small planktonic mollusks that are at the bottom of the food chain and 24 because of their dependence on calcium carbonate, they will be one of the first casualties 25 of increasing acidity in Alaska's marine waters. In recent experiments exposing live 26 pteropods to the conditions predicted by "business-as-usual" carbon emission scenarios -27 the pteropod shells showed evidence of dissolution and damage within only 48 hours. 28 Pteropods are a key food source for salmon and other species (Alaska Marine 29 Conservation Council 2011). Increased research into ocean acidification caused by the 30 saturation of water with carbon dioxide suggests that a 10% decline in pteropod production can lead to a 20% reduction in the body weight of mature salmon (Climate Solutions 2011). A decrease in these mineral levels to food web base species like pteropods, also known as sea butterflies, which make up 45% of the diet for juvenile pink salmon, can cause cascading waves of disruption up the food chain (Climate Solutions 2011).

6 <u>Non-NATIVE INVASIVE SPECIES</u>

7 Non-native invasive species are of concern throughout the ESU and DPSs and can result 8 in numerous deleterious effects to native species. For example, introduction of non-native 9 invasive species can alter the natural food webs that existed prior to their introduction, as 10 illustrated by the Asiatic freshwater clams Corbicula fluminea and Potamocorbula 11 *amurensis* in the Delta. Cohen and Moyle (2004) report that the arrival of these two clam 12 species disrupted the normal benthic community structure, and depressed phytoplankton 13 levels in the Delta due to the highly efficient filter feeding of the introduced clams. 14 Declines in phytoplankton levels have consequently resulted in reduced populations of 15 zooplankton that feed upon them, thereby reducing the forage base available to salmonids 16 transiting through the Delta and the San Francisco estuary on their ocean migrations. The 17 lack of forage base can adversely affect the health and physiological condition of 18 salmonids as they migrate to the Pacific Ocean.

Attempts to control non-native invasive plant species also can adversely affect the health and habitat suitability of salmonids within affected water systems, through either direct exposure to toxic chemicals or reductions in DO levels associated with the decomposition of vegetative matter in the water. As an example, control programs for the invasive water hyacinth and *Egeria densa* plants in the Delta must balance the toxicity of the herbicides applied to control the plants against the probability of exposure to listed salmonids during herbicide application period.

26 **4.2.6.2** Lower Yuba River

The phenotypic lower Yuba River spring-run Chinook salmon population is exposed and subject to the myriad of limiting factors, threats and stressors described above for the Central Valley ESU. Lower Yuba River phenotypic spring-run Chinook salmon generally 1 spend a few months (with some individuals remaining up to several months, or a year) in 2 the lower Yuba River prior to migrating downstream through the lower Feather River, the 3 lower Sacramento River, the Delta, and San Francisco Bay to the Pacific Ocean, where 4 they spend from two to four years growing and maturing. Following their ocean 5 residency, these fish then undertake an upstream migration through this same system, and 6 are again exposed to the associated limiting factors, threats and stressors, prior to spending a few additional months in the lower Yuba River holding and 7 subsequently spawning. 8

9 Three separate efforts have been undertaken over the past few years to identify, 10 characterize and prioritize limiting factors (i.e., "stressors") for anadromous salmonids 11 (including spring-run Chinook salmon) in the lower Yuba River. The Lower Yuba River 12 Fisheries Technical Working Group, a multi-party stakeholder group including the Corps 13 and YCWA, established a process to rank stressors as part of the "Draft Implementation 14 Plan for Lower Yuba River Anadromous Fish Habitat Restoration" (CALFED and 15 YCWA 2005). The Yuba Accord Technical Team built upon these efforts and utilized a 16 stressor analysis in the development of the Yuba Accord minimum flow requirements 17 (i.e., "flow schedules") (YCWA et al. 2007).

18 Most recently, NMFS (2009) conducted a comprehensive assessment of stressors 19 affecting spring-run Chinook salmon both within the lower Yuba River, and affecting 20 lower Yuba River populations as they migrate downstream (as juveniles) and upstream 21 (as adults) through the lower Feather River, the lower Sacramento River, and the Bay-22 Delta system.

As stated by NMFS (2009), stressor matrices, which structured hierarchically related tiers in order to prioritize stressors, were developed. After all of the variables in the matrix were identified and weighted, stressors within the matrices were sorted in descending order (from the highest to the lowest biological impact). Although the resultant sorted matrices provide a pseudo-quantitative means of comparatively ranking individual stressors, to avoid attributing unwarranted specificity to the prioritized stressor list, it was distributed into four separate quartiles ("Very High", "High", "Medium", and "Low"). 1 The ranking and quartile characterization of stressors were organized such that stressors

- 2 affecting the individual lifestages also could be ascertained.
- 3 According to NMFS (2009a), for the lower Yuba River population of spring-run Chinook
- 4 salmon, the number of stressors according to the categories of "Very High", "High",
- 5 "Medium", and "Low" that occur in the lower Yuba River or occur out of basin are
- 6 presented below by lifestage (**Table 4-3**).

7 Table 4-3. The number of stressors according to the categories of "Very High", "High", 8 "Medium", and "Low" that occur in the lower Yuba River, or occur out-of-basin, by 9 lifestage for the lower Yuba River population of spring-run Chinook salmon (Source: 10 NMFS 2009a).

| Lifestage | Location | Stressor Categories | | | |
|--|------------------|---------------------|------|--------|-----|
| | | Very High | High | Medium | Low |
| Adult Immigration and Holding | | | | | |
| | Lower Yuba River | 2 | 1 | 3 | 1 |
| | Out of Basin | 1 | 5 | 8 | 6 |
| Spawning | | | | | |
| | Lower Yuba River | 3 | 2 | 0 | 2 |
| | Out of Basin | N/A* | N/A | N/A | N/A |
| Embryo Incubation | | | | | |
| | Lower Yuba River | 1 | 0 | 4 | 0 |
| | Out of Basin | N/A | N/A | N/A | N/A |
| Juvenile Rearing and Outmigration | | | | | |
| | Lower Yuba River | 5 | 1 | 1 | 5 |
| | Out of Basin | 12 | 16 | 6 | 9 |
| * Not Applicable. These lifestages for this population only occur in the lower Yuba River. | | | | | |

11 As shown by the numbers in Table 4-3, of the total number of 94 stressors affecting all 12 identified lifestages of the lower Yuba River populations of spring-run Chinook salmon, 13 31 are within the lower Yuba River and 63 are out-of-basin. Because spawning and 14 incubation occurs only in the lower Yuba River, all of the stressors associated with these 15 lifestages occur in the lower Yuba River. Therefore, for the adult immigration and holding, and the juvenile rearing and outmigration lifestages combined, a total of 49 16 17 "Very High" and "High" stressors were identified, with 15 of those occurring in the 18 lower Yuba River and 34 occurring out-of-basin.

1 The NMFS (2009) Draft Recovery Plan states that "The lower Yuba River, below 2 Englebright Dam, is characterized as having a high potential to support a viable 3 independent population of spring-run Chinook salmon, primarily because: (1) flow and 4 water temperature conditions are generally suitable to support all lifestage requirements; 5 (2) the river does not have a hatchery on it; (3) spawning habitat availability is believed 6 not to be limiting; and (4) high habitat restoration potential".

7 The NMFS (2009) Draft Recovery Plan further states that "For currently occupied 8 habitats below Englebright Dam, it is unlikely that habitats can be restored to pre-dam 9 conditions, but many of the processes and conditions that are necessary to support a 10 viable independent population of spring-run Chinook salmon can be improved with 11 provision of appropriate instream flow regimes, water temperatures, and habitat 12 availability. Continued implementation of the Yuba Accord is expected to address these 13 factors and considerably improve conditions in the lower Yuba River."

14 PASSAGE IMPEDIMENTS/BARRIERS

Englebright Dam was not designed for fish passage and presents an impassable barrier to the upstream migration of anadromous salmonids, and marks the upstream extent of currently accessible spring-run Chinook salmon habitat in the lower Yuba River, whereas Daguerre Point Dam presents an impediment to upstream migration.

Englebright Dam, built in 1941 to retain hydraulic mining debris from the Yuba River
watershed, blocks upstream migration of fish in the lower Yuba River and, in particular,
blocks the migration of steelhead and spring-run Chinook salmon to their historic
spawning grounds (NMFS 2002).

Daguerre Point Dam has been reported to be an impediment to upstream migration of adult salmon and steelhead under certain conditions. Factors contributing to impeded adult spring-run Chinook salmon upstream passage have been suggested to include inadequate attraction flows to the ladders, proximity and orientation of the ladder entrances to the spillway, periodic obstruction of the ladders by sediment and woody debris, and other fish ladder physical design issues.

1 Sheet flow across the dam's spillway, particularly during high-flow periods, may obscure 2 ladder entrances and, thus, makes it difficult for immigrating adult salmonids to find the 3 entrances (NMFS 2007a). For example, fall-run Chinook salmon have been observed 4 attempting to leap over the dam, demonstrating that these fish may have difficulty in 5 finding the fish ladder entrances (Corps 2000). This phenomenon may particularly affect 6 spring-run Chinook salmon, because peak spring-run adult Chinook salmon upstream 7 migration occurs primarily during the relatively high-flow periods of spring through early 8 summer. Since 2001, wooden flashboards have been periodically affixed to the crest of 9 the dam during low flow periods to aid in directing the flows towards the fish ladder 10 entrances. Fish passage monitoring data from 2006 indicates that the installation of the 11 flashboards resulted in an immediate and dramatic increase in the passage of salmon up 12 the ladders, and is thought to have improved the ability of salmon to locate and enter the 13 ladders (NMFS 2007a).

14 Both the north and south fish ladders at Daguerre Point Dam, particularly the north 15 ladder, historically tended to clog with woody debris and sediment, which had the 16 potential to block passage or substantially reduce attraction flows at the ladder entrances. 17 Additionally: (1) the north and south ladders' exits are close to the spillway, potentially 18 resulting in adult fish exiting the ladder being immediately swept by flow back over the 19 dam; (2) sediment accumulates at the upstream exits of the fish ladders, reducing the 20 unimpeded passage from the ladders to the main channel, and may cause potential "fall-21 back" into the ladders; and (3) fish could jump out of the upper bays of the fishway, 22 resulting in direct mortality. Many of the past issues associated with woody debris 23 accumulation have either been eliminated or minimized since locking metal grates were 24 installed over the unscreened bays on the north and south fish ladders during 2011.

The RMT (2013) examined passage of adult Chinook upstream of Daguerre Point Dam and corresponding flow data during eight years of available data. Chinook salmon passage was observed over a variety of flow conditions, including ascending or descending flows, as well as during extended periods of stable flows. Flow thresholds prohibiting passage of Chinook salmon through the ladders at Daguerre Point Dam were not apparent in the data (RMT 2013).

1 Phenotypic spring-run Chinook salmon (those entering the lower Yuba River during 2 spring months) may remain in the lower Yuba River in areas downstream (and 3 proximate) to Daguerre Point Dam for extended periods of time during the spring and 4 summer. It is uncertain whether, or to what extent, the duration of residency in the large 5 pool located downstream of Daguerre Point Dam is associated with upstream passage 6 impediment and delay, or volitional habitat utilization prior to spawning in upstream 7 areas. However, RMT (2013) reported that temporal migrations of adult phenotypic 8 spring-run Chinook salmon to areas upstream of Daguerre Point Dam occurred over an 9 extended period of time. The tagged spring-run Chinook salmon in the lower Yuba River 10 actually migrated upstream of Daguerre Point Dam from May through September, and 11 utilized a broad expanse of the lower Yuba River during the phenotypic summer holding 12 period, including areas as far downstream as Simpson Lane Bridge (i.e., ~RM 1.8), and 13 as far upstream as the area just below Englebright Dam. A longitudinal analysis of 14 acoustic tag detection data indicated that distributions were non-random, and that the 15 spring-run Chinook salmon were selecting holding tagged locations for 16 (RMT 2013).

17 NMFS (2007) suggested that delays resulting from adult spring-run Chinook salmon 18 adult passage impediments could weaken fish by requiring additional use of fat stores 19 prior to spawning, and potentially could result in reduced spawning success (i.e., 20 production) from reduced resistance to disease, increased pre-spawning mortality, and 21 reduced egg viability. However, these statements suggesting biological effects associated 22 with fish passage issues at Daguerre Point Dam are not supported by studies or 23 referenced literature. For example, the RMT (2010b) included evaluation of water 24 temperatures at Daguerre Point Dam during the spring-run Chinook salmon adult 25 upstream immigration and holding lifestage, which addressed considerations regarding 26 both water temperature effects to pre-spawning adults and egg viability. They concluded 27 that during this lifestage, characterized as extending from April through August, water 28 temperatures [modeled] at Daguerre Point Dam are suitable and remain below the 29 reported optimum water temperature index value of 60°F at least 97% of the time over all 30 water year types during these months. Thus, it is unlikely that this represents a 31 significant source of mortality to spring-run Chinook salmon. Moreover, actual data monitored since the Yuba Accord has been implemented (October 2006 to June 2013)
demonstrates that water temperatures at Daguerre Point Dam actually remained at about
or below 60°F during the adult immigration and holding period each of the six years
(RMT 2013).

As reported by NMFS (2007), Daguerre Point Dam may adversely affect outmigration success of juvenile salmon and steelhead. During downstream migration, juvenile Chinook salmon and steelhead may be disoriented or injured as they plunge over the spillway, increasing their exposure and vulnerability to predators in the large pool at the base of the dam (NMFS 2007).

10 HARVEST/ANGLING IMPACTS

11 Fishing for Chinook salmon on the lower Yuba River is regulated by CDFW. Although 12 harvest/angler impacts were previously listed as a stressor, the magnitude of this potential 13 stressor has been reduced associated with changes in fishing regulations over time. 14 Angling regulations on the lower Yuba River are intended to protect sensitive species, in 15 particular spring-run Chinook salmon (and wild steelhead). CDFW angling regulations 16 2013-2014 (CDFW 2013a) state that the lower Yuba River from its confluence with the 17 lower Feather River up to Englebright Dam is closed year-round to salmon fishing, and 18 no take or possession of salmon is allowed.

Fishing for hatchery trout or hatchery steelhead is allowed on the lower Yuba River from its confluence with the lower Feather River up to the Highway 20 Bridge year-round. The lower Yuba River, between the Highway 20 Bridge and Englebright Dam, is closed to fishing from September through November to protect spring-run Chinook salmon spawning activity and egg incubation.

Although these regulations are intended to specifically protect spring-run Chinook salmon, anglers can potentially harass, harm and kill listed species (spring-run Chinook salmon and wild steelhead) through incidental actions while targeting non-listed species. Examples of potential angler impacts may include, but are not necessarily limited to, angler harvest, physical disturbance of salmonid redds, hooking and catch-and-release stress or mortality, including that which results from incidental hooking (CALFED and YCWA 2005).

1 **POACHING**

2 Whether poaching represents a stressor, or the extent to which spring-run Chinook3 salmon are targeted for poaching in the lower Yuba River is unknown.

4 Poaching of adult Chinook salmon at the fish ladders and at the base of Daguerre Point 5 Dam has been previously reported in several documents. Poaching has been previously 6 reported as a "chronic problem" (Falxa 1994 as cited in CALFED and YCWA 2005). 7 The spring-run Chinook salmon status report (CDFG 1998) stated that poaching was an 8 "ongoing problem" at Daguerre Point Dam. Poaching of salmon has been reported as a 9 "long-standing problem" on the Yuba River, particularly at Daguerre Point Dam (John 10 Nelson, CDFG, pers. comm., November 2000, as cited in NMFS 2005a). The Corps 11 (2001) and NMFS (2009) both refer to poaching of adult salmon at the Daguerre 12 Point Dam.

13 Although these previous reports refer in some fashion to poaching within the fish ladders 14 and immediately downstream of Daguerre Point Dam as issues, the only actual account of 15 documented poaching was provided by Nelson (2009). In his declaration, Nelson (2009) 16 stated that during his tenure at CDFW (which extended until 2006) he personally 17 observed people fishing illegally in the ladders, and further observed gear around the 18 ladders used for poaching. It is not clear regarding the time period to which he was 19 referring, although it may have been referring to the period prior to 2000 (see reference in 20 previous paragraph).

21 The VAKI Riverwatcher infrared and videographic sampling system began operations in 22 2003. CDFW monitored these operations at Daguerre Point Dam seasonally from 2003 23 through 2005. Since 2006 (with implementation of the Yuba Accord Pilot Programs 24 (2006 – 2007) and the Yuba Accord in 2008), PSMFC staff have monitored the system at 25 Daguerre Point Dam on a nearly daily basis, year-round, through the present. Over this 26 8-year period, neither CDFW nor PSMFC staff have reported poaching in the ladders, or 27 immediately downstream of Daguerre Point Dam. Thus, although poaching has been 28 reported as a stressor, it is unclear whether, or to what extent, it impacts the spring-run 29 Chinook salmon population in the lower Yuba River. According to Sprague (2011), the 30 amount of poaching from the fish ladders has not been quantified, and there does not appear to be data on the amount of poaching, so the extent of the problem is not
 well understood.

3 Moreover, it is unclear whether these previous reports of poaching were directed toward 4 spring-run or fall-run Chinook salmon. While data are not available as to the fish species 5 targeted, poachers likely target the fish that are readily available. The greatest numbers 6 of poached fish probably would be fall-run Chinook salmon because they congregate 7 below the dam in large numbers under the low-flow, clear-water conditions of October 8 and November (Corps 2001). According to NMFS (2002), fall-run Chinook salmon are 9 most likely to be subject to poaching because they are the largest salmonid population in 10 the lower Yuba River. Nevertheless, spring-run Chinook salmon also may be affected 11 because they may be present in the lower Yuba River during the periods of the highest 12 recreational use (NMFS 2002).

13 As early as 2001, the Corps (2001) suggested that although poaching is likely very 14 limited, fencing or screening of the ladder could further reduce or eliminate any 15 poaching. Nelson (2009) suggested that one measure that could reduce poaching would 16 be to place grates over the top of the ladders to restrict poacher access. He further 17 suggested that grates had been installed on other fish ladders to prevent poaching, such as 18 on the Woodbridge Irrigation District Dam fish ladders located on the Mokelumne River 19 However, Sprague (2011) stated that grates are not near Woodbridge, California. 20 recommended, due to the multiple sharp edges and the potential for resultant fish injury. 21 He further suggested that solid covers could be used, but consideration should be given to 22 the potential for how to avoid pressurizing the fish ladders during high flow events. As a 23 temporary solution addressing the potential for fish to jump out of the ladder (and 24 potential poaching within the fish ladders), in 2011 the Corps installed plywood boards 25 over the upper bays at the south ladder at Daguerre Point Dam. As previously discussed, 26 the July 25, 2011 Interim Remedy Order issued by the Court ordered the Corps to install 27 locking metal grates over all but the lower eight bays of the fish ladders at Daguerre Point 28 Dam by September 14, 2011. In response to the Interim Remedy Order issued by the 29 Court on July 25, 2011, during the summer of 2011 the Corps proceeded with installation 30 of locking metal grates on all 33 unscreened bays. Due to concerns expressed by both 31 NMFS and CDFW, the Court then reconsidered the requirement to put grates over the bays on the lowermost section of the south fish ladder at Daguerre Point Dam.
 Consequently, grates were not installed over the lower eight bays of the south fish ladder
 at Daguerre Point Dam.

4 PHYSICAL HABITAT ALTERATION (INCLUDING WATERWAY 13)

5 According to NMFS (2009), the stressor associated with physical habitat alteration 6 specifically addressed the issue of return flows and attraction of anadromous salmonids 7 into the Yuba Goldfields through Waterway 13. Various efforts have been undertaken to 8 prevent anadromous salmonids from entering the Goldfields via Waterway 13. In May 9 2005, heavy rains and subsequent flooding breached the structure at the east (upstream 10 facing) end. Subsequently, funded by USFWS, the earthen "plug" was replaced with a 11 "leaky-dike" barrier intended to serve as an exclusion device for upstream migrating adult 12 salmonids (AFRP 2010). The Corps does not have any operations or maintenance 13 responsibilities for the earthen "plug" and Waterway 13, nor has it issued any permits or 14 licenses for it. Nonetheless, until a more permanent solution is implemented, ongoing 15 issues associated with attraction of upstream migrating adult salmonids into Waterway 13 16 are considered to remain a stressor to spring-run Chinook salmon. For additional 17 information on Waterway 13, see Chapter 5 – Environmental Baseline.

18 In addition to Waterway 13 issues, physical habitat alternation stressors include Lake 19 Wildwood operations and resultant Deer Creek flow fluctuations (according to the 20 SWRCB's Revised Decision 1644, Lake Wildwood is operated by the Lake Wildwood Association — a gated community in Penn Valley, California). This stressor refers to the 21 22 potential for stranding or isolation events to occur in Deer Creek, near its confluence with 23 the lower Yuba River. Observational evidence suggests that, in the past, adult Chinook 24 salmon entered Deer Creek during relatively high flow periods, presumably for holding 25 or spawning purposes, only to subsequently become stranded in the creek when flows 26 receded due to changes in Lake Wildwood operations. Stranding may delay or prevent 27 adult Chinook salmon from spawning, or cause decreased spawning success due to 28 increased energy expenditure or stress due to delayed spawning (CALFED and 29 YCWA 2005).

1 The Sierra Streams Institute (SSI) is in the process of implementing the Deer Creek 2 Spawning Bed Enhancement Project, which is located on a tributary to the lower Yuba 3 River. From September 4-7, 2012, 250 tons of spawning gravel (~180 cubic yards) was 4 placed in the creek. Chinook salmon redd surveys were conducted after the initial 5 placement to document the number and characteristics of salmon redds created in Deer 6 Creek during the 2012 spawning season. On November 27, 2012, more than 51 salmon 7 redds were observed in Deer Creek, compared to 15 redds in 2011, and 9 redds in 2003 8 (SSI 2013). Approximately 75% of spawning activity during 2012 occurred in the newly 9 created spawning areas, with the remaining spawning activity occurring in locations 10 where spawning was observed in 2011. Gravel transport also was monitored to 11 understand the effects of higher stream flows on gravel movement, and to evaluate 12 transport of spawning gravels in Deer Creek. Tracer gravel surveys were conducted 13 during February, March, and April 2013. Based on these and other visual observations of 14 substrate deposition in Deer Creek, SSI (2013) report that it is likely that some of the 15 placed gravels remain in Deer Creek providing spawning habitat, and that some of the 16 gravels were mobilized downstream into the Yuba River to provide habitat for anadromous salmonids. To supplement existing available spawning habitat, SSI planned 17 18 to place an additional 250 tons of spawning gravel in Deer Creek from September 19 3-13, 2013.

Physical habitat alteration stressors also address habitat complexity and diversity. The
concepts of habitat complexity and diversity pertinent to the lower Yuba River were
described by CALFED and YCWA (2005), as discussed below.

Habitat complexity and diversity refer to the quality of instream physical habitatincluding, but not necessarily limited to, the following physical habitat characteristics:

Escape cover
 Feeding cover
 Allochthonous material contribution
 Alternating point-bar sequences
 Pool-to-riffle ratios
 Sinuosity
 Instream object cover
 Overhanging riparian vegetation

1 The physical structure of rivers plays a significant role in determining the suitability of 2 aquatic habitats for juvenile salmonids, as well as for other organisms upon which 3 salmonids depend for food. These structural elements are created through complex 4 interactions among natural geomorphic features, the power of flowing water, sediment 5 delivery and movement, and riparian vegetation, which provides bank stability and inputs 6 of large woody debris (Spence et al. 1996). The geomorphic conditions caused by hydraulic and dredge mining since the mid-1800s, and the construction of Englebright 7 Dam, which affects the transport of nutrients, fine and course sediments and, to a lesser 8 9 degree, woody material from upstream sources to the lower river, continue to limit 10 habitat complexity and diversity in the lower Yuba River.

LWM creates both micro- and macro-habitat heterogeneity by forming pools, back eddies and side channels and by creating channel sinuosity and hydraulic complexity. This habitat complexity provides juvenile salmonids numerous refugia from predators and water velocity, and provides efficient locations from which to feed. LWM also functions to retain coarse sediments and organic matter in addition to providing substrate for numerous aquatic invertebrates (Spence et al. 1996).

In the lower Yuba River, mature riparian vegetation is scattered intermittently, leaving much of the banks devoid of LWM and unshaded – affecting components that are essential to the health and survival of the freshwater lifestages of salmonids (NMFS 2002). Although the ability of the lower Yuba River to support riparian vegetation has been substantially reduced by the historic impacts from mining activities, the dynamic nature of the river channel results in periodic creation of high-value shaded riverine aquatic (SRA) cover for fish and wildlife (Beak 1989).

Other important components of habitat structure at the micro-scale include large boulders, coarse substrate, undercut banks and overhanging vegetation. These habitat elements offer juvenile salmonids concealment from predators, shelter from fast current, feeding stations and nutrient inputs. At the macro-scale, streams and rivers with high channel sinuosity, multiple channels and sloughs, beaver impoundments or backwaters typically provide high-quality rearing and refugia habitats (Spence et al. 1996). The
lower Yuba River can be generally characterized as lacking an abundance of
 such features.

3 Loss of Riparian Habitat and Instream Cover

4 **RIPARIAN VEGETATION**

5 As stated in CALFED and YCWA (2005), riparian vegetation, an important habitat 6 component for anadromous fish, is known to provide: (1) bank stabilization and sediment 7 load reduction; (2) shade that results in lower instream water temperatures; (3) overhead 8 cover; (4) streamside habitat for aquatic and terrestrial insects, which are important food 9 sources for rearing juvenile fishes; (5) a source of instream cover in the form of woody 10 material; and (6) allochthonous nutrient input.

11 SRA cover generally occurs in the lower Yuba River as scattered, short strips of low-12 growing woody species (e.g., Salix sp.) adjacent to the shoreline. Beak (1989) reported 13 that the most extensive and continuous segments of SRA cover occur along bars where 14 [then] recent channel migrations or avulsions had cut new channels through relatively 15 large, dense stands of riparian vegetation. SRA cover consists of instream object cover 16 and overhanging cover. Instream object cover provides structure, which promotes 17 hydraulic complexity, diversity and microhabitats for juvenile salmonids, as well as 18 escape cover from predators. The extent and quality of suitable rearing habitat and cover, 19 including SRA, generally has a strong effect on juvenile salmonid production in rivers 20 (Healey 1991 as cited in CALFED and YCWA 2005).

21 Since completion of New Bullards Bar Reservoir, the riparian community (in the lower 22 Yuba River) has expanded under summer and fall streamflow conditions that have 23 generally been higher than those that previously occurred (SWRCB 2003). However, the 24 riparian habitat is not pristine. NMFS (2005b) reports ... "The deposition of hydraulic 25 mining debris, subsequent dredge mining, and loss/confinement of the active river 26 corridor and floodplain of the lower Yuba River which started in the mid-1800's and 27 continues to a lesser extent today, has eliminated much of the riparian vegetation along 28 the lower Yuba River. In addition, the large quantities of cobble and gravel that 29 remained generally provided poor conditions for re-establishment and growth of riparian

1 vegetation. Construction of Englebright Dam also inhibited regeneration of riparian 2 vegetation by preventing the transport of any new fine sediment, woody debris, and 3 nutrients from upstream sources to the lower river. Subsequently, mature riparian 4 vegetation is sparse and intermittent along the lower Yuba River, leaving much of the 5 bank areas unshaded and lacking in large woody debris. This loss of riparian cover has 6 greatly diminished the value of the habitat in this area."

7 Where hydrologic conditions are supportive, riparian and wetland vegetative 8 communities are found adjacent to the lower Yuba River and on the river sides of 9 retaining levees. These communities are dynamic and have changed over the years as the 10 river meanders. The plant communities along the river are a combination of remnant 11 Central Valley riparian forests, foothill oak/pine woodlands, agricultural grasslands, and 12 orchards (Beak 1989).

According to CALFED and YCWA (2005), the lower Yuba River, especially in the vicinity of Daguerre Point Dam and the Yuba Goldfields, is largely devoid of sufficient riparian vegetation to derive the benefits (to anadromous salmonids) discussed above (**Figure 4-4**).

17 In 2012, YCWA conducted a riparian habitat study in the Yuba River from Englebright 18 Dam to the confluence with the Feather River (see Technical Memorandum 6-2 in 19 YCWA 2013). Field efforts included descriptive observations of woody and riparian 20 vegetation, cottonwood inventory and coring, and a large woody material (LWM) survey. 21 The study was performed by establishing eight LWM study sites and seven riparian 22 habitat study sites. One LWM study site was established within each of eight distinct 23 reaches (i.e., Marysville, Hallwood, Daguerre Point Dam, Dry Creek, Parks Bar, 24 Timbuctoo Bend, Narrows, and Englebright Dam). Riparian habitat sites were 25 established in the same locations as the LWM study sites, with the exception of the 26 Marysville study site. Riparian information regarding the Marysville Reach was 27 developed, but no analysis was performed because of backwater effects of the Feather 28 River.

29 The RMT contracted Watershed Sciences Inc. to use existing LiDAR to produce a map of 30 riparian vegetation stands by type. The resulting data was subject to a field validation



Figure 4-4. Vegetation communities in the lower Yuba River vicinity (Source: CALFED and
 YCWA 2005).

and briefly summarized in WSI (2010) and the data were also utilized in YCWA's
Riparian Study Technical Memorandum 6-2 (YCWA 2013).

Based on field observations, YCWA (2013) reported that all reaches supported woody
species in various lifestages - mature trees, recruits, and seedlings were observed within
all reaches. Where individuals or groups of trees were less vigorous, beaver (*Castor canadensis*) activity was the main cause, although some trees in the Marysville Reach
appeared to be damaged by human camping.

The structure and composition of riparian vegetation was largely associated with four landforms. Cobble-dominated banks primarily supported bands of willow shrubs with scattered hardwood trees. Areas with saturated soils or sands supported the most complex riparian areas and tended to be associated with backwater ponds. Scarps and levees supported lines of mature cottonwood and other hardwood species, typically with a simple understory of Himalayan blackberry or blue elderberry shrubs. Bedrock dominated reaches had limited riparian complexity and supported mostly willow shrubs
 and cottonwoods (YCWA 2013).

3 Based on analysis of the mapping data, RMT (2013) reported that the majority of the 4 woody species present in the river valley include, in order of most to least number of 5 individuals: various willow species (Salix sp. and Cephalanthus occidentalis); Fremont 6 cottonwood (Populus fremontii) (i.e., cottonwoods); blue elderberry (Sambucus nigra 7 ssp. caerulea); black walnut (Juglans hindsii); Western sycamore (Platanus racemosa); 8 Oregon ash (Fraxinus latifolia); white alder (Alnus rhombifolia); tree of heaven 9 (Ailanthus altissima); and grey pine (Pinus sabiniana). Willow species could not be 10 differentiated by species using remote sensing information. Willow on the lower Yuba 11 River are dominated by dusky sandbar willow (Salix melanopsis) and narrow leaf willow 12 (Salix exigua), and relative dominance of the two species shifts respectively in the 13 downstream direction (WSI 2010). Other species occurring are arundo willow (Salix 14 lasiolepsis), Goodings willow (Salix goodingii) and red willow (Salix laevigata). 15 Goodings and red willow comprise 6.4% of the willow according to a limited field 16 validation survey (WSI 2010).

17 Cottonwoods are one of the most abundant woody species in the study area, and the most 18 likely source of locally-derived large instream woody material due to rapid growth rates 19 and size of individual stems commonly exceeding 2 feet in diameter and 50 feet in length. 20 Cottonwoods exist in all life stages including as mature trees, recruits, or saplings, and as 21 seedlings. Cottonwoods are more abundant in downstream areas of the study area 22 relative to upstream. Cottonwoods are distributed laterally across the valley floor. Of the 23 estimated 18,540 cottonwood individuals/stands, 12% are within the bankfull channel 24 (flows of 5,000 cfs or less), and 39% are within the floodway inundation zone (flows 25 between 5,000 and 21,100 cfs). However, recruitment patterns of cottonwood have not 26 been analyzed with respect to time or with any more detail regarding channel location 27 (YCWA 2013).

A total of 97 cottonwood trees were cored to estimate age. Age estimates ranged from 11 to 87 years. The cottonwood tree age analysis resulted in age estimates that place the year of establishment for trees in a range of years from ± 7 to 16 years, which is too wide to allow for linking the establishment of trees to any year's specific conditions
 (YCWA 2013).

YCWA conducted a historical aerial photograph analysis to describe changes over time to total vegetation delineated within the valley walls, riparian vegetation delineated within 50 feet of the active river channel,² and channel alignment (see Technical Memorandum 6-2 in YCWA 2013). To determine the cumulative change over time³ in total vegetative cover and riparian vegetation cover for the Marysville, Timbuctoo Bend, Narrows, and Englebright Dam study sites, YCWA compared the aerial photographs from 1937 and 2010.

10 Cumulative changes in vegetative cover in the Englebright Dam and Narrows study sites 11 decreased. For the remaining study sites, including Marysville, Hallwood, Daguerre 12 Point Dam, Dry Creek, Parks Bar, and Timbuctoo Bend study sites, the cumulative 13 change in vegetative cover increased. The least amount of vegetation change over time 14 was observed in the Englebright Dam, Narrows and Marysville sites. The Dry Creek, 15 Daguerre Point Dam and Hallwood sites had the greatest vegetated area, and YCWA 16 identified those sites as the most dynamic (i.e., both decreased in vegetative cover 17 through 1970 and then increased through 2010).

18 Cumulative changes in riparian vegetation cover in the Englebright Dam and Narrows 19 study sites decreased with very little detectable change for the Narrows study site. For 20 the remaining study sites, the cumulative change in riparian vegetation cover increased. 21 The observed changes for the Englebright Dam, Narrows and Marysville study sites were 22 very small. For the Dry Creek and Parks Bar study sites, the greatest changes were 23 observed, with dramatic increases in riparian vegetation cover. The magnitude of change 24 of riparian vegetation between photoset years (in a stepwise comparison) was greater than 25 that seen in the cumulative riparian vegetation cover change.

² Total vegetation is inclusive of riparian vegetation.

³ Cumulative change describes the changes to observable area for either total vegetation or riparian vegetation from the earliest photo date to the most recent photo date.

1 INSTREAM WOODY MATERIAL

Instream woody material provides escape cover and relief from high current velocities for
juvenile salmonids and other fishes. LWM also contributes to the contribution of
invertebrate food sources, and micro-habitat complexity for juvenile salmonids (NMFS
2007). Snorkeling observations in the lower Yuba River have indicated that juvenile
Chinook salmon had a strong preference for near-shore habitats with instream woody
material (JSA 1992).

8 There is currently a lack of consensus regarding the amount of instream woody material 9 occuring in the lower Yuba River (Corps 2012d). It has been suggested (CALFED and 10 YCWA 2005) that the presence of Englebright Dam has resulted in decreased recruitment 11 of LWM to the lower Yuba River, although no surveys or studies were cited to support 12 these statements. Some woody material may not reach the lower Yuba River due to 13 collecting on the shoreline and sinking in Englebright Reservoir (Corps 2012d). 14 However, Englebright Dam does not functionally block woody material from reaching 15 the lower Yuba River because there is no woody material removal program implemented 16 for Englebright Reservoir, and accumulated woody material therefore spills over the dam 17 during uncontrolled flood events (R. Olsen, Corps, pers. comm. 2011, as cited in 18 Corps 2012d).

19 About 8.7 miles of the lower Yuba River downstream of Englebright Dam, distributed 20 among study sites per reach, were surveyed and evaluated for pieces of wood (YCWA 21 2013). The number of pieces of wood was relatively similar above and below Daguerre 22 Point Dam (i.e., about 5,100 and 5,750 pieces, respectively). Woody material was 23 generally found in bands of willow (Salix sp.) shrubs near the wetted edge, dispersed 24 across open cobble bars, and stranded above normal high-flow indicators. Most of the 25 woody material was diffuse and located on floodplains and high floodplains, with only 26 about a guarter of the material in heavy concentrations (YCWA 2013).

Most (77-96%) pieces of wood found in each reach were smaller than 25 feet in length and smaller than 24 inches in diameter, which is the definition of large woody material (LWM). These pieces would be typically floated by flood flows and trapped within willows and alders above the 21,100 cfs line, which is defined as the flow delineating the
 floodway boundary (YCWA 2013).

Instream woody material was not evenly distributed throughout the reaches. For the
smaller size classes (i.e., shorter than 50 feet, less than 24 inches in diameter), the
greatest abundance of pieces was found in the Hallwood or Daguerre Point Dam reaches,
with lower abundances above and below these reaches (YCWA 2013).

The largest size classes of LWM (i.e., longer than 50 feet and greater than 24 inches in 7 8 diameter) were rare or uncommon (i.e., fewer than 20 pieces total) with no discernible 9 distribution. Pieces of this larger size class were counted as "key pieces", as were any 10 pieces exceeding 25 inches in diameter and 25 feet in length and showing any 11 morphological influence (e.g., trapping sediment or altering flow patterns). A total of 15 12 key pieces of LWM were found in all study sites, including six in the Marysville study 13 site. Few of the key pieces were found in the active channel or exhibiting channel 14 forming processes (YCWA 2013).

15 Loss of Natural River Morphology and Function

According to NMFS (2009), "Loss of Natural River Morphology and Function" is the result of river channelization and confinement, which leads to a decrease in riverine habitat complexity, and thus, a decrease in the quantity and quality of juvenile rearing habitat. Additionally, this primary stressor category includes the effect that dams have on the aquatic invertebrate species composition and distribution, which may have an effect on the quality and quantity of food resources available to juvenile salmonids.

According to NMFS (2009), attenuated peak flows and controlled flow regimes have
altered the lower Yuba River's geomorphology and have affected the natural meandering
of the river downstream of Englebright Dam.

As reported by RMT (2013), preliminary evaluation of available data collected to date related to Yuba River fluvial geomorphology indicates that the Yuba River downstream of Englebright Dam has complex river morphological characteristics. Evaluation of the morphological units in the Yuba River as part of the spatial structure analyses indicates that, in general, the sequence and organization of morphological units is non-random, indicating that the channel has been self-sustaining of sufficient duration to establish an
 ordered spatial structure (RMT 2013).

The Yuba River downstream of Englebright Dam exhibits lateral variability in its formprocess associations (RMT 2013). In the Yuba River, morphological unit organization highlights the complexity of the channel geomorphology, as well as the complex and diverse suite of morphological units. The complexity in the landforms creates diversity in the flow hydraulics which, in turn, contributes to a diversity of habitat types available for all riverine lifestages of anadromous salmonids in the Yuba River downstream of Englebright Dam (RMT 2013).

10 In the lower Yuba River, anadromous salmonids spawn in mean substrate sizes ranging 11 from about 50 to 150 mm, and most of the lower Yuba River from Englebright Dam to 12 the confluence with the Feather River is characterized by average substrate particle sizes 13 The exceptions are sand/silt areas near the within this size range (RMT 2013). 14 confluence with the Feather River, and the boulder/bedrock regions in the upper sections 15 of Timbuctoo Bend and most of the Englebright Dam Reach. However, gravel 16 augmentation funded by the Corps in the Englebright Dam Reach over the past several 17 years has spurred spawning activity and Chinook salmon redd construction in this reach. 18 The net result is an increase in the spatial distribution of spawning habitat availability in 19 the river, particularly for early spawning (presumably spring-run) Chinook salmon 20 (RMT 2013).

21 Loss of Floodplain Habitat

22 NMFS (2009) listed the loss of floodplain habitat in the lower Yuba River as one of the 23 key stressors affecting anadromous salmonids (including spring-run Chinook salmon). 24 NMFS (2009) stated ..."Historically, the Yuba River was connected to vast floodplains 25 and included a complex network of channels, backwaters and woody material. The legacy 26 of hydraulic and dredger mining is still evident on the lower Yuba River where, for much 27 of the river, dredger piles confine the river to an unnaturally narrow channel. The 28 consequences of this unusual and artificial geomorphic condition include reduced 29 floodplain and riparian habitat and resultant limitations in fish habitat, particularly for 30 rearing juvenile salmonids."

NMFS (2009) further stated that in the lower Yuba River, controlled flows and decreases in peak flows has reduced the frequency of floodplain inundation resulting in a separation of the river channel from its natural floodplain. Within the Yuba Goldfields area (RM 8– 14), confinement of the river by massive deposits of cobble and gravel derived from hydraulic and dredge mining activities resulted in a relatively simple river corridor dominated by a single main channel and large cobble-dominated bars, with little riparian and floodplain habitat (DWR and PG&E 2010).

8 Loss of off-channel habitats such as floodplains, riparian, and wetland habitats has 9 substantially reduced the productive capacity of the Central Valley for many native fish 10 and wildlife species, and evidence is growing that such habitats were once of major 11 importance for the growth and survival of juvenile salmon (Moyle 2002). Recent 12 observations on the lower Yuba River indicate that remnant side channels and associated 13 riparian vegetation play a similar role by providing flood refugia, protection from 14 predators, and abundant food for young salmonids and other native fishes. These habitats 15 also promote extended rearing and expression of the stream-type rearing characteristic of 16 spring-run Chinook salmon (DWR and PG&E 2010).

17 As reported by RMT (2013), despite some flow regulation, the channel and floodplain in 18 the lower Yuba River are highly connected, with floods spilling out onto the floodplain more frequently than commonly occurs for unregulated semiarid rivers. Some locations 19 20 exhibit overbank flow well below 5,000 cfs, while others require somewhat more than 21 that. In any given year, there is an 82% chance the river will spill out of its bankfull 22 channel and a 40% chance that the floodway will be fully inundated. These results 23 demonstrate that floodplain inundation occurs with a relatively high frequency in the 24 lower Yuba River compared to other Central Valley streams which, in turn, contributes to 25 a diversity in habitats available for anadromous salmonids (RMT 2013).

26 RMT (2013) conducted a flood-frequency analysis of the annual peak discharges 27 recorded at the USGS stream gage near Marysville (#11421000) that showed average 28 annual return periods of 1.25 years and 2.5 years for the bankfull and flood discharges, 29 respectively. Bankfull flows for similar rivers are generally assumed to occur with return 30 periods of 1.5-2 years. The fact that the lower Yuba River is less than this implies that the channel is naturally undersized relative to generalized expectations and flows spill
 into the floodplain at a more frequent rate (RMT 2013).

3 <u>ENTRAINMENT</u>

4 According to NMFS (2009), entrainment of juvenile salmonids remains a stressor in the 5 lower Yuba River. Entrainment represents a suite of potential negative impacts to 6 juvenile fish that may occur while, or after, the fish encounter a diversion facility in 7 operation. For instance, entrainment impacts may include the non-volitional recruitment 8 of juveniles past a diversion facility and/or screening structure, or impingement upon 9 diversion screens and physical damage to fish caused by diversion activities. It has been 10 suggested that as juvenile salmonids pass Daguerre Point Dam, physical injury may occur 11 as they pass over the dam or through its fish ladders (SWRI 2002).

12 Water diversions in the lower Yuba River generally begin in the early spring and extend 13 through the fall. As a result, potential threats to juvenile salmonids occur at the 14 Hallwood-Cordua and South Yuba/Brophy diversions (NMFS 2009). The relatively 15 recent fish screen constructed at the Hallwood-Cordua diversion is considered a notable 16 improvement over the previous design, and is believed to reduce the amount of fry and juvenile entrainment at the diversion. The new diversion fish screen is believed to reduce 17 18 loss rates of emigrating fall-run Chinook salmon at this location. However, predation 19 losses of emigrating fry and juvenile fall-run Chinook salmon may remain a limiting 20 factor at this location. In addition, the configuration of the current return pipe and flows 21 though the pipe may also be a limiting factor (CALFED and YCWA 2005).

22 As previously described, the South Yuba/Brophy system diverts water through an 23 excavated channel from the south bank of the lower Yuba River in the vicinity of 24 Daguerre Point Dam. The water is then subsequently diverted through a porous rock dike 25 that is intended to exclude fish. The current design of this rock structure does not meet 26 current NMFS or CDFW juvenile fish screen criteria (SWRI 2002), and additional issues 27 regarding predation in the diversion channel and the rate of water bypassing the rock 28 gabion and returning to the lower Yuba River through the diversion channel have been 29 raised as potential stressors.

1 **PREDATION**

2 Predation can occur in three forms: (1) natural; (2) predation resulting from a relative 3 increase in predator habitat and opportunity near major structures and diversions; and (3) 4 predation resulting from minimal escape cover and habitat complexity for prey species 5 (CALFED and YCWA 2005). For the purpose of stressor identification in this BA, 6 predation includes the predation associated with increases in predator habitat and 7 predation opportunities for piscivorous species created by major structures and 8 diversions, and predation resulting from limited amounts of prev escape cover in the 9 lower Yuba River.

10 The extent of predation on juvenile Chinook salmon in the lower Yuba River is not well 11 documented (NMFS 2009). Although predation is a natural component of salmonid 12 ecology, the rate of predation of salmonids in the lower Yuba River has potentially 13 increased through the introduction of non-native predatory species such as striped bass 14 (*Morone saxatilis*), largemouth bass (*Micropterus salmoides*) and American shad (*Alosa* 15 *sapidissima*) and through the alteration of natural flow regimes and the development of 16 structures that attract predators (NMFS 2009).

17 Predatory fish are known to congregate around structures in the water including dams, 18 diversions and bridges, where their foraging efficiency is improved by shadows, 19 turbulence and boundary edges (CDFG 1998). Thus, juvenile salmonids can also be 20 adversely affected by Daguerre Point Dam on their downstream migration. Daguerre 21 Point Dam creates a large plunge pool at its base, which provides ambush habitat for 22 predatory fish in an area where emigrating juvenile salmonids may be disoriented after 23 plunging over the face of the dam into the deep pool below (NMFS 2002). The 24 introduced predatory striped bass and American shad have been observed in this pool 25 (CALFED and YCWA 2005). In addition to introduced predatory species, several native 26 fish species also prey on juvenile salmonids in the lower Yuba River, including 27 Sacramento pikeminnow, hardhead and large juvenile and adult rainbow trout/steelhead 28 (CALFED and YCWA 2005). It has been suggested that the rate of predation of juvenile 29 salmonids passing over dams in general, and Daguerre Point Dam in particular, may be unnaturally high (NMFS 2007), although specific studies addressing this suggestion have
 not been conducted.

3 In addition to the suggestion of increased rates of predation resulting from disorientation 4 of juveniles passing over Daguerre Point Dam into the downstream plunge pool, it also 5 has been suggested that unnaturally high predation rates may also occur in the diversion 6 channel associated with the South Yuba/Brophy diversion (NMFS 2007). Other 7 structure-related predation issues include the potential for increased rates of predation of 8 juvenile salmonids: (1) in the entryway of the Hallwood-Cordua diversion canal upstream 9 of the fish screen; and (2) at the point of return of fish from the bypass pipe of the 10 Hallwood-Cordua diversion canal into the lower Yuba River.

11 HATCHERY EFFECTS

12 Although no fish hatcheries are located on the lower Yuba River, and the river continues 13 to support a persistent population of spring-run Chinook salmon that spawn downstream 14 of Englebright Dam, the genetic integrity of the fish expressing the phenotypic 15 characteristics of spring-run Chinook salmon is presently uncertain. CDFG (1998) 16 suggested that spring-run Chinook salmon populations may be hybridized to some degree 17 with fall-run Chinook salmon due to lack of spatial separation of spawning habitat. Also, 18 the observation of adipose fin clips on adult Chinook salmon passing upstream through 19 the VAKI system at Daguerre Point Dam during the spring demonstrates that hatchery 20 straying into the lower Yuba River has and continues to occur, most likely from the 21 FRFH (NMFS 2009; RMT 2013).

22 FEATHER RIVER FISH HATCHERY GENETIC CONSIDERATIONS

Spring-run Chinook salmon from the FRFH were planted in the lower Yuba River during (CDFG 1991). In addition, it is possible that some hatchery-reared juvenile Chinook salmon from the FRFH may move into the lower Yuba River in search of rearing habitat. Some competition for resources with naturally spawned spring-run Chinook salmon could occur as a result (YCWA et al. 2007). The remainder of this discussion pertains to hatchery effects associated with the straying of adult Chinook salmon into the lower Yuba River. The FRFH is the only hatchery in the Central Valley that currently produces spring-run
 Chinook salmon. The FRFH was constructed in 1967 to compensate for anadromous
 salmonid spawning habitat lost with construction of the Oroville Dam. The FRFH has a
 goal of releasing 2,000,000 spring-run Chinook salmon smolts annually (DWR 2004c).

From 1962 to 1966, spring-run Chinook salmon were trapped and trucked above Oroville
Dam. Beginning in 1967, spring-run Chinook salmon were collected for artificial
propagation at FRFH as the construction of Oroville Dam was completed. The program
is funded by the DWR and managed by CDFW (NMFS 2004).

9 The program was founded with local native stock collected at the FRFH. Early attempts 10 to over-summer spring-run at the hatchery resulted in high mortality and the decision to 11 allow the run to hold in the river until September 1. Prior to 2004, FRFH hatchery staff 12 differentiated spring-run Chinook salmon from fall-run Chinook salmon by opening the 13 ladder to the hatchery on September 1 (NMFS 2009). Those fish ascending the ladder 14 from September 1 through September 15 were assumed to be spring-run Chinook salmon 15 while those ascending the ladder after September 15 were assumed to be fall-run (Kastner 16 2003 as cited in NMFS 2009). This practice led to considerable hybridization between 17 spring- and fall-run Chinook salmon (DWR 2004c). Since 2004, the FRFH fish ladder 18 remains open during the spring months, closing on June 30, and those fish ascending the 19 ladder are marked with an external flov tag and returned to the river. This practice allows 20 FRFH staff to identify those previously marked fish as spring-run when they re-enter the 21 ladder in September. Only floy-tagged fish are spawned with floy-tagged fish in the 22 month of September. No other fish are spawned during this time, as part of an effort to 23 prevent hybridization with fall-run, and to introduce a temporal separation between 24 stocks in the hatchery. During the FRFH spring-run spawning season, all heads from 25 adipose fin-clipped fish are taken and sent to CDFW's laboratory in Santa Rosa for tag 26 extraction and decoding. The tag information will be used to test the hypothesis that 27 early spring-run spawners will produce progeny that maintain that run fidelity.

Regardless of recent improved FRFH practices, previous practices appear to have
resulted in hybridization between "spring-run" and "fall-run" Chinook salmon. The
following discussion was taken from Garza et al. (2008).

1 Evaluation of the FRFH "spring-run" stock found that it is genetically most similar to the 2 FRFH fall-run stock, as indicated both by clustering on the phylogeographic trees and by 3 comparison of the [standardized variance in allele frequencies between the sample years] 4 (F_{ST}) values, and is nested within the fall-run group of populations in all analyses (Garza 5 et al. 2008). F_{ST} values between the FRFH "spring-run" and naturally-spawned spring-6 run are in the low end of the range of values for fall-run populations to spring-run 7 populations, but not the lowest. In addition, they are the essentially the same as those of 8 FRFH fall-run to spring-run populations. This demonstrates convincingly that the FRFH 9 "spring-run" stock is dominated by fall-run ancestry. However, Garza et al. (2008) also 10 found very slight, but significant, differentiation between the two FRFH stocks, which is 11 concordant with the results of Hedgecock et al. (unpublished study as cited in Garza et al. 12 2008) on these stocks. In addition, Garza et al. (2008) found a strong signal of linkage 13 (gametic phase) disequilbrium, absent in all other population samples, in the FRFH 14 "spring-run" stock. Garza et al. (2008) interpreted this as evidence that the FRFH 15 "spring" run retains remnants of the phenotype and ancestry of the Feather River spring-16 run Chinook salmon that existed prior to the dam and hatchery (as opposed to representing a hatchery selection-created and maintained phenotypic variant), but that has 17 18 been heavily introgressed by fall-run Chinook salmon through some combination of 19 hatchery practices and natural hybridization, induced by habitat concentration due to lack 20 of access to spring-run Chinook salmon habitat above the dam. This suggests that it may 21 be possible to preserve some additional component of the ancestral Central Valley spring-22 run Chinook salmon genomic variation through careful management of this stock that can 23 contribute to the recovery of the ESA-listed Central Valley spring-run Chinook salmon 24 ESU, although it will not be possible to reconstitute a "pure" spring-run stock from 25 these fish.

The FRFH spring-run Chinook salmon population is part of the Central Valley spring-run Chinook salmon ESU (70 FR 37160). At the time of issuance of the final rule regarding the listing status of the Central Valley ESU of spring-run Chinook salmon, NMFS (70 FR 37160) recognized that naturally spawning spring-run Chinook in the Feather River are genetically similar to the FRFH spring-run Chinook stock, and that the hatchery stock shows evidence of introgression with Central Valley fall-run Chinook salmon. NMFS also stated that FRFH stock should be included in the ESU because the FRFH spring-run
Chinook salmon stock may play an important role in the recovery of spring-run Chinook
salmon in the Feather River Basin, as efforts progress to restore natural spring-run
populations in the Feather and Yuba Rivers (70 FR 37160).

Although the FRFH spring-run Chinook salmon population is part of the Central Valley
spring-run Chinook salmon ESU, concern has been expressed that straying of FRFH fish
into the lower Yuba River may represent an adverse impact due to the potential influence
of previous hatchery management practices on the genetic integrity of FRFH spring-run
Chinook salmon.

10 STRAYING INTO THE LOWER YUBA RIVER

The RMT (2013) reported that substantially higher amounts of straying of adipose finclipped Chinook salmon into the lower Yuba River occur than that which was previously believed. Although no quantitative analyses or data were presented, NMFS (2007) stated that some hatchery fish stray into the lower Yuba River and that these fish likely come from the FRFH.

16 Some information indicating the extent to which adipose-clipped Chinook salmon 17 originating from the FRFH return to the lower Yuba River is available from coded wire 18 tag analysis. During the October through December 2010 carcass survey period in the 19 lower Yuba River, the RMT collected heads from fresh Chinook salmon carcasses with 20 adipose fin clips, and sent the heads to the CDFW coded wire tag (CWT) interpretive 21 center. In April of 2011, the results of the interpretation of the CWTs became available. 22 Of the 333 Chinook salmon heads sent to the CDFW interpretive center, 11 did not 23 contain a CWT, 8 were fall-run Chinook salmon from the Coleman National Fish 24 Hatchery, 2 were from the RST captured and tagged juveniles in the lower Yuba River, 1 25 was a naturally-spawned fall-run Chinook salmon from the Feather River, 1 was a fall-26 run Chinook salmon from the Mokelumne River Hatchery, and 310 were Chinook 27 salmon from the FRFH (234 spring-run and 76 fall-run Chinook salmon). Thus, for all 28 CWT hatchery-origin fish returning to the Yuba River from out-of-basin sources, 97% 29 were from the FRFH. However, this information does not indicate the percentage of 30 hatchery contribution from the FRFH to the phenotypic spring-run Chinook salmon run in the lower Yuba River, because, among other reasons, all of these heads were collected
during the fall and represent a mixture of phenotypic spring- and fall-run Chinook salmon
spawning in the lower Yuba River (RMT 2013).

4 Additional information that can be used to assess the amount of straying of FRFH 5 Chinook salmon into the lower Yuba River is provided from VAKI Riverwatcher data 6 collected from 2004 through 2011 (RMT 2013). The estimated numbers of adipose fin-7 clipped spring-run Chinook salmon that passed upstream of Daguerre Point Dam from 8 2004 through 2011 that were derived from the VAKI Riverwatcher data are an indicator 9 of the minimum number of Chinook salmon of hatchery origin (most likely of FRFH 10 origin) that strayed into the lower Yuba River. The following discussion of adipose fin-11 clipped spring-run Chinook salmon is from RMT (2013). Discussion of the procedure 12 utilized by the RMT (2013) to first differentiate phenotypic spring-run from phenotypic 13 fall-run Chinook salmon is provided in Section 4.2.7.2, below.

14 Because the VAKI Riverwatcher systems located at both the north and south ladder of 15 Daguerre Point Dam can record both silhouettes and electronic images of each fish 16 passage event, the systems were able to differentiate Chinook salmon with adipose fins 17 clipped or absent from Chinook salmon with their adipose fins intact. Thus, annual series 18 of daily counts of Chinook salmon with adipose fins clipped (i.e., ad-clipped fish) and 19 with adipose fins intact (i.e., not ad-clipped fish) that passed upstream of Daguerre Point 20 Dam from March 1, 2004 through February 29, 2012 were obtained. The estimated 21 numbers of spring-run Chinook salmon of hatchery (i.e., ad-clipped fish) and potentially 22 non-hatchery origin (i.e., not ad-clipped fish) passing upstream of Daguerre Point Dam 23 for the last eight years of available VAKI Riverwatcher data are presented in Table 4-4.

24 Relationships between Spring-run Chinook Salmon Straying into the lower Yuba River 25 AND Attraction Flows and Water Temperatures

As reported by RMT (2013), to evaluate the influence of "attraction" flows and water temperatures on the straying of adipose fin-clipped adult phenotypic spring-run Chinook salmon into the lower Yuba River, variables related to flows and water temperatures in the lower Yuba River and the lower Feather River were developed and statistically related to the weekly proportions of adipose fin-clipped phenotypic spring-run Chinook

1 Table 4-4. Estimated numbers of Chinook salmon, ad-clipped and not ad-clipped 2 phenotypic spring-run Chinook salmon that passed upstream of Daguerre Point Dam 3 annually from 2004 through 2011 (Source: RMT 2013).

| Year | Demarcation Date | Chinook Salmon Passage Upstream of Daguerre Point Dam | | | | |
|------|---------------------|---|---------------------------|------------|----------------|--------------|
| | | All Chinook | Spring-run Chinook Salmon | | | |
| | | Salmon | Total | Ad-Clipped | Not Ad-Clipped | % Ad-Clipped |
| 2004 | 8/1/04 | 5,927 | 738 | 72 | 666 | 10 |
| 2005 | 8/24/05 | 11,374 | 3,592 | 676 | 2,916 | 19 |
| 2006 | 9/6/06 | 5,203 | 1,326 | 81 | 1,245 | 6 |
| 2007 | 9/4/07 | 1,394 | 372 | 38 | 334 | 10 |
| 2008 | 8/10/08 | 2,533 | 521 | 15 | 506 | 3 |
| 2009 | 7/9/09 | 5,378 | 723 | 213 | 510 | 29 |
| 2010 | 7/6/10 | 6,469 | 2,886 | 1,774 | 1,112 | 61 |
| 2011 | 9/7/11 | 7,785 | 1,159 | 323 | 836 | 28 |

salmon (relative to all spring-run Chinook salmon) passing upstream of Daguerre Point
Dam during each of the 8 years when annual VAKI Riverwatcher counts at Daguerre
Point Dam are available. Details of this analytical evaluation are provided in RMT
(2013).

Results of the RMT (2013) analysis suggest that there is a moderately strong ($R^2=0.72$) 9 10 and highly significant (P < 0.000001) relationship between the percentage of adipose finclipped spring-run Chinook salmon contribution to the weekly spring-run Chinook 11 12 salmon total counts at Daguerre Point Dam and the attraction flow and water temperature 13 indices four weeks prior. The attraction flow index explained 20.4% of the data 14 variability, the attraction water temperature index explained 27.5% of the variability, and the interaction term explained 24.4% of the variability in the proportion of adipose fin-15 16 clipped phenotypic spring-run Chinook salmon passing Daguerre Point Dam weekly 17 (RMT 2013). Figure 4-5 displays the 3-D response surface produced by the fitted 18 logistic model.

19 The analysis described above showed that an estimated 72% of the variation in the 20 proportion of adipose fin-clipped phenotypic spring-run Chinook salmon passing 21 upstream of Daguerre Point Dam can be accounted for by the ratio of lower Yuba River 22 flow relative to lower Feather River flow, and the ratio of lower Yuba River water 23 temperature relative to lower Feather River water temperature, four weeks prior to the

4



Figure 4-5. Relationship of the weekly percentage of adipose fin-clipped contribution to the weekly phenotypic spring-run Chinook salmon count at Daguerre Point Dam as function of the weekly attraction flow and water temperature indices calculated four weeks prior to the week of passage at Daguerre Point Dam (Source: RMT 2013).

time of passage at Daguerre Point Dam. In other words, the higher the Yuba River flows
relative to Feather River flows, combined with the lower the Yuba River water
temperatures relative to Feather River water temperatures, the higher the percentage of
fin-clipped Chinook salmon passing upstream of Daguerre Point Dam four weeks later
(RMT 2013).

As described in RMT (2013), the acoustically-tagged phenotypic spring-run Chinook salmon spent variable and extended periods of time holding below Daguerre Point Dam after being tagged and prior to passing upstream of Daguerre Point Dam, with a range of 0 to 116 days. Based on all 67 acoustically-tagged spring-run Chinook salmon that passed upstream of Daguerre Point Dam, the average holding time before passing upstream of Daguerre Point Dam was about 50 days. For the phenotypic acousticallytagged spring-run Chinook salmon that passed upstream of Daguerre Point Dam by the

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annual spring-run Chinook salmon demarcation date for each year, the average holding periods before passing upstream of Daguerre Point Dam were approximately 51, 41, and 57 days during 2009, 2010 and 2011, respectively. Therefore, it would be expected that attraction of adipose fin-clipped fish to the lower Yuba River associated with flows and water temperatures in the lower Yuba River relative to the lower Feather River would occur at least several weeks prior to passage of phenotypic spring-run Chinook salmon upstream of Daguerre Point Dam (RMT 2013).

8 While the variation in the proportion of adipose fin-clipped phenotypic spring-run 9 Chinook salmon passing Daguerre Point Dam was best explained with ratios of flows and 10 water temperatures in the lower Yuba and Feather rivers four weeks prior to passage at 11 Daguerre Point Dam, the acoustically-tagged individuals exhibited a somewhat longer 12 duration of holding on average. However, due to the relatively small sample size of 13 acoustically-tagged spring-run Chinook salmon passing upstream of Daguerre Point Dam 14 (N=67), the short duration of the study, and based on the highly variable holding duration 15 (i.e., 0-116 days), the average holding time calculated for the acoustically-tagged spring-16 run Chinook salmon is considered to be a general approximation of holding duration 17 downstream of Daguerre Point Dam (RMT 2013). Therefore, consideration of holding 18 duration downstream of Daguerre Point Dam supports the observation that the ratios of 19 flows and water temperatures in the lower Yuba River relative to the lower Feather River 20 four weeks prior to passage of spring-run Chinook salmon at Daguerre Point Dam may be 21 influencing the attraction of adipose fin-clipped spring-run Chinook salmon of FRFH-22 origin into the lower Yuba River (RMT 2013).

23 LOWER YUBA RIVER GENETIC CONSIDERATIONS

Spring-run Chinook salmon historically acquired and maintained genetic integrity through reproductive (spatial-temporal) isolation from other Central Valley Chinook salmon runs. However, construction of dams has prevented access to headwater areas and much of this historical reproductive isolation has been compromised, resulting in intermixed life history traits in many remaining habitats (YCWA 2010).

Between 1900 and 1941, debris dams constructed on the lower Yuba River by theCalifornia Debris Commission to retain hydraulic mining debris, now owned and

operated by the Corps, completely or partially blocked the migration of Chinook salmon and steelhead to historic spawning and rearing habitats (CDFG 1991a; Wooster and Wickwire 1970; Yoshiyama et al. 1996). Englebright Dam (constructed in 1941) completely blocks spawning runs of Chinook salmon and steelhead, and is the upstream limit of fish migration. Fry (1961) reported that a small spring-run Chinook salmon population historically occurred in the lower Yuba River, but the run virtually disappeared by 1959.

8 Since the completion of New Bullards Bar Reservoir in 1970 by YCWA, higher, colder 9 flows in the lower Yuba River have improved conditions for over-summering and 10 spawning of spring-run Chinook salmon in the lower Yuba River (YCWA et al. 2007). 11 As of 1991, a remnant spring-run Chinook salmon population reportedly persisted in the 12 lower Yuba River downstream of Englebright Dam maintained by fish produced in the 13 lower Yuba River, fish straying from the Feather River, or fish previously and 14 infrequently stocked from the FRFH (CDFG 1991). In the 1990s, relatively small 15 numbers of Chinook salmon that exhibit spring-run phenotypic characteristics were 16 reported to have been observed in the lower Yuba River (CDFG 1998). Although precise 17 escapement estimates are not available, the USFWS testified at the 1992 SWRCB lower 18 Yuba River hearing that "...a population of about 1,000 adult spring-run Chinook 19 salmon now exists in the lower Yuba River" (San Francisco Bay RWQCB 2006).

If spring-run Chinook salmon were extirpated from the lower Yuba River in 1959 (Fry 1961) and, as reported by CDFG (1991), a population of spring-run Chinook salmon became reestablished since the 1970s due to improved habitat conditions and fish straying from the Feather River or stocked and straying from the FRFH, then it is likely that spring-run Chinook salmon on the lower Yuba river do not represent a "pure" ancestral genome.

There also is concern that the existing spring-run Chinook salmon population has interbred with fall-run Chinook salmon and, as a result, it is a hybrid species and not a true spring-run species (Corps 2001). In addition to the effects of hatchery straying, an additional issue regarding the genetic integrity of phenotypic spring-run Chinook salmon in the lower Yuba River pertains to the loss or reduction of reproductive isolation.

1 Spring-run Chinook salmon acquired and maintained genetic integrity through spatial-2 temporal isolation from other Central Valley Chinook salmon runs. Historically, spring-3 run Chinook salmon were temporally isolated from winter-run, and largely isolated in 4 both time and space from the fall-run. Much of this historical spatial-temporal integrity 5 has broken down, resulting in intermixed life history traits in many remaining habitats. 6 Consequently, the present self-sustaining, persistent populations of spring-run Chinook 7 salmon in the upper Sacramento, lower Yuba, and lower Feather rivers may be 8 hybridized to some degree with fall-run Chinook salmon (YCWA et. al 2007).

9 Englebright Dam is a complete migration barrier to anadromous fish, precluding 10 migration of Chinook salmon to historical holding and spawning areas upstream of the 11 dam. Consequently, both fall-run and spring-run Chinook salmon are restricted to areas 12 below the dam. Because the spawn timing overlaps between the two runs and they 13 potentially interbreed, genetic swamping of the relatively smaller numbers of spring-run 14 Chinook salmon by more abundant fall-run fish could occur (DWR and PG&E 2010).

15 The presence of Englebright Dam has necessitated that spring-run Chinook salmon 16 spawn in areas that were believed to formerly represent fall-run Chinook salmon 17 spawning areas. Although the lower Yuba River continues to support a persistent 18 population of spring-run Chinook salmon that now are restricted to spawning downstream of Englebright Dam, the genetic integrity of the fish expressing the 19 20 phenotypic characteristics of spring-run Chinook salmon is presently uncertain. For 21 example, CDFG (1998) suggests that spring-run populations may be hybridized to some 22 degree with fall-run populations due to lack of spatial separation of spawning habitat for 23 the two runs of Chinook salmon in the lower Yuba River.

In the report titled *Salmonid Hatchery Inventory and Effects Evaluation* (NMFS 2004), through an analysis of Yuba River Chinook salmon tissues, NMFS genetically linked the spring-run and fall-run populations, which exhibit a merged run timing similar to that found in the Feather River.

In conclusion, available information indicates that: (1) the phenotypic spring-run Chinook salmon in the lower Yuba River actually represents hybridization between spring- and fall-run Chinook salmon in the lower Yuba River, and hybridization with

1 Feather River stocks including the FRFH spring-run Chinook salmon stock, which itself 2 represents a hybridization between Feather River fall- and spring-run Chinook salmon 3 populations; and (2) straying from FRFH origin "spring-run" Chinook salmon into the 4 lower Yuba River occurs, and that this rate of straying is associated with the relative 5 proportion of lower Yuba River flows and water temperatures to lower Feather River 6 flows and water temperatures ("attraction flows and water temperatures"); and (3) the 7 FRFH spring-run Chinook salmon is included in the ESU, in part because of the 8 important role this stock may play in the recovery of spring-run Chinook salmon in the 9 Feather River Basin, including the Yuba River (70 FR 37160). Although straying of 10 FRFH "spring-run" Chinook salmon into the lower Yuba River has oftentimes been 11 suggested to represent an adverse impact on lower Yuba River spring-run Chinook 12 salmon stocks, it is questionable whether the phenotypic spring-run Chinook salmon in 13 the lower Yuba River represents an independent population. The RMT (2013) recently 14 reported that data obtained through the course of implementing the RMT's M&E 15 Program demonstrate that phenotypically "spring-running" Chinook salmon in the lower 16 Yuba River do not represent an independent population – rather, they represent an 17 introgressive hybridization of the larger Feather-Yuba river regional population.

18 JUVENILE STRANDING AND REDD DEWATERING

In the California State Water Resources Control Board's (SWRCB) 2001 Decision (D)-1644, the SWRCB directed YCWA to submit a plan that described the scope and duration of future flow fluctuation studies to verify that Chinook salmon and steelhead redds are being adequately protected from dewatering with implementation of D-1644 criteria (YCWA 1992). The monitoring and evaluation plan contained the following objectives (JSA 2003):

- Determine the potential magnitude of redd dewatering in relation to the timing
 and magnitude of flow fluctuations and reductions
- Determine the potential magnitude of fry stranding in relation to the timing,
 magnitude, and rate of flow fluctuations and reductions
- Evaluate the effectiveness of the D-1644 flow fluctuation and reduction criteria
 in protecting redds and fry

Recommend additional measures to protect redds and fry from flow fluctuations and reductions if warranted

3 The studies combined habitat mapping, field surveys, and information on the timing and 4 distribution of fry rearing in the Yuba River to evaluate the effectiveness of D-1644 flow 5 fluctuation and reduction criteria in protecting Chinook salmon and steelhead fry. Two 6 studies were conducted and summarized in the 2007 and 2008 Lower Yuba River Redd 7 Dewatering and Fry Stranding Annual Report (JSA 2008) to the SWRCB, and results from an additional study were reported in a progress report in 2010 (ICF Jones & Stokes 8 9 2010). A preliminary draft report providing the results of all survey activities conducted 10 during 2007 through 2011 was produced in 2012 (ICF Jones & Stokes 2012), although 11 additional evaluation and reporting of the data is ongoing.

The first *Lower Yuba River Redd Dewatering and Fry Stranding Study* was conducted in April 2007 to evaluate bar and off-channel stranding of juvenile salmonids associated with a flow reduction of 1,300-900 cfs at Smartsville at a ramping rate of 100 cfs per hour. Bar stranding was again evaluated in June with a temporary flow reduction of 1,600-1,300 cfs at a rate of 100 cfs per hour. Snorkel surveys were conducted between Rose Bar, located ~2.5 miles downstream of Englebright Dam, and the Highway 20 Bridge, located ~5.7 miles downstream of Englebright Dam.

19 During the April 5, 2007 drawdown, field crews observed eight stranded salmon fry in 20 the interstitial spaces of substrates on bar slopes (perpendicular to shoreline) ranging 21 from 0.5 to 5.5% in slope. No stranded fish were observed during surveys conducted on 22 June 18, 2007. The presence of both juvenile Chinook salmon and O. mykiss were 23 confirmed in shallow, near-shore areas adjacent to the study sites, suggesting that the risk of bar stranding is greatly reduced by June. Following the April 5, 2007 flow reductions, 24 25 juvenile salmon were found in 16 of the 24 disconnected off-channel sites (ICF Jones & 26 Stokes 2012). Most of the fish that had become isolated in off-channel sites were 30-50 27 mm fry. Out of the 16 sites where isolation of fry was observed, 70% of the fish were 28 found in the four largest sites, which accounted for nearly 60% of the total wetted area 29 that had become disconnected from the main river. According to ICF Jones & Stokes 30 (2012), these four sites were unique in that they were all associated with man-made

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features within or adjacent to the main river channel (e.g., diversion channels, ponds and
 bridge piers).

An updated *Lower Yuba River Redd Dewatering and Fry Stranding Study* was subsequently conducted from May 29, 2008 through June 4, 2008 with a scheduled flow reduction on June 1, 2008. A total of seven stranded trout fry ranging between 30-35 mm were observed in the interstitial spaces of substrates on bar slopes ranging from 2.0 to 5.7% in slope.

Juvenile salmon were found isolated in seven of the 12 off-channel sites that had become 8 9 disconnected from the main river by the June 1, 2008 event. One site accounted for only 10 about 7% of the total wetted area that had been disconnected from the main river, but 11 nearly 80% of the total number of juvenile salmon that had been isolated by the June 1, 12 2008 event. A total of 13 steelhead fry were found isolated in 2 of the 12 off-channel 13 sites that had become disconnected from the main river by the June 1, 2008 event. 14 Nearly all of these fish were 30-50 mm fry that had been isolated in a single backwater 15 pool adjacent to the main river in the Timbuctoo Reach (ICF Jones & Stokes 2012).

JSA (2008) suggested that the preliminary findings indicated that juvenile *O. mykiss* fry may be less vulnerable to off-channel stranding than juvenile Chinook salmon because of their more restricted distribution and inability to access off-channel areas under late spring flow conditions. Long-term monitoring of several isolated off-channel sites confirmed that some sites can support juvenile salmonids for long periods and even produce favorable summer rearing conditions.

A 2010 study was conducted from June 21, 2010 through July 1, 2010, with a scheduled flow reduction between June 28 and June 30 from approximately 4,000 cfs to 3,200 cfs as measured at the Smartsville Gage. As reported by ICF Jones & Stokes (2010), fish stranding surveys were conducted on June 21, 22, and 23 to identify potential stranding areas and document habitat conditions and fish presence before the flow reduction, and were repeated on June 29, June 30, and July 1 to document the incidence of fish stranding and habitat conditions after the flow reduction.

After the June flow reduction, a total of six juvenile salmon and 46 juvenile trout was observed in seven of the 26 off-channel sites that had become fully or nearly disconnected (≤0.1 foot deep) from the main river. Most of the stranded fish were
juvenile trout 30-70 mm in length that had become isolated in five off-channel sites
above Daguerre Point Dam. Below Daguerre Point Dam, observations of stranded fish
were limited to six juvenile salmon and two juvenile trout at two study sites
(ICF Jones & Stokes 2010).

Hydrologic and operating conditions in January and February 2011 provided the first
opportunity to evaluate the effect of a winter flow reduction on the incidence of bar
stranding. A series of three successive flow reductions were evaluated. Following a 3week period of relatively stable flows, Englebright Dam releases were reduced from
3,000-2,600 cfs on January 31, 2,600-2,200 cfs on February 7, and 2,200-2,000 cfs on
February 11.

The first event was a 400-cfs flow reduction (3,000–2,600 cfs) conducted from 8:00 AM to 10:00 AM at a target rate of 200 cfs per hour on January 31, 2011. This event resulted in a 2.1–2.5 inch drop in water surface elevation and a rate of change of 0.6–0.8 inch per hour at the three study sites. Field crews searched a total of 764 square feet of dewatered shoreline and found a total of 20 stranded salmon fry (30-40 mm long) and six stranded steelhead (50-90 mm long) (ICF Jones & Stokes 2012).

During the second event on February 7, 2011, flows were again reduced by 400 cfs (2,600–2,200 cfs) from 8:00 AM to 10:00 AM, but at a target rate of 100 cfs per hour. This event resulted in a 1.8–2.1 inch drop in water surface elevation and a rate of change of 0.4–0.5 inch per hour at the three study sites. Field crews searched a total of 560 square feet of dewatered shoreline and found a total of 10 stranded salmon fry (30-40 mm long) and no steelhead (ICF Jones & Stokes 2012).

During the third event on February 11, 2011, flows were reduced by 200 cfs (2,200– 2,000 cfs) from 2:00 AM to 4:00 AM at a target rate of 100 cfs per hour. This event resulted in a 0.8–1.3 inch drop in water surface elevation and a rate of change of 0.4–0.7 inch per hour at the three study sites. Field crews searched a total of 248 square feet of dewatered shoreline and found a total of four stranded salmon fry (30-40 mm long) and no steelhead (ICF Jones & Stokes 2012).

1 4.2.7 Viability of Central Valley Spring-run Chinook Salmon

2 The "Viable Salmonid Population" (VSP) concept was developed by McElhany et al. 3 (2000) to facilitate establishment of Evolutionarily Significant Unit (ESU)-level delisting 4 goals and to assist in recovery planning by identifying key parameters related to 5 population viability. Four key parameters were identified by McElhany et al. (2000) as the key to evaluating population viability status: (1) abundance; (2) productivity; (3) 6 7 diversity; and (4) spatial structure. McElhany et al. (2000) interchangeably use the term 8 population growth rate (i.e., productivity over the entire life cycle) and productivity. 9 Good et al. (2007) used the term productivity when describing this VSP parameter, which 10 also is the term used for this parameter in this BA. The following discussion regarding 11 the four population viability population parameters was taken directly from 12 NMFS (2009).

Abundance is an important determinant of risk, both by itself and in relationship to other factors (McElhany et al. 2000). Small populations are at a greater risk for extinction than larger populations because risks that affect the population dynamics operate differently on small populations than in large populations. A variety of risks are associated with the dynamics of small populations, including directional effects (i.e., density dependence compensatory and depensatory), and random effects (i.e., demographic stochasticity, environmental stochasticity, and catastrophic events).

20 The parameter of productivity and factors that affect productivity provide information on 21 how well a population is "performing" in the habitats it occupies during the life cycle 22 (McElhany et al. 2000). Productivity and related attributes are indicators of a 23 population's performance in response to its environment and environmental change and 24 variability. Intrinsic productivity (the maximum production expected for a population 25 sufficiently small relative to its resource supply not to experience density dependence), 26 the intensity of density dependence, and stage-specific productivity (productivity realized 27 over a particular part of the life cycle) are useful in assessing productivity 28 of a population.

Diversity refers to the distribution of traits within and among populations, and these traits
 range in scale from DNA sequence variation at single genes to complex life-history traits

1 (McElhany et al. 2000). Traits can be completely genetic or vary due to a combination of 2 genetics and environmental factors. Diversity in traits is an important parameter because: 3 (1) diversity allows a species to use a wide array of environments; (2) diversity protects a 4 species against short-term spatial and temporal changes in its environment; and (3) 5 genetic diversity provides the raw material for surviving long-term environmental 6 changes (McElhany et al. 2000). Some of the varying traits include run timing, spawning 7 timing, age structure, outmigration timing, etc. Straying and gene flow strongly influence 8 patterns of diversity within and among populations (McElhany et al. 2000).

9 Spatial structure reflects how abundance is distributed among available or potentially 10 available habitats, and how it can affect overall extinction risk and evolutionary processes 11 that may alter a population's ability to respond to environmental change. A population's 12 spatial structure encompasses the geographic distribution of that population, as well as 13 the processes that generate or affect that distribution (McElhany et al. 2000). A 14 population's spatial structure depends fundamentally on habitat quality, spatial 15 configuration, and dynamics as well as the dispersal characteristics of individuals in the 16 population. Potentially suitable but unused habitat is an indication of the potential for 17 population growth.

18 **4.2.7.1 ESU**

19 To determine the current viability of the spring-run Chinook salmon ESU, NMFS 20 (2009a) used the historical population structure of spring-run Chinook salmon presented 21 in Lindley et al. (2007) and the concept of VSP for evaluating populations described by 22 McElhany et al. (2000). Lindley et al. (2004) identified 26 historical populations within 23 the spring-run ESU; 19 were independent populations, and 7 were dependent populations. 24 Of the 19 independent populations of spring-run that occurred historically, only three 25 remain, in Deer, Mill, and Butte creeks. Extant dependent populations occur in Battle, 26 Antelope, Big Chico, Clear, Beegum, and Thomes creeks, as well as in the Yuba River, 27 the Feather River below Oroville Dam, and in the mainstem Sacramento River below 28 Keswick Dam (NMFS 2009a).

Lindley et al. (2007) provide criteria to assess the level of risk of extinction of Pacificsalmonids based on population size, recent population decline, occurrences of

catastrophes within the last 10 years that could cause sudden shifts from a low risk state
to a higher one, and the impacts of hatchery influence. Although these criteria were
developed for application to specific populations, insight to the viability of the spring-run
Chinook salmon ESU can be obtained by examining population trends within the context
of these criteria.

6 VIABLE SALMONID POPULATION (VSP) PARAMETERS AND APPLICATION

7 ABUNDANCE

According to NMFS (2009a), spring-run Chinook salmon in the Central Valley declined drastically in the mid- to late 1980s before stabilizing at very low levels in the early to mid-1990s. Since the late 1990s, there does not appear to be a trend in basin-wide abundance (NMFS 2009a). Since NMFS presented these data, additional abundance estimates are available for the spring-run Chinook salmon ESU.

13 Central Valley-wide spring-run Chinook salmon abundance estimates are available 14 through GrandTab (CDFW 2013). Since 1983, in-river estimates for the lower Feather 15 River have not been included in the system-wide estimates, although FRFH estimates are 16 provided separately. Additionally, spring-run Chinook salmon are not estimated in 17 GrandTab for the lower Yuba River, and all lower Yuba River Chinook salmon 18 escapement estimates are reported as fall-run Chinook salmon. For the Sacramento River 19 system (not including the FRFH or the lower Yuba River) since 1983, spring-run 20 Chinook salmon run size estimates have ranged from a high of 24,903 in 1998 to a low of 21 1,404 in 1993. For the past five years (2008 - 2012), the abundance of in-river spawning 22 Central Valley spring-run Chinook salmon has steadily declined from a high of 11,927 in 23 2008 to a low of 2,962 in 2010, before increasing to 5,439 in 2011 and 18,511 in 2012.

The spring-run Chinook salmon run size estimate for the Sacramento River system (not including the FRFH or the lower Yuba River) over the past three consecutive years for which data are available averaged 8,971 fish (i.e., 2,962 fish in 2010, 5,439 fish in 2011, and 18,511 fish in 2012).

1 **Ρ***RODUCTIVITY*

The spring-run Chinook salmon run size estimate for the Sacramento River system (not including the FRFH or the lower Yuba River) over the past three consecutive years totaled 26,912 fish, thereby exceeding both the minimum total escapement value of 2,500 (Lindley et al. 2007), as well as the mean value of 833 fish per year identified by NMFS (2011a).

7 From 1983 through 2012, the annual contribution of spring-run Chinook salmon from the 8 FRFH to the total annual run size in the Sacramento River system has ranged from a high 9 of 76.9% (4,672 fish) in 1993 to a low of 5.6% (1,433 fish) in 1986. As an indicator of 10 the FRFH influence on spring-run Chinook salmon in the Sacramento River system, the 11 average annual percent contribution of FRFH spring-run Chinook salmon relative to the 12 total annual run in the Sacramento River system was 31.2% over the entire 30-year 13 period (1983-2012), and was 20.7% over the last 10 years (2003-2012). The percent 14 contribution of FRFH to the total population of Central Valley spring-run Chinook 15 salmon does not represent straying *per se*. The guidelines presented in Figure 1 in 16 Lindley et al. (2007) present extinction risk levels corresponding to different amount, 17 duration and source of hatchery strays, taking into consideration whether hatchery strays 18 are from within the ESU, the diversity group, and from a "best management practices" 19 hatchery. These criteria indicate a high extinction risk if hatchery straying represents 20 more than 20% hatchery contribution for one generation or more than 10% for four 21 generations from a hatchery within a given diversity group, or more than 50% hatchery 22 contribution for one generation or more than 15% for four generations from a best 23 management practices hatchery within a given diversity group. Although not technically 24 representing straying, the average contribution of spring-run Chinook salmon from the 25 FRFH to the total annual run size in the Sacramento River system has been 26.4% over 26 the most recent generation, 21.6% over the two most recent generations, 19.8% over the 27 three most recent generations, and 19.9% over the four most recent generations assuming 28 a three-year life cycle. According to NMFS (2011a), recent anomalous conditions in the 29 coastal ocean, along with consecutive dry years affecting inland freshwater conditions, 30 have contributed to statewide escapement declines.

1 SPATIAL STRUCTURE

2 Lindley et al. (2007) indicated that of the 19 independent populations of spring-run that 3 occurred historically, only three (Butte, Mill, and Deer creeks) remain, and their current 4 distribution makes the spring-run ESU vulnerable to catastrophic disturbance (e.g., 5 disease outbreaks, toxic spills, or volcanic eruptions). Butte, Mill, and Deer Creeks all occur in the same biogeographic region (diversity group), whereas historically, 6 7 independent spring-run populations were distributed throughout the Central Valley among at least three diversity groups (i.e., the Basalt and Porous Lava Diversity Group, 8 9 the Northern Sierra Nevada Diversity Group, and the Southern Sierra Nevada Diversity 10 Group). In addition, dependent spring-run populations historically persisted in the 11 Northwestern California Diversity Group (Lindley et al. 2004). Currently, there are 12 dependent populations of spring-run Chinook salmon in the Big Chico, Antelope, Clear, 13 Thomes, Battle, and Beegum creeks, and in the Sacramento, Feather, and Yuba rivers 14 (Lindley et al. 2007).

Spring-run Chinook salmon have been reported more frequently in several upper Central Valley creeks, but the sustainability of these runs is still unknown (NMFS 2004). In 2004, NMFS reported that Butte Creek spring-run cohorts had recently utilized all available habitat in the creek, so the population cannot expand further. It is unknown if individuals have opportunistically migrated to other systems. The spatial structure of the Central Valley spring-run Chinook salmon ESU has been reduced with the extirpation of all San Joaquin River Basin spring-run populations (NMFS 2004).

22 Diversity

As discussed in NMFS (2009a), diversity, both genetic and behavioral, provides a species the opportunity to track environmental changes. As a species' abundance decreases, and spatial structure of the ESU is reduced, a species has less flexibility to track changes in the environment. Spring-run Chinook salmon reserve some genetic and behavioral variation in that in any given year, at least two cohorts are in the marine environment and, therefore, are not exposed to the same environmental stressors as their freshwater cohorts (NMFS 2009a).

1 Genetic analysis of natural and hatchery spring-run Chinook salmon stocks in the Central 2 Valley reveal that the southern Cascades spring-run population complex has retained its 3 genetic integrity (NMFS 2004). However, although spring-run produced at the FRFH are 4 part of the spring-run Chinook salmon ESU (70 FR 37160, June 28, 2005), they 5 compromise the genetic diversity of naturally-spawned spring-run Chinook salmon (NMFS 2009a). The spring-run hatchery stock introgressed with the fall-run hatchery 6 7 stock, and both are genetically linked with the natural populations in the Feather River 8 (NMFS 2004). The FRFH program has affected the diversity of the Central Valley 9 spring-run Chinook salmon and, together with the loss of the San Joaquin River Basin 10 spring-run populations, the diversity of the Central Valley spring-run Chinook salmon 11 ESU has been reduced (NMFS 2004).

12 SUMMARY OF THE VIABILITY OF THE CENTRAL VALLEY SPRING-RUN CHINOOK SALMON ESU

13 According to NMFS (2005a), threats from hatchery production, climatic variation, 14 predation, and water diversions persist. Because the Central Valley spring-run Chinook 15 salmon ESU is confined to relatively few remaining streams and continues to display 16 broad fluctuations in abundance, high quality critical habitat containing spawning sites 17 with adequate water and substrate conditions, or rearing sites with adequate floodplain 18 connectivity, cover, and water conditions (i.e., key primary constituent elements of 19 critical habitat that contribute to its conservation value) is considered to be limited and 20 the population is at a moderate risk of extinction.

According to NMFS (2009a), spring-run Chinook salmon fail the representation and redundancy rule for ESU viability, because the current distribution of independent populations has been severely constricted to only one of their former geographic diversity groups. NMFS (2009a) concluded that the Central Valley spring-run Chinook salmon ESU is at moderate risk of extinction in 100 years.

In 2011, NMFS completed a 5-year status review of the Central Valley spring-run Chinook salmon ESU. According to NMFS (2011b), new information for the Central Valley spring-run Chinook salmon ESU suggests an increase in extinction risk. With a few exceptions, Central Valley spring-run Chinook salmon escapements has declined over the past 10 years, in particular since 2006 (NMFS 2011b). Overall, the recent 1 declines have been significant but not severe enough to qualify as a catastrophe under the 2 criteria of Lindley et al. (2007). On the positive side, spring-run Chinook salmon appear 3 to be repopulating Battle Creek, home to a historical independent population in the Basalt 4 and Porous Lava diversity group that was extirpated for many decades. Similarly, the 5 spring-run Chinook salmon population in Clear Creek has been increasing, although 6 Lindley et al. (2004) classified this population as a dependent population, and thus it is not expected to exceed the low-risk population size threshold of 2,500 fish (i.e., annual 7 8 spawning run size of about 833 fish).

9 The status of the Central Valley spring-run Chinook salmon ESU has probably 10 deteriorated on balance since the 2005 status review and Lindley et al.'s (2007) 11 assessment, with two of the three extant independent populations of spring-run Chinook 12 salmon slipping from low or moderate extinction risk to high extinction risk (NMFS 13 2011b). Butte Creek remains at low risk, although it is on the verge of moving towards 14 high risk (NMFS 2011b). By contrast, spring-run Chinook salmon in Battle and Clear 15 creeks have increased in abundance over the last decade, reaching levels of abundance 16 that place these populations at moderate extinction risk (NMFS 2011b).

17 In summary, NMFS (2011b) states that the status of the Central Valley spring-run 18 Chinook salmon ESU has probably deteriorated since the 2005 status review. From 19 2007-2009, the Central Valley experienced drought conditions and low river and stream 20 discharges, which are generally associated with lower survival of Chinook salmon 21 (NMFS 2011b). There is a possibility that with the recent cessation of the drought and a 22 return to more typical patterns of upwelling and sea-surface temperatures that declining 23 trends in abundance may reverse in the near future (NMFS 2011b). According to NMFS 24 (2011b), improvements in the status of two spring-run Chinook salmon populations in the 25 Central Valley are not sufficient to warrant a downgrading of the ESU extinction risk, 26 and the degradation in status of three formerly low- or moderate-risk independent 27 populations is cause for concern. New information available since Good et al. (2005) 28 indicates an increased extinction risk (NMFS 2011b).

1 4.2.7.2 Lower Yuba River

As previously discussed, the VSP concept was developed by McElhany et al. (2000) in order to facilitate establishment of ESU-level delisting goals and to assist in recovery planning by identifying key parameters related to population viability. The four parameters established by McElhany et al. (2000) included abundance, productivity, spatial structure and genetic and life-history diversity, although McElhany et al. (2000) did not provide quantitative criteria that would allow assessment of whether particular populations or ESUs/DPSs are viable.

9 Lindley et al. (2007) characterized the spring-run Chinook salmon population in the 10 lower Yuba River as data deficient, and therefore did not characterize its viability. In 11 2007, there was limited information on the current population size of spring-run Chinook 12 salmon in the lower Yuba River, although NMFS (2009) stated that ongoing monitoring 13 is providing additional information.

14 ABUNDANCE AND PRODUCTIVITY

15 RUN DIFFERENTIATION (SPRING-RUN VS. FALL-RUN CHINOOK SALMON)

Prior to application of VSP performance indicators or the extinction risk criteria, it is
necessary to differentiate between annually returning spring-run and fall-run Chinook
salmon in the lower Yuba River.

However, as reported by RMT (2013), there is no discernible genetic differentiation available to determine spring-run Chinook salmon, only phenotypic differentiation. The phenotypic expression is often obscure, requiring application of advanced statistical techniques to VAKI Riverwatcher and other datasets in order to identify the phenotypic differences in run timing. The following discussion of differentiating phenotypic springrun from phenotypic fall-run Chinook salmon in the lower Yuba River is generally taken from RMT (2013).

Infrared-imaging technology has been used to monitor fish passage at Daguerre Point
Dam in the lower Yuba River since 2003 using VAKI Riverwatcher systems to document
specific observations used to address VSP parameters of adult abundance and diversity.
The VAKI Riverwatcher infrared systems produced by VAKI Aquaculture Systems Ltd.,

of Iceland, provided a tool for monitoring fish passage year-round. The VAKI
 Riverwatcher system records both silhouettes and electronic images of each fish passage
 event in both of the Daguerre Point Dam fish ladders. By capturing silhouettes and
 images, fish passage can be accurately monitored even under turbid conditions.

5 The VAKI Riverwatcher systems located at both the north and south ladder of Daguerre
6 Point Dam were able to record and identify the timing and magnitude of passage for
7 Chinook salmon at Daguerre Point Dam during most temporal periods of a given year.

8 Prior to applying any analysis of temporal modalities to the 8 annual time series of 9 Chinook salmon daily VAKI counts, the annual daily count series at each ladder were 10 adjusted to account for days when the VAKI Riverwatcher systems were not fully 11 operational. The procedure used to obtain complete annual daily count series of Chinook 12 salmon migrating upstream of Daguerre Point Dam is provided in RMT (2013).

The daily time series of Chinook salmon moving upstream of Daguerre Point Dam resulting from the previous step were further analyzed and temporal modalities were explored to differentiate spring-run from fall-run Chinook salmon each year. For a full description of the run differentiation process, see RMT (2013).

17 Figure 4-6 and Figure 4-7 display the daily number of Chinook salmon that passed 18 upstream of Daguerre Point Dam during the 2004 to the 2011 biological years (March 1 19 through February 28) and the fitted generalized logistic functions describing the 20 distributions of spring-run and fall-run Chinook salmon resulting from the application of 21 the annually variable temporal demarcation procedure. Finally, Table 4-5 summarizes 22 the total number of spring-run and fall-run Chinook salmon estimated to have passed 23 upstream of Daguerre Point Dam annually, and the estimated annual percentage of 24 spring-run Chinook salmon relative to all Chinook salmon each year.



Figure 4-6. Daily number of Chinook salmon passing upstream of Daguerre Point Dam during the 2004 to 2007 biological years. Bars indicate the VAKI Riverwatcher daily counts and lines indicate the predicted daily distributions of spring-run (blue line) and fall-run (orange line) Chinook salmon based on the fitting of two generalized logistic functions to the data. The demarcation date differentiating the two runs of Chinook salmon is indicated for each year (Source: RMT 2013).



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Figure 4-7. Daily number of Chinook salmon passing upstream of Daguerre Point Dam during the 2008 to 2011 biological years. Bars indicate the VAKI Riverwatcher daily counts and lines indicate the predicted daily distributions of spring-run (blue line) and fall-run (orange line) Chinook salmon based on the fitting of two generalized logistic functions to the data. The demarcation date differentiating the two runs of Chinook salmon is indicated for each year. (Source: RMT 2013)
1 Table 4-5. Annual number of spring-run and fall-run Chinook salmon estimated to have 2 passed upstream of Daguerre Point Dam, and the estimated annual percentage of spring-3 run Chinook salmon relative to all Chinook salmon each year. (Source: RMT 2013)

| Run | | Biological Year | | | | | | | | | | | | | |
|---------------------------|-------|-----------------|-------|-------|-------|-------|-------|-------|--|--|--|--|--|--|--|
| | 2004 | 2005 | 2006 | 2007 | 2008 | 2009 | 2010 | 2011 | | | | | | | |
| Spring-run Chinook Salmon | 738 | 3,592 | 1,326 | 372 | 521 | 723 | 2,886 | 1,159 | | | | | | | |
| | 12.5% | 31.6% | 25.5% | 26.7% | 20.6% | 13.4% | 44.6% | 14.9% | | | | | | | |
| Fall-run Chinook Salmon | 5,189 | 7,782 | 3,877 | 1,022 | 2,012 | 4,655 | 3,583 | 6,626 | | | | | | | |
| | 87.5% | 68.4% | 74.5% | 73.3% | 79.4% | 86.6% | 55.4% | 85.1% | | | | | | | |

4

5 ANNUAL ABUNDANCE OF SPRING-RUN CHINOOK SALMON

For the period (2004-2011) during which VAKI Riverwatcher data are available, the
annual number of spring-run Chinook salmon estimated to have passed upstream of
Daguerre Point Dam ranged from 372 in 2007 to 3,592 in 2005, with an average of 1,415
(RMT 2013). The abundance of spring-run Chinook salmon during the past two years
has been substantially higher than the three years prior (RMT 2013).

As previously described by NMFS (2011a), populations with a low risk of extinction 11 12 (less than 5% chance of extinction in 100 years) are those with a minimum total 13 escapement of 2,500 spawners in 3 consecutive years (mean of 833 fish per year). For 14 the last three consecutive years, an estimated total of 4,768 spring-run Chinook salmon 15 have passed upstream of Daguerre Point Dam, with an average of 1,589 fish per year 16 (RMT 2013). However, as further discussed below, the annual abundances of phenotypic 17 spring-run Chinook salmon in the lower Yuba River are strongly influenced by hatchery 18 fish (RMT 2013).

19 TRENDS IN THE ANNUAL ABUNDANCE OF SPRING-RUN CHINOOK SALMON

The statistical approach recommended by Lindley et al. (2007) was followed by RMT (2013) to examine whether the abundance of lower Yuba River spring-run Chinook salmon exhibited a statistically significant linear trend over time during the eight most recent years for which VAKI Riverwatcher data are available. The natural logarithms of the abundance estimates of lower Yuba River spring-run Chinook salmon for the eight most recent years (2004-2011) were linearly regressed against time (year) using a simple
least-squares approach (RMT 2013). The estimated slope of the resulting line is a
measure of the average rate of change of the abundance in the population over time.

4 Figure 4-8 displays the antilogarithmic transformation of the estimated annual number of 5 spring-run Chinook salmon passing upstream of Daguerre Point Dam from 2004-2011 6 (RMT 2013). Figure 4-8 demonstrates that the abundance of spring-run Chinook salmon 7 in the lower Yuba River has exhibited a very slight increase over the eight years examined. However, the coefficient of determination is very weak ($r^2 = 0.0005$) and the 8 slope is not statistically significantly different from zero (P = 0.96), indicating that the 9 10 positive trend is not significant (RMT 2013). The relationship indicates that the 11 phenotypic spring-run Chinook salmon annual abundance over this time period is stable, 12 and is not exhibiting a significant declining trend (RMT 2013). These abundance and 13 trend considerations would correspond to low extinction risk according to NMFS criteria 14 (Lindley et al. 2007). However, the RMT (2013) questions the applicability of any of 15 these criteria addressing extinction risk, because they presumably apply to independent 16 populations and, as previously discussed, lower Yuba River anadromous salmonids



18 Figure 4-8. Temporal trend and estimated annual number of phenotypic adult spring-run

17

¹⁹ Chinook salmon passing upstream of Daguerre Point Dam from 2004 through 2011.

^{20 (}Source: RMT 2013)

represent introgressive hybridization of larger Feather-Yuba river populations, with
 substantial contributions of hatchery-origin fish to the annual runs. As previously
 mentioned, the annual abundances of phenotypic spring-run Chinook salmon in the lower
 Yuba River are strongly influenced by hatchery fish, as discussed below.

5 ANNUAL ABUNDANCE OF ADIPOSE FIN-CLIPPED AND NON ADIPOSE FIN-CLIPPED SPRING-RUN 6 CHINOOK SALMON

Because the VAKI Riverwatcher systems located at both the north and south ladder of Daguerre Point Dam can record both silhouettes and electronic images of each fish passage event, the systems were able to differentiate Chinook salmon with adipose fins clipped or absent from Chinook salmon with their adipose fins intact. Thus, annual series of daily counts of Chinook salmon with adipose fins clipped (i.e., ad-clipped fish) and with adipose fins intact (i.e., not ad-clipped fish) that passed upstream of Daguerre Point Dam from March 1, 2004 through February 29, 2012 were obtained by RMT (2013).

14 The estimated numbers of spring-run Chinook salmon of hatchery (i.e., ad-clipped fish) 15 and potentially non-hatchery origin (i.e., not ad-clipped fish) passing upstream of 16 Daguerre Point Dam for the last eight years of available VAKI Riverwatcher data are presented in Table 4-6. Examination of Table 4-6 demonstrates a sharp increase in the 17 18 annual percent contribution of ad-clipped phenotypic spring-run Chinook salmon to the 19 total estimated annual run beginning in 2009 and extending through 2011 (RMT 2013). 20 This may be due, in part, to the fact that FRFH-origin spring-run Chinook salmon were 21 fractionally marked prior to 2005 and 100% marked thereafter. These fish would have 22 returned as age-3 fish during 2008. Also, fractional marking of fall-run hatchery fish at 23 the FRFH started during 2006, and these fish may return, to some extent, as phenotypic 24 spring-run Chinook salmon. Age 3 fish would have returned during 2009. The first full 25 year (age 3 and age 4) of recovery data from the CFM program occurred during 2010. 26 Evaluation of the lower Yuba River carcass survey data indicated that hatchery-origin 27 Chinook salmon comprised an estimated 71% of the total 2010 Chinook salmon run (Kormos et al. 2012, as cited in RMT 2013), although it was not possible to differentiate 28 29 between phenotypic spring- and fall-run Chinook salmon in the lower Yuba River carcass 30 surveys (RMT 2013).

1 Table 4-6. Estimated numbers of Chinook salmon, ad-clipped and non ad-clipped 2 phenotypic spring-run Chinook salmon that passed upstream of Daguerre Point Dam 3 annually from 2004 through 2011. (Source: RMT 2013)

| Year | Demarcation | Chinook Salmon Passage Upstream of Daguerre Point Dam | | | | | | | | | | | | |
|------|-------------|---|---------------------------|------------|----------------|--------------|--|--|--|--|--|--|--|--|
| | Data | All Chinook | Spring-run Chinook Salmon | | | | | | | | | | | |
| | Date | Salmon | Total | Ad-Clipped | Not Ad-Clipped | % Ad-Clipped | | | | | | | | |
| 2004 | 8/1/04 | 5,927 | 738 | 72 | 666 | 10 | | | | | | | | |
| 2005 | 8/24/05 | 11,374 | 3,592 | 676 | 2,916 | 19 | | | | | | | | |
| 2006 | 9/6/06 | 5,203 | 1,326 | 81 | 1,245 | 6 | | | | | | | | |
| 2007 | 9/4/07 | 1,394 | 372 | 38 | 334 | 10 | | | | | | | | |
| 2008 | 8/10/08 | 2,533 | 521 | 15 | 506 | 3 | | | | | | | | |
| 2009 | 7/9/09 | 5,378 | 723 | 213 | 510 | 29 | | | | | | | | |
| 2010 | 7/6/10 | 6,469 | 2,886 | 1,774 | 1,112 | 61 | | | | | | | | |
| 2011 | 9/7/11 | 7,785 | 1,159 | 323 | 836 | 28 | | | | | | | | |

4

5 The average contribution of adipose fin-clipped phenotypic spring-run Chinook salmon 6 to the total annual run size in the lower Yuba River, as inferred by the percentage of 7 adipose fin-clipped fish passing upstream of Daguerre Point Dam during the annual 8 defined phenotypic period, has been 20.8% over the eight years of available data and, 9 assuming a 3-year generation, the four most recent 3-year running averages of adipose 10 fin-clipped phenotypic spring-run Chinook salmon to the total annual run size have been 11 39.6%, 31.3%, 14.2%, and 6.4%, respectively. The average contribution of adipose fin-12 clipped phenotypic spring-run Chinook salmon to the total annual run sizes of these four 13 generations is 22.9%. The RMT (2013) recognized that there are limitations to simply 14 using percent adipose fin-clipped spring-run Chinook salmon passing through the VAKI 15 Riverwatcher systems as an estimate of total hatchery influence, and that resulting 16 estimates should be considered as minimum estimates. It is important to note that the 17 adipose fin-clipped phenotypic spring-run Chinook salmon abundance represents a 18 minimum indicator of hatchery-origin individuals due to fractional marking of spring-run 19 hatchery fish prior to 2005, and constant fractional marking (CFM) of fall-run hatchery 20 fish at the FRFH since 2006 which may return as phenotypic spring-run Chinook salmon. 21 It also is recognized that the hatchery influence criterion presumably is applicable to an

independent, genetically distinct population. However, as previously discussed, the phenotypic spring-run Chinook salmon in the lower Yuba River actually represents hybridization between spring- and fall-run Chinook salmon in the lower Yuba River, and hybridization with Feather River stocks including the FRFH spring-run Chinook salmon
 stock, which itself represents a hybridization between Feather River fall- and spring-run
 Chinook salmon populations.

4 APPLICABILITY OF ADDITIONAL VSP PARAMETERS AND EXTINCTION RISK CRITERIA

5 The M&E Program Framework developed by the RMT (2010) utilized VSP performance indicators that were identified based on the precept that the lower Yuba River 6 7 anadromous salmonid populations represented independent populations. However, the 8 RMT has identified a substantial amount of reproductive interaction between lower Yuba 9 River and lower Feather River anadromous salmonid stocks. As described in RMT 10 (2013), phenotypic spring-run Chinook salmon in the lower Yuba River likely represents 11 hybridization between spring- and fall-run Chinook salmon in the lower Yuba River, 12 hybridization with Feather River fall- and spring-run Chinook salmon stocks, and 13 hybridization with the FRFH spring-run Chinook salmon stock, which itself represents 14 hybridization between Feather River fall- and spring-run Chinook salmon populations. 15 Additionally, it is likely that anadromous O. mykiss stocks are similarly hybridized, with 16 fluid intermixing of lower Feather River and lower Yuba River fish.

17 The recognition of the extent of hybridization and lack of reproductive isolation of lower Yuba River and lower Feather River anadromous salmonid stocks logically constrains the 18 19 manner in which the VSP concept can be applied to the lower Yuba River, because many 20 of the VSP metrics are designed to evaluate the viability of discrete, independent 21 populations. Even the simplified approach suggested by Lindley et al. (2007) to evaluate 22 'extinction risk' is of limited applicability in the evaluation of highly introgressed 23 populations whose evaluation metrics are directly influenced by other stocks, and out-of-24 basin factors.

Lindley et al. (2007) provide criteria to assess the level of risk of extinction of Pacific salmonids based on population size, recent population decline, occurrences of catastrophes within the last 10 years that could cause sudden shifts from a low risk state to a higher one, and the impacts of hatchery influence. Populations with a low risk of extinction (less than 5% chance of extinction in 100 years) are those with a minimum total escapement of 2,500 spawners in 3 consecutive years (mean of 833 fish per year), no apparent decline in escapement, no catastrophic declines within the last 10 years, and
a low hatchery influence (NMFS 2011a). The overall estimated risk of extinction for the
population is determined by the highest risk score for any category Lindley et al. (2007).
While more detailed population viability assessment (PVA) models could be constructed
to assess Chinook salmon populations, Lindley et al. (2007) suggest any PVA results
should be compared with the results of applying their simpler criteria to estimate status
(NMFS 2011a).

8 Only some of the VSP performance indicators identified in the RMT (2010) M&E 9 Program framework and some of the extinction risk criteria provided by Lindley et al. 10 (2007) are appropriate for application specifically to lower Yuba River anadromous 11 salmonids. VSP performance indicators regarding spatial structure are applicable to the 12 habitat conditions in the lower Yuba River. Similarly, the catastrophe occurrence 13 extinction risk criterion also is applicable to the lower Yuba River. The extinction risk 14 criteria including abundance, and trends in abundance are of limited applicability and 15 serve as illustrative comparative measures in consideration of the non-independent 16 salmonid populations in the lower Yuba River. The hatchery risk extinction criterion 17 does not appear to be applicable to the non-independent lower Yuba River salmonid 18 populations. Considerations regarding each of these applicabilities are discussed below.

19 SPATIAL STRUCTURE

According to McElhany et al. (2000), spatial structure reflects how abundance is distributed among available or potentially available habitats, and how it can affect overall extinction risk and evolutionary processes that may alter a population's ability to respond to environmental change. A population's spatial structure depends fundamentally on habitat quality, spatial configuration, and dynamics, as well as on the dispersal characteristics of individuals in the population.

26 Performance indicators and analytics addressing spatial structure include spatial 27 organization of morphological units (e.g., lateral variability/diversity, adjacency, 28 randomness, and abundance), persistence of morphological units through time, and the 29 quality, number, size and distribution of morphological units available for spawning 30 Chinook salmon. Additional considerations include floodplain connectivity,

entrenchment, channel sinuosity, substrate size, changes in topographic depth, scour and
 fill processes, bankfull and flood flow recurrence interval, and maintenance of watershed
 processes to maintain suitable habitat for anadromous salmonid lifestages.

As stated in the M&E Plan (RMT 2010a), the spatial structure evaluation includes examination of maintenance of watershed processes and regulatory management practices to create and maintain suitable habitat for all freshwater lifestages of spring-run and fall-run Chinook salmon, and steelhead. As discussed in RMT (2013), one of the performance indicators preliminarily evaluated by Wyrick and Pasternack (2012) is whether the sequence of morphological units in the lower Yuba River is non-random. Highly disturbed systems often degrade into homogeneity or randomness.

11 Of the 12 major near-bankfull morphological units, the most uniformly distributed (i.e., 12 randomly located) units are slackwater, slow glide, and lateral bar. As an example of 13 non-uniform distribution, pool units were predominantly found in the upstream reaches 14 (i.e., Englebright and Timbuctoo Bend) and the downstream reach (i.e., Marysville), but 15 were less abundant in the middle, wider reaches (i.e., Daguerre Point Dam and Dry 16 Creek). Consequently, evaluation of the morphological units in the lower Yuba River as 17 part of the spatial structure analyses indicates that, in general, the sequence of 18 morphological units is non-random, indicating that the channel has been self-sustaining 19 of sufficient duration to establish an ordered spatial structure (refer to RMT 2013 for 20 additional discussion).

Another new method for analyzing the morphological unit organization that Wyrick and Pasternack (2012) developed is an adjacency probability analysis, which evaluates the frequency at which each morphological unit is adjacent to every other unit, and compares that against random adjacency expectations. Results of this analysis indicate that the inchannel units near the thalweg typically exhibit low adjacency probabilities to the bar units, although they do exhibit higher-than-random probabilities to other in-channel units.

Wide, diverse rivers should also exhibit lateral variability in its form-process associations. In the lower Yuba River, morphological unit organization highlights the complexity of the channel geomorphology, as well as the complex and diverse suite of potential habitat at any given location in the Yuba River. The above summary (described in more detail in RMT 2013) illustrates that spatial structure of morphological units in the
 lower Yuba River is complex, diverse, and persistent.

3 CATASTROPHE OCCURRENCE

4 According to Lindley et al. (2007), the catastrophe criteria trace back to Mace and Lande 5 (1991), and the underlying theory is further developed by Lande (1993). The following 6 discussion was taken from Lindley et al. (2007). The overall goal of the catastrophe 7 criteria is to capture a sudden shift from a low risk state to a higher one. Catastrophes are 8 defined as instantaneous declines in population size due to events that occur randomly in 9 time, in contrast to regular environmental variation, which occurs constantly and can 10 have both positive and negative effects on the population. Lindley et al. (2007) view 11 catastrophes as singular events with an identifiable cause and only negative immediate 12 consequences, as opposed to normal environmental variation which can produce very 13 good as well as very bad conditions. Some examples of catastrophes include disease 14 outbreaks, toxic spills, or volcanic eruptions. A high risk situation is created by a 90% 15 decline in population size over one generation. A moderate risk event is one that is 16 smaller but biologically significant, such as a year-class failure.

17 EXTINCTION RISK CRITERIA AND APPLICATION

18 Lindley et al. (2007) characterized the spring-run Chinook salmon population in the 19 lower Yuba River as data deficient, and therefore did not characterize its viability. In 20 2007, there was limited information on the current population size of spring-run Chinook salmon in the lower Yuba River. NMFS' 5 Year Status Review for the Central Vallev 21 22 Spring-run Chinook Salmon ESU (NMFS 2011) reported that the annual spawning run 23 size of spring-run Chinook salmon in the lower Yuba River generally ranges from a few 24 hundred to a few thousand fish with the annual trend closely following the annual 25 abundance trend of the Feather River Hatchery spring-run Chinook salmon population. 26 NMFS (2011a) concluded that the Yuba River spring-run Chinook salmon population 27 satisfies the moderate extinction risk criteria for abundance, but likely falls into the high 28 risk category for hatchery influence.

1 Criteria to assess extinction risk of Pacific salmonids are based on population size, recent 2 population decline, occurrences of catastrophes within the last 10 years, and the impacts 3 of hatchery influence (Lindley et al. 2007). As previously discussed, for the last three 4 consecutive years, an estimated total of 4,768 phenotypic spring-run Chinook salmon 5 have passed upstream of Daguerre Point Dam, with an average of 1,589 fish per year. 6 Catastrophes have not occurred in the Yuba River Basin, nor have catastrophic declines 7 been observed within the phenotypic spring-run Chinook salmon abundance estimates 8 within the last ten years. The abundance of phenotypic spring-run Chinook salmon in the 9 lower Yuba River has exhibited a very slight increase over the eight years examined, 10 although the positive trend is not statistically significant. These abundance and trend 11 considerations would correspond to low extinction risk according to NMFS criteria 12 (Lindley et al. 2007). However, RMT (2013) questions the applicability of any of these 13 criteria addressing extinction risk, because they presumably apply to independent 14 populations and, as previously discussed, lower Yuba River anadromous salmonids 15 represent introgressive hybridization of larger Feather-Yuba river populations, with 16 substantial contributions of hatchery-origin fish to the annual runs. For additional 17 discussion, see RMT (2013).

18 The average contribution of adipose fin-clipped phenotypic spring-run Chinook salmon 19 to the total annual run size in the lower Yuba River, as inferred by the percentage of 20 adipose fin-clipped fish passing upstream of Daguerre Point Dam during the annual 21 defined phenotypic period, has been 20.8% over the eight years of available data and, 22 assuming a 3-year generation, the four most recent 3-year running averages of adipose 23 fin-clipped phenotypic spring-run Chinook salmon to the total annual run size have been 24 39.6%, 31.3%, 14.2%, and 6.4%, respectively. The average contribution of adipose fin-25 clipped phenotypic spring-run Chinook salmon to the total annual run sizes of these four 26 generations is 22.9%. RMT (2013) recognized that there are limitations to simply using 27 percent adipose fin-clipped spring-run Chinook salmon passing through the VAKI 28 Riverwatcher systems as an estimate of total hatchery influence, and that resulting 29 estimates should be considered as minimum estimates. As previously discussed, it is 30 important to note that the adipose fin-clipped phenotypic spring-run Chinook salmon 31 abundance represents a minimum indicator of hatchery-origin individuals due to

fractional marking of spring-run hatchery fish prior to 2006, and constant fractional
 marking (CFM) of fall-run hatchery fish at the FRFH which may return as phenotypic
 spring-run Chinook salmon.

It also is recognized that the hatchery influence criterion presumably is applicable to an independent, genetically distinct population (RMT 2013). However, as previously discussed, the phenotypic spring-run Chinook salmon in the lower Yuba River actually represents hybridization between spring- and fall-run Chinook salmon in the lower Yuba River, and hybridization with Feather River stocks including the FRFH spring-run Chinook salmon stock, which itself represents a hybridization between Feather River falland spring-run Chinook salmon populations.

11 Although straying of FRFH-origin Chinook salmon into the lower Yuba River occurs, 12 available information indicates that: (1) the FRFH spring-run Chinook salmon is included 13 in the ESU, in part because of the important role this stock may play in the recovery of 14 spring-run Chinook salmon in the Feather River Basin, including the Yuba River (70 FR 15 37160); (2) the spring-run Chinook program at FRFH is an Integrated Recovery Program 16 which seeks to aid in the recovery and conservation of Central Valley spring-run Chinook 17 salmon (DWR 2009a); and (3) fish produced at FRFH are intended to spawn in the wild 18 or be genetically integrated with the targeted natural population as FRFH broodstock 19 (DWR 2009a).

20 4.2.8 Public Review Draft Recovery Plan Considerations

According to NMFS (2005) *Recommendations for the Contents of Biological Assessments and Biological Evaluations* pertaining to status of the species in the action
area, a BA should:

- Identify any recovery plan implementation that is occurring in the action area,
 especially priority one action items from recovery plans.
- The NMFS Draft Recovery Plan establishes three population levels to help guide recovery efforts for existing populations, referred to as Core 1, 2, and 3 populations. The NMFS Draft Recovery Plan (pg. 65) identifies lower Yuba River spring-run Chinook

salmon [and steelhead] populations as Core 1 populations. Core 1 populations form the
 foundation of the recovery strategy, and Core 1 populations should be the first focus of an
 overall recovery effort (NMFS 2009).

To meet recovery objectives for the diversity groups, the conceptual recovery scenarios for the spring-run Chinook salmon ESU (pg. 99) [and the steelhead DPS (pg. 123)] include: (1) securing extant populations by implementing key habitat restoration actions, particularly in the near term; and (2) establishment of additional viable independent populations.

9 The NMFS Draft Recovery Plan states, that in order to secure a viable independent 10 population of spring-run Chinook salmon (pg. 116), [and to secure the extant population 11 and promote a viable population of steelhead (pg. 140)], in the lower Yuba River, several 12 key near-term and long-term habitat restoration actions were identified, including the 13 following:

- 14 □ Continued implementation of the Yuba Accord flow schedules to provide 15 suitable habitat (flow and water temperature) conditions for all lifestages 16 Improvements to adult salmonid upstream passage at Daguerre Point Dam 17 Improvements to juvenile salmonid downstream passage at Daguerre Point Dam 18 Implementation of a spawning gravel augmentation program in the uppermost 19 reach (i.e., Englebright Dam to the Narrows) of the lower Yuba River 20 Improvements to riparian habitats for juvenile salmonid rearing 21 Creation and restoration of side-channel habitats to increase the quantity and 22 quality of off-channel rearing (and spawning) areas 23 □ Implementation of projects to increase floodplain habitat availability to improve 24 habitat conditions for juvenile rearing 25 The NMFS Draft Recovery Plan includes Priority 1, Priority 2 and Priority 3 recovery 26 actions. The NMFS Draft Recovery Plan Appendix C (pgs. 2, 3) states "According to
- 27 NMFS' 1990 Endangered and Threatened Species Listing and Recovery Priority

1 Guidelines (55 FR 24296), recovery actions identified in a Recovery Plan are to be 2 assigned priorities of 1 to 3, as follows:

- 3 Priority 1 An action that must be taken to prevent extinction or to identify those
 4 actions necessary to prevent extinction
- 5 Priority 2 An action that must be taken to prevent a significant decline in 6 population numbers, habitat quality, or other significant negative impacts short of 7 extinction
- 8 *Priority 3 All other actions necessary to provide for full recovery of the species.*"

9 The NMFS Draft Recovery Plan (pg. 161) identifies the following proposed action as a
10 Priority 1 recovery action for the Yuba River:

11 **Recovery Action 1.9.6.1.** Develop and implement a phased approach to salmon 12 reintroduction planning to recolonize historic habitats above Englebright Dam. 13 Implement actions to: (1) enhance habitat conditions including providing flows and 14 suitable water temperatures for successful upstream and downstream passage, holding, 15 spawning and rearing; and (2) improve access within the area above Englebright Dam, 16 including increasing minimum flows, providing passage at Our House, New Bullards 17 Bar, and Log Cabin dams, and assessing feasibility of passage improvement at natural 18 barriers. The phased approach should include:

- 20 Conduct habitat evaluations
- 21 Conduct 3-5 year pilot testing program
- 22 Implement long-term fish passage program

The spring-run Chinook salmon conceptual recovery scenario also includes reintroduction of spring-run Chinook salmon to the candidate areas of the North Fork, Middle Fork and South Fork Yuba rivers. Reintroduction of anadromous salmonids above Englebright Dam has been the subject of recent and current investigations. Evaluation of habitat suitability for anadromous salmonids upstream of Englebright Dam was recently undertaken (DWR 2007), but those evaluations have yet to be finalized as part of the Upper Yuba River Watershed Studies Program. Currently, NMFS is
 evaluating the feasibility of providing passage for anadromous salmonids at Englebright
 Dam. Hence, the conceptual recovery scenario does not further discuss specific
 restoration actions associated with reintroduction.

5 The NMFS Draft Recovery Plan (pg. 161) identifies the following proposed action as a
6 Priority 1 recovery action for the Yuba River:

Recovery Action 1.9.6.2. Improve spawning habitat in the lower river by gravel
restoration program below Englebright Dam and improve rearing habitat by increasing
floodplain habitat availability.

Also, a gravel restoration program below Englebright Dam is discussed as a Priority 2
action on pg. 73, and lower Yuba River floodplain habitat availability considerations are
discussed as Priority 2 actions on pgs. 73, 74, 76, and 92 of Appendix C in NMFS (2009).

Proposed recovery action 1.9.6.2 actually includes two separate proposed actions: (1) improve spawning habitat in the lower river by gravel restoration program below Englebright Dam; and (2) improve rearing habitat by increasing floodplain habitat availability. Each of these is discussed separately, below.

- (1) Improve spawning habitat in the lower river by gravel restoration program below
 Englebright Dam. The Corps completed the injection of 500 tons of gravel
 approximately 200 yards downstream of Englebright on November 30, 2007
 (Grothe 2011). The Corps completed additional injections of 5,000 tons of gravel
 on January 13, 2011, August 21, 2012 and August 14, 2013.
- 22 (2) Improve rearing habitat by increasing floodplain habitat availability. Since the 23 NMFS Draft Recovery Plan was noticed in the Federal Register on October 6, 24 2009, substantial efforts have been undertaken to identify, develop and consider 25 the relative merits of habitat restoration actions in the lower Yuba River. The 26 need for restoration actions, identification of the specific actions themselves, and 27 the relative merits of the actions to expand habitat and accomplish the goals of the 28 Oroville FERC Relicensing Habitat Expansion Agreement (HEA) were presented 29 in a report submitted to the HEA Steering Committee during early November

1 2009 (YCWA et al. 2009). This report represents a comprehensive consideration 2 of such restoration actions developed for the lower Yuba River. The YCWA et al. 3 (2009) report identified several factors that continue to limit juvenile spring-run Chinook salmon [and steelhead] rearing habitat suitability in the lower Yuba 4 5 River, including: (1) sparse and restricted amounts of riparian vegetation and 6 associated instream object and overhanging object cover; (2) limited aquatic habitat complexity and diversity; and (3) altered natural river function and 7 morphology in the lower Yuba River. Shaded Riverine Aquatic (SRA) habitat 8 9 generally occurs in the lower Yuba River as scattered, short strips, with the most 10 extensive and continuous segments of SRA habitat occurring along bars where 11 recent channel migrations or avulsions have cut new channels through stands of 12 riparian vegetation.

13 Regarding juvenile salmonid rearing habitat, the NMFS Draft Recovery Plan states that, 14 in order to secure a viable independent population of spring-run Chinook salmon (pg. 15 116), [and to secure the extant population and promote a viable population of steelhead 16 (pg.140)] in the lower Yuba River, the following key near-term and long-term habitat 17 restoration actions should be implemented: (1) the creation and restoration of side 18 channel habitats to increase the quantity and quality of off-channel rearing (and 19 spawning) areas; (2) improvements to riparian habitats for juvenile salmonid rearing; and 20 (3) implementation of projects to increase floodplain habitat availability to improve 21 habitat conditions for juvenile rearing. Of the proposed actions regarding juvenile 22 rearing, the actions that would be most beneficial and cost-effective for juvenile rearing 23 habitat, and the actions that would yield the most immediate benefits, are the creation of 24 new side-channel habitats associated with existing stands of riparian vegetation that are 25 not presently hydraulically connected to the river channel (YCWA 2010). Specifically, 26 new side-channel habitats would: (1) increase and maintain existing riparian vegetation; 27 (2) provide instream object and overhanging object cover; (3) provide new SRA, and 28 associated allochthonous food sources for rearing juveniles; (4) increase aquatic habitat 29 complexity and diversity; (5) provide habitats more consistent with those previously 30 available in the upper watershed; and (6) provide predator escape cover, and overall 31 increased survival of juvenile spring-run Chinook salmon and steelhead.

1 The NMFS Draft Recovery Plan (pg. 83) states "The [Draft Plan's recovery] scenarios 2 represent some of the many possible combinations of populations, restoration actions, 3 risk minimization and threat abatement. Different scenarios may fulfill the biological requirements for recovery". The NMFS Draft Recovery Plan (pg. 83) further states "As 4 5 this Recovery Plan is implemented over time, additional information will become 6 available to help determine whether the threats have been abated, to further develop 7 understanding of the linkages between threats and Chinook salmon and steelhead 8 population responses, and to evaluate the viability of Chinook salmon and steelhead in 9 the Central Valley Domain ... Such information is expected to lead to adjustments in 10 recovery expectations and restoration actions and, thus, recovery scenarios."

The NMFS Draft Recovery Plan (pg. 208) states that it may not be necessary to reintroduce fish to all of the listed river and creek systems to meet the recovery criteria for Central Valley spring-run Chinook salmon [and steelhead]. "*It may not be necessary to re-establish populations to all of these rivers. The highest priority areas are the Little Sacramento River, the McCloud River, the North Fork American River, and the San Joaquin River.*"

17 4.3 Central Valley Steelhead DPS

18 **4.3.1 ESA Listing Status**

19 On March 19, 1998 (63 FR 13347) NMFS listed the California Central Valley steelhead 20 ESU as "threatened", concluding that the risks to Central Valley steelhead had 21 diminished since the completion of the 1996 status review based on a review of existing 22 and recently implemented state conservation efforts and federal management programs 23 (e.g., CVPIA, AFRP, CALFED) that address key factors for the decline of this species. 24 The California Central Valley steelhead ESU included all naturally spawned populations 25 of steelhead in the Sacramento and San Joaquin rivers and their tributaries, but excluded 26 steelhead from the tributaries of San Francisco and San Pablo bays (NMFS 2004b).

On June 14, 2004, NMFS proposed listing determinations for 27 ESUs of West Coast
salmon and *O. mykiss*, including the California Central Valley steelhead ESU. In the

proposed rule, NMFS concluded that steelhead were not in danger of extinction, but were likely to become endangered within the foreseeable future throughout all or a significant portion of their range and, thus, proposed that steelhead remain listed as threatened under the ESA. Steelhead from the Coleman National Fish Hatchery and the FRFH, as well as resident populations of *O. mykiss* (rainbow trout) below impassible barriers that co-occur with anadromous populations, were included in the California Central Valley steelhead ESU and, therefore, also were included in the proposed listing.

8 During the 2004 comment period on the proposed listings, the USFWS provided 9 comments that the USFWS does not use NMFS' ESU policy in any USFWS ESA listing 10 decisions. As a result of the comments received, NMFS re-opened the comment period to 11 receive comments on a proposed alternative approach to delineating "species" of West 12 Coast O. mykiss (70 FR 67130). NMFS proposed to depart from past practice of applying 13 the ESU Policy to O. mykiss stocks, and instead proposed to apply the DPS Policy in 14 determining "species" of O. mykiss for listing consideration. NMFS noted that within a 15 discrete group of O. mykiss populations, the resident and anadromous life forms of O. 16 mykiss remain "markedly separated" as a consequence of physical, physiological, 17 ecological, and behavioral factors, and may therefore warrant delineation as separate 18 DPSs (71 FR 834).

19 NMFS issued a policy for delineating distinct population segments of Pacific salmon in 20 1991 (56 FR 58612; November 20, 1991). Under this policy, a group of Pacific salmon 21 populations is considered an "Evolutionarily Significant Unit" if it is substantially 22 reproductively isolated from other conspecific populations, and it represents an important 23 component in the evolutionary legacy of the biological species. Further, an ESU is 24 considered to be a "Distinct Population Segment" (and thus a "species") under the 25 ESA. In 1996, NMFS and USFWS adopted a joint policy for recognizing DPSs under the 26 ESA (DPS Policy; 61 FR 4722; February 7, 1996). The DPS Policy adopted criteria 27 similar to, but somewhat different from, those in the ESU Policy for determining when a group of vertebrates constitutes a DPS - The group must be discrete from other 28 29 populations, and it must be significant to its taxon. A group of organisms is discrete if it 30 is "markedly separated from other populations of the same taxon as a consequence of 31 physical, physiological, ecological, and behavioral factors." Significance is measured

with respect to the taxon (species or subspecies) as opposed to the full species (71 FR
834). Although the ESU Policy did not by its terms apply to steelhead, the DPS Policy
stated that NMFS will continue to implement the ESU Policy with respect to "Pacific
salmonids" (which included *O. mykiss*). In a previous instance of shared jurisdiction
over a species (Atlantic salmon), NMFS and USFWS used the DPS Policy in their
determination to list the Gulf of Maine DPS of Atlantic salmon as endangered (65 FR
69459; November 17, 2000).

Given NMFS and USFWS shared jurisdiction over *O. mykiss*, and consistent with joint
NMFS and USFWS approaches for Atlantic salmon, it was concluded that application of
the joint DPS policy to was logical, reasonable, and appropriate for identifying DPSs of *O. mykiss* (71 FR 834). Moreover, NMFS determined that use of the ESU policy —
originally intended for Pacific salmon — should not continue to be extended to *O. mykiss*, a type of salmonid with characteristics not typically exhibited by Pacific salmon
(71 FR 834).

15 On January 5, 2006 NMFS issued a final decision that defined Central Valley steelhead 16 as a DPS rather than an ESU, and retained the status of Central Valley steelhead as 17 threatened (71 FR 834). The DPS includes all naturally spawned anadromous O. mykiss 18 (steelhead) populations below natural and manmade impassable barriers in the 19 Sacramento and San Joaquin Rivers and their tributaries, excluding steelhead from San 20 Francisco and San Pablo Bays and their tributaries (63 FR 13347). Steelhead in two 21 artificial propagation programs — the Coleman National Fish Hatchery, and FRFH 22 steelhead hatchery programs are considered to be part of the DPS. NMFS determined 23 that these artificially propagated stocks are no more divergent relative to the local natural 24 population(s) than what would be expected between closely related natural populations 25 within the DPS (71 FR 834).

As previously discussed, the ESA requires that NMFS review the status of listed species under its authority at least every five years and determine whether any species should be removed from the list or have its listing status changed. In August 2011, NMFS completed a 5-year status review of the Central Valley steelhead DPS. Based upon a review of available information, NMFS (2011c) recommended that the Central Valley

1 steelhead DPS remain classified as a threatened species. However, NMFS (2011c) also 2 indicated that the biological status of the DPS has declined since the previous status 3 review in 2005 and, therefore, NMFS recommend that the DPS's status is reassessed in 2 4 to 3 years if it does not respond positively to improvements in environmental conditions 5 and management actions. In the interim period, NMFS also recommended that the status 6 of the DPS should be monitored and the most recent genetic information for the DPS, 7 including information for the four steelhead hatchery stocks, should be reviewed to re-8 assess the DPS membership status of the Nimbus and Mokelumne River hatcheries. New 9 information resulting from the genetics review should be incorporated into any updated 10 status review for the DPS (NMFS 2011c).

11 4.3.2 Critical Habitat Designation

12 On February 16, 2000 (65 FR 7764), NMFS published a final rule designating critical 13 habitat for Central Valley steelhead. This critical habitat includes all river reaches 14 accessible to listed steelhead in the Sacramento and San Joaquin rivers and their 15 tributaries in California, including the lower Yuba River upstream to Englebright Dam. 16 NMFS proposed new Critical Habitat for spring-run Chinook salmon and Central Valley 17 steelhead on December 10, 2004 (69 FR 71880) and published a final rule designating 18 critical habitat for these species on September 2, 2005. This critical habitat includes the 19 lower Yuba River (70 FR 52488) from the confluence with the lower Feather River 20 upstream to Englebright Dam.

21 4.3.2.1 Primary Constituent Elements

The critical habitat designation (70 FR 52488) lists PCEs, which are physical or biological elements essential for the conservation of the listed species. The PCEs include sites essential to support one or more lifestages of the DPS (sites for spawning, rearing, migration, and foraging). The specific PCEs include:

- 1 🖸 Estuarine areas
- 2 D Nearshore marine areas
- 3 Offshore marine areas

The most recent discussion of PCEs in the Central Valley is in the CVP/SWP OCAP
Biological Opinion (NMFS 2009a). The following summary descriptions of the current
conditions of the PCEs for the Central Valley steelhead DPS were taken from
NMFS (2009a).

8 Freshwater Spawning Habitat

9 According to NMFS (2009), steelhead in the Sacramento River spawn primarily between 10 Keswick Dam and Red Bluff Diversion Dam during the winter and spring. The highest 11 density spawning area is likely in the upstream portion of this area in the vicinity of the 12 city of Redding, although detailed surveys of steelhead spawning in the mainstem 13 Sacramento River are not available. Most Sacramento River steelhead probably spawn in 14 the tributary streams. Steelhead spawn in Clear Creek mostly within a couple miles of 15 Whiskeytown Dam but spawning extends for about 10 miles downstream of the dam (M. 16 Brown, pers. comm. as cited in Reclamation 2008). Steelhead spawn in the Feather River 17 from the fish barrier dam downstream to Gridley with nearly 50% of all spawning 18 occurring the first mile of the low flow channel (DWR 2003). Steelhead spawn in the 19 American River from Nimbus Dam (RM 23) downstream to the lowest riffle in the river 20 at Paradise Beach (RM 5). Most spawning is concentrated in the upper seven miles of the 21 river (Hannon and Deason 2008). Steelhead (and/or rainbow trout) spawn in the 22 Stanislaus River from Goodwin Dam downstream to approximately the city of Oakdale. 23 Steelhead spawning surveys have not been conducted in the Stanislaus River so detailed 24 spawning distribution is unknown but based on observations of trout fry, most spawning 25 occurs upstream of Orange Blossom Bridge.

26 Freshwater Rearing Habitat

Juvenile steelhead reside in freshwater for a year or more, so they are more dependent on
freshwater rearing habitat than are the ocean type Chinook salmon in the Central Valley.
Steelhead rearing occurs primarily in the upstream reaches of the rivers where channel

1 gradients tend to be higher and, during the warm weather months, where temperatures are 2 maintained at more suitable levels by cool water dam releases. The Sacramento River 3 contains a long reach of suitable water temperatures even during the heat of the summer. 4 Steelhead rearing in the Sacramento River occurs mostly between Keswick Dam (RM 5 302) and Butte City (RM 169) with the highest densities likely to be upstream of Red 6 Bluff Diversion Dam. Steelhead rearing in Clear Creek is concentrated in the upper river higher gradient areas but probably occurs down to the mouth. Steelhead rearing in the 7 8 Feather River is concentrated in the low flow channel where temperatures are most 9 suitable (DWR 2004c). Steelhead rearing in the American River occurs down to Paradise 10 Beach, with concentrations during the summer on most major riffle areas and highest 11 densities near the higher density spawning areas. Steelhead rearing in the Stanislaus 12 River occurs upstream of Orange Blossom Bridge, where gradients are highest. The 13 highest rearing densities are upstream of Knights Ferry (Kennedy and Cannon 2002).

14 Freshwater Migration Corridors

15 Steelhead migrate during the winter and spring of the year, as juveniles, from the rearing 16 areas described above downstream through the rivers and the Delta to the ocean. The 17 habitat conditions they encounter during migration from the upstream reaches of the 18 rivers downstream to the Delta generally become less suitable as fish move away from 19 their natal streams until they reach the ocean. The generally non-turbulent flows and 20 sand substrates found in the lower river reaches are not preferred types of habitat, so 21 steelhead do not likely reside for extended periods in these areas except when food 22 supplies, such as smaller young fish, are abundant and temperatures are suitable. 23 Predatory fishes such as striped bass tend to be more abundant in the lower rivers and the 24 Delta. Emigration conditions for juvenile steelhead in the Stanislaus River down through 25 the San Joaquin River and the south Delta tend to be less suitable than conditions for 26 steelhead emigrating from the Sacramento River and its tributaries.

Adult steelhead migrate upstream from the ocean to their spawning grounds near the terminal dams primarily during the fall and winter months. Flows are generally lower during the upstream migrations than during the outmigration period. Areas where their upstream progress can be affected are the Delta Cross Channel Gates, RBDD, and
 Anderson Cottonwood Irrigation District Diversion Dam.

3 <u>ESTUARINE HABITAT AREAS</u>

Steelhead use the San Francisco estuary as a rearing area and migration corridor between their upstream rearing habitat and the ocean. The San Francisco Bay estuarine system includes the waters of San Francisco Bay, San Pablo Bay, Grizzley Bay, Suisuin Bay, Honker Bay, and can extend as far upstream as Sherman Island during dry periods. At times steelhead likely remain for extended periods in areas of suitable habitat quality where food such as young herring, salmon and other fish and invertebrates is available.

10 NEARSHORE COASTAL MARINE AND OFFSHORE MARINE AREAS

The most recent discussion of PCEs for the Central Valley steelhead DPS (NMFS 2009a) did not include the PCEs of nearshore coastal marine and offshore marine areas. Although relatively little is known about steelhead utilization of nearshore coastal marine and offshore marine areas, it is reasonable to assume that the discussion of these PCEs previously provided for spring-run Chinook salmon in Section 4.1 of this BA generally is applicable to steelhead.

17 **4.3.3** Historical Distribution and Abundance

According to NMFS (2009), steelhead historically occurred naturally throughout the Sacramento and San Joaquin River basins, although stocks have been extirpated from large areas in both basins. The California Advisory Committee on Salmon and Steelhead (CDFG 1988) reported a reduction in Central Valley steelhead habitat from 6,000 miles historically to 300 miles.

NMFS (2009) reported that prior to dam construction, water development and watershed perturbations, Central Valley steelhead were distributed throughout the Sacramento and San Joaquin rivers (Busby et al. 1996; McEwan 2001). Steelhead were found from the upper Sacramento and Pit rivers (now inaccessible due to Shasta and Keswick dams) south to the Kings and possibly the Kern River systems, and in both east- and west-side Sacramento River tributaries (Yoshiyama et al. 1996). Lindley et al. (2006) estimated 1 that historically there were at least 81 independent Central Valley steelhead populations 2 distributed primarily throughout the eastern tributaries of the Sacramento and San 3 Joaquin rivers. Presently, impassable dams block access to 80% of historically available 4 habitat, and block access to all historical spawning habitat for about 38% of historical 5 populations (Lindley et al. 2006). Existing wild steelhead stocks in the Central Valley 6 are mostly confined to the upper Sacramento River and its tributaries, including Antelope 7 Creek, Deer Creek, and Mill Creek, and the Yuba River. Populations may exist in Big 8 Chico and Butte creeks, and a few wild steelhead are produced in the American and 9 Feather rivers (McEwan 2001).

Until recently, steelhead were thought to be extirpated from the San Joaquin River
system. Recent monitoring has detected small self-sustaining populations of steelhead in
the Stanislaus, Mokelumne, and Calaveras rivers, and other streams previously thought to
be devoid of steelhead (McEwan 2001).

14 It is possible that naturally spawning populations exist in many other streams but are 15 undetected due to lack of monitoring programs (IEP Steelhead Project Work Team 1999, 16 as cited in NMFS 2009). Incidental catches and observations of steelhead juveniles also 17 have occurred on the Tuolumne and Merced Rivers during fall-run Chinook salmon 18 monitoring activities, indicating that steelhead are widespread, throughout accessible 19 streams and rivers in the Central Valley (Good et al. 2005). Naturally spawning 20 populations of steelhead also occur in the Feather, Yuba, American, and Mokelumne 21 rivers, but these populations have had substantial hatchery influence and their ancestries 22 are not clear (Busby et al. 1996). Steelhead runs in the Feather and American rivers are 23 sustained largely by the FRFH and Nimbus Hatchery (McEwan and Jackson 1996). 24 Steelhead also currently occur in the Stanislaus, Calaveras, Merced, and Tuolumne rivers 25 (NMFS 2009).

Historic Central Valley steelhead run sizes are difficult to estimate because of the lack of data, but McEwan (2001) suggested that steelhead run sizes may have approached one to two million adults annually. McEwan and Jackson (1996) suggested that by the early 1960s, the steelhead run size had declined to about 40,000. Over the last 30 years the steelhead populations in the upper Sacramento River have declined substantially (NMFS

1 2009). In 1996, NMFS estimated the Central Valley total run size based on dam counts. 2 hatchery returns, and past spawning surveys was probably fewer than 10,000 fish. Both 3 natural and hatchery runs have declined since the 1960s. Counts at RBDD averaged 4 1,400 fish from 1991 to 1996, compared to counts in excess of 10,000 fish in the late 5 1960s (McEwan and Jackson 1996). American River redd surveys and associated 6 monitoring from 2002 through 2007 indicate that only a few hundred steelhead spawn in 7 the river and a portion of those spawners originated from Nimbus Hatchery (Hannon and 8 Deason 2008).

9 Specific information regarding steelhead spawning within the mainstem Sacramento 10 River is limited due to lack of monitoring (NMFS 2004). Currently, the number of 11 steelhead spawning in the Sacramento River is unknown because redds cannot be 12 distinguished from a large resident rainbow trout population that has developed as a 13 result of managing the upper Sacramento River for coldwater species.

The lack of sustained monitoring programs for steelhead throughout most of the Central Valley persists to the present time. There is a paucity of reliable data to estimate run sizes of steelhead in the Central Valley, particularly wild stocks. However, some steelhead escapement monitoring surveys have been initiated in upper Sacramento River tributaries (e.g., Beegum, Deer, and Antelope Creeks) using snorkel methods similar to spring-run Chinook escapement surveys (NMFS 2009a).

There is a general lack of steelhead population monitoring in most of the Central Valley (NMFS 2009a). Lindley et al. (2007) stated that there are almost no data with which to assess the status of any of the Central Valley steelhead populations. They further stated that Central Valley steelhead populations are classified as data deficient, with the exceptions restricted to streams with long-running hatchery programs including Battle Creek and the Feather, American and Mokelumne rivers.

- According to NMFS (2007a), in the *Updated Status Review of West Coast Salmon and Steelhead* (Good et al. 2005), the Biological Review Team made the following conclusion based on steelhead Chipps Island trawl data:
- 29 "If we make the fairly generous assumptions (in the sense of generating large estimates of
- 30 spawners) that average fecundity is 5,000 eggs per female, 1% of eggs survive to reach

Chipps Island, and 181,000 smolts are produced (the 1998-2000 average), about 3,628
 female steelhead spawn naturally in the entire Central Valley."

In the Yuba River, definitive historic population estimates do not exist for steelhead, but
it is likely that the river supported large steelhead runs in the 1800s (USFWS 1995).
McEwan and Jackson (1996) reported that the Yuba River historically supported the
largest, naturally reproducing, persistent population of steelhead in the Central Valley.

7 Prior to construction of Englebright Dam in 1941, CDFW fisheries biologists stated that 8 they observed large numbers of steelhead spawning in the uppermost reaches of the Yuba 9 River and its tributaries (CDFG 1998; Yoshiyama et al. 1996). After construction of 10 Englebright Dam in 1941, CDFW estimated that only approximately 200 steelhead 11 spawned in the lower Yuba River annually before New Bullards Bar Reservoir was 12 completed in 1969. From 1970 to 1979, CDFW annually stocked 27,270-217,378 13 fingerlings, yearlings, and sub-catchables from Coleman National Fish Hatchery into the 14 lower Yuba River (CDFG 1991a). CDFW stopped stocking steelhead into the lower 15 Yuba River in 1979. Based on angling data, CDFW estimated a run size of 2,000 16 steelhead in the lower Yuba River in 1975 (CDFG 1991a). McEwan and Jackson (1996) 17 reported that, as of 1996, the status of the lower Yuba River steelhead population was 18 unknown, but it appeared to be stable and able to support a significant sport fishery. 19 CDFW currently manages the river to protect natural steelhead through strict "catch-and-20 release" fishing regulations.

21 **4.3.4 General Life History and Habitat Requirements**

Steelhead exhibits perhaps the most complex suite of life-history traits of any species of
Pacific salmonid. Members of this species can be anadromous or freshwater residents
and, under some circumstances, members of one form can apparently yield offspring of
another form (YCWA 2010).

26 "Steelhead" is the name commonly applied to the anadromous form of the biological 27 species *O. mykiss*. The physical appearance of *O. mykiss* adults and the presence of 28 seasonal runs and year-round residents indicate that both anadromous (steelhead) and 29 resident rainbow trout exist in the lower Yuba River downstream of Englebright Dam,

1 although no definitive visual characteristics have been identified to distinguish young 2 steelhead from resident trout (SWRI et al. 2000). Zimmerman et al. (2009) analyzed 3 otolith strontium:calcium (Sr:Ca) ratios in 964 otolith samples comprised of young-of-4 year, age-1, age-2, age-3, and age-4+ fish to determine maternal origin and migratory 5 history (anadromous vs. non-anadromous) of O. mykiss collected in Central Valley rivers 6 between 2001 and 2007, including the lower Yuba River. The proportion of steelhead 7 progeny in the lower Yuba River (about 13%) was intermediate to the other rivers 8 examined (Sacramento, Deer Creek, Calaveras, Stanislaus, Tuolumne, and Merced), 9 which ranged from about 4% in the Merced River to 74% in Deer Creek (Zimmerman et 10 al. 2009). Results from Mitchell (2010) indicate O. mykiss in the lower Yuba River are 11 exhibiting a predominately residential life history pattern. He found that 14% of scale 12 samples gathered from 71 O. mykiss moving upstream and trapped in the fish ladder at 13 Daguerre Point Dam from November 1, 2000, through March 28, 2001, exhibited an 14 anadromous life history. Thus, it is recognized that both anadromous and resident life 15 history strategies of O. mykiss have been and continue to be present in the lower Yuba River. 16

The RMT (2013) developed representative temporal distributions for specific steelhead lifestages in the lower Yuba River through review of previously conducted studies, as well as recent and currently ongoing data collection activities of the M&E Program. As with spring-run Chinook salmon, the resultant lifestage periodicities are intended to encompass the majority of activity for a particular lifestage, and are not intended to be inclusive of every individual in the population. The lifestage-specific periodicities for steelhead in the lower Yuba River are summarized in **Table 4-7**, and are discussed below.

24 4.3.4.1 Adult Immigration and Holding

Adult migration from the ocean to spawning grounds occurs during much of the year, with peak migration occurring in the fall or early winter. Central Valley steelhead are known to use the Sacramento River as a migration corridor to spawning areas in upstream tributaries. Historically, steelhead likely did not utilize the mainstem Sacramento River downstream from the present location of Shasta Dam, except as a migration corridor to and from headwater streams (NMFS 2009).

| Lifestage | Ja | n | Feb | м | ar | A | or | Ма | ıy | Jι | ın | Jı | ıl | Au | ıg | Se | еp | 0 | ct | No | v | De | C |
|------------------------------|----|---|-----|---|----|---|----|----|----|----|----|----|----|----|----|----|----|---|----|----|---|----|---|
| Steelhead | | | | | | | | | | | | | | | | | | | | | | | |
| Adult Immigration & Holding | | | | | | | | | | | | | | | | | | | | | | | |
| Spawning | | | | | | | | | | | | | | | | | | | | | | | |
| Embryo Incubation | | | | | | | | | | | | | | | | | | | | | | | |
| Fry Rearing | | | | | | | | | | | | | | | | | | | | | | | |
| Juvenile Rearing | | | | | | | | | | | | | | | | | | | | | | | |
| Juvenile Downstream Movement | | | | | | | | | | | | | | | | | | | | | | | |

1 Table 4-7. Lifestage-specific periodicities for steelhead in the lower Yuba River 2 (Source: RMT 2013).

3 Migration through the Sacramento River mainstem begins in July, peaks at the end of 4 September, and continues through February or March (Bailey 1954; Hallock et al. 1961 5 both as cited in McEwan and Jackson 1996). Counts made at RBDD from 1969 through 6 1982 (Hallock 1989 as cited in McEwan and Jackson 1996) and on the Feather River 7 (Painter et al. 1977) follow the above pattern, although some fish were counted as late as 8 April and May. Weekly counts at Clough Dam on Mill Creek during a 10-year period 9 from 1953 to 1963 showed a similar migration pattern as well, with a peak in migration 10 during mid-November and another peak during February (NMFS 2009a). This second 11 peak is not reflected in counts made in the Sacramento River mainstem (Bailey 1954; 12 Hallock et al. 1961; both as cited in McEwan and Jackson 1996) or at RBDD (Hallock 13 1989 as cited in McEwan and Jackson 1996).

14 According to NMFS (2009a), Central Valley steelhead are mostly 'winter steelhead' and 15 may contain some 'summer steelhead' (the naming convention refers to the seasonal 16 period of adult upstream migration). Winter steelhead mature in the ocean and arrive on the spawning grounds nearly ready to spawn, whereas summer steelhead enter freshwater 17 with immature gonads and typically spend several months in freshwater before spawning. 18 19 The reported minimum depth for successful passage is about 7 inches (Reiser and Bjornn 1979 as cited in McEwan and Jackson 1996). Excessive water velocity (>10 to 13 ft/s) 20 21 and obstacles may prevent access to upstream spawning grounds (NMFS 2009a).

Smolt (Yearling+) Emigration

The optimal temperature range during adult upstream migration is unknown for Central
 Valley steelhead stocks (NMFS 2009a). Prolonged exposure to water temperatures above
 73°F is reported to be lethal to adult steelhead (Moyle 2002). Based on northern stocks,
 the optimal temperature range for migrating adult steelhead is 46 to 52°F (Bovee 1978;
 Reiser and Bjornn 1979; Bell 1986; all as cited in McEwan and Jackson 1996).

6 The immigration of adult steelhead in the lower Yuba River has been reported to occur 7 from August through March, with peak immigration from October through February 8 (CALFED and YCWA 2005; McEwan and Jackson 1996). CDFG (1984a) reported that 9 during the drought years of 1976-1977, two steelhead immigration peaks were observed – 10 one in October and one in February. CDFG (1991a) reported that steelhead enter the 11 lower Yuba River as early as August, migration peaks in October through February, and 12 may extend through March. In addition, they report that a run of "half-pounder" 13 steelhead occurred from late-June through the winter months.

14 The RMT (2010b) examined preliminary data and identified variable annual timing of O. 15 *mykiss* ascending the fish ladders at Daguerre Point Dam since the VAKI Riverwatcher 16 infrared and videographic sampling system began operations in 2003. For example, 17 Massa et al. (2010) state that peak passage of steelhead at Daguerre Point Dam occurred 18 from April through June during 2007. They also suggest that the apparent disparity 19 between the preliminary data and other reports of steelhead adult immigration periodicity 20 may be explained by the previously reported (Zimmerman et al. 2009; Mitchell 2010) 21 relatively high proportion of resident (vs. anadromous) O. mykiss occurring in the lower 22 Yuba River, because the VAKI Riverwatcher system did document larger (>40.6 cm) O. 23 *mykiss* ascending the fish ladders at Daguerre Point Dam during the winter months 24 (December through February). The observed timing of larger O. mykiss ascending the fish ladders at Daguerre Point Dam more closely corresponds with previously reported 25 26 adult steelhead immigration periodicities. The RMT (2010b; 2013) identified the period 27 extending from August through March as encompassing the majority of the upstream 28 migration and holding of adult steelhead in the lower Yuba River.

1 4.3.4.2 Adult Spawning

Central Valley adult steelhead generally begin spawning in late December and spawning
extends through March, but also can range from November through April (CDFG 1986).
Steelhead adults typically spawn from December through April with peaks from January
through March in small streams and tributaries where cool, well oxygenated water is
available year-round (Hallock et al. 1961; McEwan 2001). Based on all available
information collected to date, the RMT (2013) recently identified the steelhead spawning
period as extending from January through April.

9 Central Valley steelhead spawn downstream of dams on every major tributary within the
10 Sacramento and San Joaquin River systems. Due to water development projects, most
11 spawning is now confined to lower stream reaches below dams. In a few streams, such as
12 Mill and Deer Creeks, steelhead still have access to historical spawning areas (NMFS
13 2009a).

14 The female steelhead selects a site with good intergravel flow, digs a redd with her tail, 15 usually in the coarse gravel of the tail of a pool or in a riffle, and deposits eggs while an 16 attendant male fertilizes them (NMFS 2009). Spawning occurs mainly in gravel 17 substrates (particle size range of about 0.2-4.0 inches). Sand-gravel and gravel-cobble 18 substrates are also used, but these must be highly permeable and contain less than 5% 19 sand and silt for the water to be able to provide sufficient oxygen to the incubating eggs. 20 Adults tend to spawn in shallow areas (6–24 inches deep) with moderate water velocities 21 (about 1 to 3.6 ft/s) (Bovee 1978 as cited in McEwan and Jackson 1996; Hannon and 22 Deason 2007 as cited in Reclamation 2008). The optimal temperature range for 23 spawning has been reported to range from 39 to 52°F (Bovee 1978; Reiser and Bjornn 24 1979; Bell 1986 all as cited in McEwan and Jackson 1996). Egg mortality begins to 25 occur at 56°F (McEwan and Jackson 1996).

Unlike Chinook salmon, Central Valley steelhead may not die after spawning (McEwan and Jackson 1996). Some may return to the ocean and repeat the spawning cycle for two or three years. The percentage of adults surviving spawning is generally thought to be low for Central Valley steelhead, but varies annually and between stocks. Acoustic tagging of Central Valley steelhead kelts from the Coleman Hatchery indicates survival rates can be high, especially for Central Valley steelhead reconditioned by holding and
 feeding at the hatchery prior to release. Some return immediately to the ocean and some
 remain and rear in the Sacramento River (NMFS 2009a).

4 Steelhead spawning has been reported to generally extend from January through April in 5 the lower Yuba River (CALFED and YCWA 2005; CDFG 1991a; YCWA et al. 2007). 6 The RMT conducted a pilot redd survey from September 2008 through April 2009 (RMT 7 2010a). Surveys were not conducted during March, which is a known time for steelhead 8 spawning in other Central Valley rivers, due to high flows and turbidity. An extensive 9 area redd survey was conducted by surveyors kayaking from the downstream end of the 10 Narrows pool to the Simpson Lane Bridge. During the extensive area redd survey, redds 11 that were categorized as steelhead based on redd size criteria were reportedly observed 12 from October through April. However, some of those redds categorized as steelhead, 13 particularly during October, may actually have been small Chinook salmon redds because 14 the size criteria used to identify steelhead redds was found to be 53% accurate for 15 identifying steelhead redds in the Feather River (USFWS 2008a).

16 Campos and Massa (2010b and 2011) synthesized results of near-census redd surveys 17 conducted on the lower Yuba River during the 2009 and 2010 survey periods. During 18 both annual survey efforts, a substantial proportion of the weekly strata in the January 19 through April time periods were not sampled due to elevated flows and associated 20 turbidity levels. Nonetheless, RMT (2013) demonstrated that based upon cumulative 21 temporal distribution curves, the steelhead spawning period in the lower Yuba River is 22 generally characterized to extend from January through April.

23 Steelhead spawning has been reported to primarily occur in the lower Yuba River 24 upstream of Daguerre Point Dam (SWRI et al. 2000; YCWA et al. 2007). Kozlowski 25 (2004) states that field observations during winter and spring 2000 (YCWA unpublished 26 data) indicated that the majority of steelhead spawning in the lower Yuba River occurred 27 from Long Bar upstream to the Narrows, with the highest concentration of redds 28 observed upstream of the Highway 20 Bridge. USFWS (2007) data were collected on O. 29 mykiss redds in the lower Yuba River during 2002, 2003, and 2004, with approximately 30 98% of the redds located upstream of Daguerre Point Dam. During the pilot redd survey

conducted from the fall of 2008 through spring of 2009, the RMT (2010) report that most
(65%) of the steelhead redds were observed upstream of Daguerre Point Dam. Female
steelhead construct redds within a range of depths and velocities in suitable gravels,
oftentimes in pool tailouts and heads of riffles. In the lower Yuba River, steelhead have
also been observed to spawn in side channel areas (YCWA unpublished data).

6 4.3.4.3 Embryo Incubation

California Central Valley adult steelhead eggs incubate within the gravel and hatch from
approximately 19 to 80 days at water temperatures ranging from 60°F to 40°F,
respectively (NMFS 2009). After hatching, the young fish (alevins) remain in the gravel
for an extra two to six weeks before emerging from the gravel and taking up residence in
the shallow margins of the stream.

12 Steelhead embryo incubation generally occurs from December through June in the 13 Central Valley. The RMT (2013) identified the period of January through May as 14 encompassing the majority of the steelhead embryo incubation period in the lower Yuba 15 River. Following deposition of fertilized eggs in the redd, they are covered with loose 16 gravel. Central Valley steelhead eggs can reportedly survive at water temperature ranges 17 of 35.6°F to 59°F (Myrick and Cech 2001). Steelhead eggs reportedly have the highest 18 survival rates at water temperature ranges of 44.6°F to 50.0°F (Myrick and Cech 2001). 19 Studies conducted at or near 54.0°F report high survival and normal development of 20 steelhead incubating embryos, a relatively low mortality of incubating steelhead embryos 21 is reported to occur at 57.2°F, and a sharp decrease in survival has been reported for O. 22 *mykiss* embryos incubated above 57.2°F (RMT 2010b).

Steelhead eggs hatch in three to four weeks at 50°F to 59°F, and fry emerge from the gravel four to six weeks later (Shapovalov and Taft 1954). Steelhead embryo development requires a constant supply of well oxygenated water. This implies a loose gravel substrate allowing high permeability, with little silt or sand deposition during the development time period. Merz et al. (2004) showed that spawning substrate quality influenced a number of physical parameters affecting egg survival including temperature, dissolved oxygen, and substrate permeability. The entire egg incubation lifestage encompasses the time when adult steelhead spawn
 through the time when emergent fry exit the gravel (CALFED and YCWA 2005). In the
 lower Yuba River, steelhead embryo incubation generally occurs from January through
 May (CALFED and YCWA 2005; SWRI 2002).

5 4.3.4.4 Juvenile Rearing and Outmigration

6 As reported in NMFS (2009a), juvenile Central Valley steelhead may migrate to the 7 ocean after spending one to three years in freshwater (McEwan and Jackson 1996). Upon 8 emergence from the gravel, the fry move to shallow protected areas associated with the 9 stream margin (Royal 1972; Barnhart 1986; both as cited in McEwan and Jackson 1996). 10 Steelhead fry tend to inhabit areas with cobble-rubble substrate, a depth less than 14 11 inches, and temperature ranging from 45 to 60°F (Bovee 1978 as cited in McEwan and 12 Jackson 1996). Myrick (1998, as cited in Reclamation 2008) found steelhead from the 13 Feather and Mokelumne rivers preferred temperatures between 62.5°F and 68°F.

In general, it has been reported that after emergence steelhead fry move to shallow-water, low velocity habitats, such as stream margins and low gradient riffles, and will forage in open areas lacking instream cover (Hartman 1965; Everest et al. 1986; Fontaine 1988). As fry increase in size and their swimming abilities improve in late summer and fall, juvenile steelhead have been reported to increasingly use areas with cover and show a preference for higher velocity, deeper mid-channel areas near the thalweg (Hartman 1965; Everest and Chapman 1972; Fontaine 1988).

Juvenile steelhead have been reported to occupy a wide range of habitats, preferring deep pools as well as higher velocity rapid and cascade habitats (Bisson et al. 1982; 1988). During the winter period of inactivity, steelhead prefer low velocity pool habitats with large rocky substrate or woody debris for cover (Hartman 1965; Swales et al. 1986; Raleigh et al. 1984; Fontaine 1988). During periods of low temperatures and high flows associated with the winter months, juvenile steelhead seek refuge in interstitial spaces in cobble and boulder substrates (Bustard and Narver 1975; Everest et al. 1986).

Older juveniles use riffles and larger juveniles may also use pools and deeper runs
(Barnhart 1986 as cited in McEwan and Jackson 1996). However, specific depths and

1 habitats used by juvenile rainbow trout can be affected by predation risk (Brown and 2 Brasher 1995). Central Valley steelhead can show mortality at constant temperatures of 3 77°F although they can tolerate 85°F for short periods (Myrick and Cech 2001). Juvenile 4 steelhead in northern California rivers reportedly exhibited increased physiological stress, 5 increased agonistic activity, and a decrease in forage activity after ambient stream 6 temperatures exceeded 71.6°F (Nielsen et al. 1994). Hatchery reared steelhead in thermal 7 gradients selected temperatures of 64-66°F while wild caught steelhead selected temperatures around 63°F (Myrick and Cech 2001). An upper water temperature limit of 8 9 65°F is preferred for growth and development of Sacramento River and American River 10 juvenile steelhead (NMFS 2002a).

In the lower Yuba River, juvenile steelhead exhibit variable durations of rearing. The RMT (2010b) distinguished fry, juvenile, and yearling+ lifestages through evaluation of bi-weekly length-frequency distributions of *O. mykiss* captured in rotary screw traps in the lower Yuba River, and other studies that report length-frequency estimates (Mitchell 2010; CDFG 1984a). Some juvenile *O. mykiss* may rear in the lower Yuba River for short periods (up to a few months) and others may spend from one to three years rearing in the river.

18 Some age-0 *O. mykiss* disperse downstream soon after emerging and continue throughout 19 the year (Kozlowski 2004). Thus, the steelhead fry (individuals less than about 45 mm) 20 lifestage generally extends from the time of initial emergence (based upon accumulated 21 thermal units from the time of egg deposition through hatching and alevin incubation) 22 until three months following the end of the spawning period. YCWA (2010) identified 23 the fry rearing lifestage as generally extending from mid-March through July, and 24 identified the juvenile rearing lifestage as extending year-round. Based on all 25 information collected to date, the RMT (2013) identified the steelhead fry rearing period 26 as extending from April through July.

Juvenile steelhead have been reported to rear in the lower Yuba River for up to 1 year or more (SWRI 2002). CDFG (1991a) reported that juvenile steelhead rear throughout the year in the lower Yuba River, and may spend from 1 to 3 years rearing in the river. Scale analysis conducted by Mitchell (2010) indicates the presence of at least four age categories for *O. mykiss* in the lower Yuba River that spent 1, 2, or 3 years in freshwater
 and 1 year at sea before returning to the lower Yuba River to spawn.

3 Based on the combined results from electrofishing and snorkeling surveys conducted 4 during the late 1980s, CDFG (1991a) reported that juvenile steelhead were observed in 5 all river reaches downstream of the Englebright Dam and, in addition to Chinook salmon, 6 were the only fish species observed in the Narrows Reach. They also indicated that most 7 juvenile steelhead rearing occurred above Daguerre Point Dam. SWRI et al. (2000) 8 summarized data collection in the lower Yuba River obtained from 1992 through 2000. 9 Since 1992, Jones and Stokes Associates (JSA) biologists conducted fish population 10 surveys in the lower Yuba River using snorkel surveys to determine annual and seasonal 11 patterns of abundance and distribution of juvenile O. mykiss (and Chinook salmon) 12 during the spring and summer rearing periods. The primary rearing habitat for juvenile 13 O. mykiss is upstream of Daguerre Point Dam. In 1993 and 1994, snorkeling surveys 14 indicated that the population densities and overall abundance of juvenile O. mykiss (age 0 15 and 1+) were substantially higher upstream of Daguerre Point Dam, with decreasing 16 abundance downstream of Daguerre Point Dam.

17 Similarly, Kozlowski (2004) found higher abundances of juvenile O. mykiss above 18 Daguerre Point Dam, relative to downstream of Daguerre Point Dam. Kozlowski (2004) 19 observed age-0 O. mykiss throughout the entire study area, with highest densities in 20 upstream habitats and declining densities with increasing distance from the Narrows. 21 Approximately 82% of juvenile O. mykiss were observed upstream of Daguerre Point 22 Dam. Kozlowski (2004) suggested that the distribution of age-0 O. mykiss appeared to be 23 related to the distribution of spawning adults. SWRI et al. (2000) suggested that higher 24 abundances of juvenile O. mykiss above Daguerre Point Dam may have been due to larger numbers of spawners, greater amounts of more complex, high quality cover, and 25 26 lower densities of predators such as striped bass and American shad, which reportedly 27 were restricted to areas below Daguerre Point Dam.

In the lower Yuba River, Kozlowski (2004) reports that juvenile *O. mykiss* were observed in greater numbers in pool habitats than in run habitats. He suggests that results of his study indicated a relatively higher degree of habitat complexity, suitable for various lifestages, in the reaches just below the Narrows compared to farther downstream. The
 Narrows reach includes greater occurrence of pool-type microhabitat suitable for juvenile
 O. mykiss rearing, as well as small boulders and cobbles preferred by the age-0 emerging
 lifestage (Kozlowski 2004).

Juvenile *O. mykiss* apparently demonstrate a proclivity for near-bank areas, rather than
open-channel habitats, in the lower Yuba River. USFWS (2008a) reports 258
observations of juvenile *O. mykiss* and 244 observations of juvenile Chinook salmon, all
but 8 of them made near the river banks in the lower Yuba River.

9 A broad range of O. mykiss size classes have been observed in the lower Yuba River 10 during spring and summer snorkeling, electrofishing, and angling surveys (SWRI et al. 11 2000). Juvenile O. mykiss ranging in size from 40-150 mm were commonly observed 12 upstream of Daguerre Point Dam. Numerous larger juveniles and resident trout up to 18 13 inches long were also commonly observed in the mainstem upstream and downstream of 14 Daguerre Point Dam (SWRI et al. 2000). Age 0 (young-of-the-year) O. mykiss were 15 clearly shown by the distinct mode in lengths of fish caught by electrofishing (40-100 16 mm fork length). A preliminary examination of scales indicated that most yearling (age 17 1+) and older O. mykiss were represented by fish greater than 110 mm long, including 18 most if not all of the fish caught by hook and line. The sizes of age 0 and 1+ O. mykiss 19 indicated substantial annual growth of O. mykiss in the lower Yuba River. Seasonal 20 growth of age 0 O. mykiss was evident from repeated sampling in 1992 and 1999, but 21 actual growth rates could not be estimated because of continued recruitment of fry (newly 22 emerged juveniles) or insufficient sample sizes (SWRI et al. 2000).

23 Mitchell (2010) reports that analysis of scale growth patterns of juvenile O. mykiss in the 24 lower Yuba River indicates a period of accelerated growth during the spring peaking 25 during the summer months, followed by decelerated growth during the fall and winter. 26 Following the second winter, juvenile O. mykiss in the lower Yuba River exhibit reduced 27 annual growth in length with continued growth in mass until reaching reproductive age. 28 Additionally, more rapid juvenile and adult O. mykiss growth occurred in the lower Yuba 29 River compared to the lower Sacramento River and Klamath River O. mykiss, with 30 comparable growth rates to O. mykiss in the upper Sacramento River (Mitchell 2010).

1 CDFG (1991a) reports that juvenile steelhead in the lower Yuba River rear throughout 2 the year, and may spend from one to three years in the river before emigrating primarily 3 from March to June. Salvage data at the Hallwood-Cordua fish screen suggest that most 4 juvenile fish initiated their downstream movements immediately preceding and following 5 a new moon, indicating the presence of lunar periodicity in the timing or outmigration 6 patterns in the lower Yuba River (Kozlowski 2004).

Based on all information collected to date, the RMT (2013) identified the steelhead
juvenile rearing period as extending year-round, and the steelhead juvenile downstream
movement period as extending from April through September.

10 In the lower Yuba River, some young-of-year (YOY) O. mykiss are captured in rotary 11 screw traps (RSTs) located downstream of Daguerre Point Dam during late-spring and 12 summer, indicating movement downstream. However, at least some of this downstream movement may be associated with the pattern of flows in the river. Water transfer 13 14 monitoring in 2001, 2002, and 2004 (YCWA and SWRCB 2001; YCWA 2003; YCWA 15 2005), generally from about mid-June through September, indicated that the character of 16 the initiation of the water transfers could potentially affect juvenile O. mykiss 17 downstream movement. Based upon the substantial differences in juvenile O. mykiss 18 downstream movements (RST catch data) noted between the 2001 study, and the 2002 19 and 2004 studies, it was apparent that the increases in juvenile O. mykiss downstream 20 movement associated with the initiation of the 2001 water transfers were avoided due to a 21 more gradual ramping-up of flows that occurred in 2002 and 2004 (YCWA et al. 2007).

22 Numerous studies have been conducted regarding temperature preference, mortality, and 23 water temperature growth-related relationships for O. mykiss. As previously described, 24 some steelhead may rear in freshwater for up to three years before emigrating as 25 yearling+ smolts, whereas other individuals move downstream shortly after emergence as 26 post-emergent fry, or rear in the river for several months and move downstream as 27 juveniles without exhibiting the ontogenetic characteristics of smolts. Presumably, these 28 individuals continue to rear and grow in downstream areas (e.g., lower Feather River, 29 Sacramento River, and Upper Delta) and undergo the smoltification process prior to entry

into saline environments. Thus, fry and juvenile rearing occur concurrently with post emergent fry and juvenile downstream movement.

3 4.3.4.5 Smolt Emigration

4 Most juvenile steelhead spend one to three years in fresh water before emigrating to the 5 ocean as smolts (Shapovalov and Taft 1954). During their downstream migration, 6 juvenile steelhead undergo a process referred to as smoltification, which is a physiologic 7 transformation and osmoregulatory pre-adaptation to residence in saline environs. 8 Physiologic expressions of smoltification include increased gill ATPase and thyroxin 9 levels, and more slender body form which is silvery in appearance. The primary period 10 of steelhead smolt outmigration from rivers and creeks to the ocean generally occurs 11 from January to June (NMFS 2009).

In the Sacramento River, juvenile steelhead migrate to the ocean in spring and early summer at 1 to 3 years of age with peak migration through the Delta in March and April (Reynolds et al. 1993 as cited in NMFS 2009). Hallock et al. (1961) found that juvenile steelhead in the Sacramento River Basin migrate downstream during most months of the year, but the peak emigration period occurred in the spring, with a much smaller peak in the fall (NMFS 2009).

18 According to NMFS (2009a), steelhead are present at Chipps Island between at least 19 October and July, according to catch data from the USFWS Chipps Island Trawl. It 20 appears that adipose fin-clipped steelhead have a different emigration pattern than 21 unclipped steelhead. Adipose fin-clipped steelhead showed distinct peaks in catch 22 between January and March corresponding with time of release, whereas unclipped 23 steelhead were more evenly distributed over a period of six months or more. These 24 differences are likely an artifact of the method and timing of hatchery releases (NMFS 25 2009a).

Steelhead successfully smolt at water temperatures in the 43.7°F to 52.3°F range (Myrick and Cech 2001). The optimum water temperature range for successful smoltification in young steelhead has been reported as 44.0°F to 52.3°F (Rich 1987 as cited in NMFS 2009). Wagner (1974) reported smolting ceased rather abruptly when water temperatures
increased to 57°F-64°F. NMFS (2009a) reported that water temperatures under 57°F are
 considered best for smolting.

In the lower Yuba River, the steelhead smolt emigration period has been reported to extend from October through May (CALFED and YCWA 2005; SWRI 2002; YCWA et al. 2007). The RMT's (2010b; 2013) review of all available data indicate that yearling+ steelhead smolt emigration may extend from October through mid-April.

For the purposes of impact assessment, the RMT (2010b) developed separate water temperature index values for the yearling+ smolt emigration lifestages distinct from values for juvenile steelhead rearing and/or outmigration as juveniles from the lower Yuba River. They assumed that juvenile steelhead that exhibit extended rearing in the lower Yuba River undergo the smoltification process and volitionally emigrate from the river as yearling+ individuals.

13 **4.3.4.6** Lifestage-Specific Water Temperature Suitabilities

14 Since the RMT prepared its November 2010 water temperature objectives memorandum, 15 additional water temperature monitoring and life history investigations of anadromous 16 salmonids in the lower Yuba River have been conducted by the RMT. Through review of 17 previously conducted studies, as well as recent and currently ongoing data collection 18 activities of the M&E Program, the RMT (2013) developed the following representative 19 steelhead lifestage-specific periodicities and primary locations for water temperature 20 suitability evaluations. The locations used for water temperature evaluations correspond 21 to Smartsville, Daguerre Point Dam, and Marysville.

- Adult Immigration and Holding (August through March) Smartsville, Daguerre
 Point Dam, and Marysville
- 24 D Spawning (January through April) Smartsville and Daguerre Point Dam
- 25 📮 Embryo Incubation (January through May) Smartsville and Daguerre Point Dam
- 26 Juvenile Rearing and Downstream Movement (Year-round) Daguerre Point
 27 Dam and Marysville

Smolt (Yearling+) emigration (October through mid-April) – Daguerre Point Dam
 and Marysville

3 Steelhead lifestage-specific WTI values are provided in **Table 4-8**. The lifestages and 4 periodicities presented in Table 4-8 differ from those presented in Table 4-7 due to 5 specific lifestages that have the same or distinct upper tolerable WTI values.

| Lifestage | Upper Tolerance WTI | Ja | an | F | eb | Ма | r | A | or | M | ay | Jı | un | J | ul | Αι | ug | Se | əp | 0 | ct | N | v | D | ec |
|---|---------------------------|----|----|---|----|----|---|---|----|---|----|----|----|---|----|----|----|----|----|---|----|---|---|---|----|
| Adult Migration | 68°F | | | | | | | | | | | | | | | | | | | | | | | | |
| Adult Holding | 65°F | | | | | | | | | | | | | | | | | | | | | _ | | | |
| Spawning | 57°F | | | | | | | | | | | | | | | | | | | | | | | | |
| Embryo Incubation | 57°F | | | | | | | | | | | | | | | | | | | | | | | | |
| Juvenile Rearing and Downstream Movement | 68°F | | - | L | | | _ | | L | _ | _ | | | - | _ | _ | _ | _ | _ | | | _ | _ | | |
| Smolt (Yearling+) Emigration | 55°F | | | | | - | - | | | | | | | | | | | | | Γ | _ | _ | | | |

6 Table 4-8. Steelhead lifestage-specific upper tolerance WTI values.

7 Recent water temperature monitoring data in the lower Yuba River are available for the 8 period extending from 2006 into June 2013, during which time operations have complied 9 with the Yuba Accord. Figure 4-9 displays daily water temperature monitoring results 10 from October 2006 through June 2013 at Smartsville, Daguerre Point Dam, and 11 Marysville water temperature gages, with steelhead lifestage-specific upper tolerance 12 WTI values. Water temperatures at all three gages are always below the upper tolerance 13 WTI values for juvenile rearing and downstream movement, and adult immigration and 14 holding. The upper tolerance spawning and embryo incubation WTI value is never 15 exceeded at Smartsville, and is generally not exceeded at Daguerre Point Dam with the 16 exception of the end of May of some years. The smolt (yearling+) emigration upper 17 tolerance WTI value generally is not exceeded at the Smartsville Gage, and is not exceeded at the Daguerre Point Dam and Marysville gages after mid-November. 18



1

2 Figure 4-9. Lower Yuba River monitored water temperatures and steelhead upper 3 tolerance WTI values.

4 4.3.5 Limiting Factors, Threats and Stressors

As stated by NMFS (2005b), the factors affecting the survival and recovery of Central 5 6 Valley steelhead and their habitat are similar to those affecting spring-run Chinook 7 salmon and are primarily associated with habitat loss (McEwan 2001). McEwan and 8 Jackson (1996) attribute this habitat loss and other impacts to steelhead habitat primarily 9 to water development resulting in inadequate flows, flow fluctuations, blockages, and 10 entrainment into diversions. Other effects on critical habitat related to land use practices 11 and urbanization have also contributed to steelhead declines (Busby et al. 1996). 12 Although many of the factors affecting spring-run Chinook salmon habitat are common 13 to steelhead, some stressors, especially summer water temperatures, cause greater effects 14 to steelhead because juvenile steelhead rear in freshwater for more than one year. 15 Because most suitable habitat has been lost to dam construction, juvenile steelhead 16 rearing is generally confined to lower elevation stream reaches, where water temperatures 17 during late summer and early fall can be sub-optimal (NMFS 2005b).

18 Many of the improvements to critical habitat that have benefited spring-run Chinook 19 salmon, including water management through the CVPIA Section 3406(b)(2) water supply and the CALFED Environmental Water Account, improved screening conditions at water diversions, and changes in inland fishing regulations (there is no ocean steelhead fishery) also benefit Central Valley steelhead (NMFS 2005b). However, many dams and reservoirs in the Central Valley do not have water storage capacity or release mechanisms necessary to maintain suitable water temperatures for steelhead rearing through the critical summer and fall periods, especially during critically dry years (McEwan 2001).

7 4.3.5.1 DPS

According to the NMFS Draft Recovery Plan (NMFS 2009), threats to Central Valley steelhead are similar to those for spring-run Chinook salmon and fall into three broad categories: (1) loss of historical spawning habitat; (2) degradation of remaining habitat; and (3) threats to the genetic integrity of the wild spawning populations from hatchery steelhead production programs in the Central Valley. Also, as for spring-run Chinook salmon, the potential effects of long-term climate change also may adversely affect steelhead and their recovery.

15 In 1998, NMFS concluded that the risks to Central Valley steelhead had diminished, 16 based on a review of existing and recently implemented state conservation efforts and 17 federal management programs (e.g., CVPIA, AFRP, CALFED) that address key factors 18 for the decline of this species (NMFS 2009). NMFS stated that Central Valley steelhead 19 were benefiting from two major conservation initiatives, being simultaneously 20 implemented: (1) the CVPIA, which was passed by Congress in 1992; and (2) the 21 CALFED Program, a joint state/federal effort implemented in 1995. The following 22 discussion of these two programs was taken directly from NMFS (2009).

23 The CVPIA is specifically intended to remedy habitat and other problems associated with 24 the construction and operation of the CVP. The CVPIA has two key features related to 25 steelhead. First, it directs the Secretary of the Interior to develop and implement a 26 program that makes all reasonable efforts to double natural production of anadromous 27 fish in Central Valley streams (Section 3406(b)(1)) by the year 2002. The AFRP was 28 initially drafted in 1995 and subsequently revised in 1997. Funding has been 29 appropriated since 1995 to implement restoration projects identified in the AFRP 30 Second, the CVPIA dedicates up to 800,000 acre-feet of water planning process.

annually for fish, wildlife, and habitat restoration purposes (Section 3406(b)(2)) and
provides for the acquisition of additional water to supplement the 800,000 acre-feet
(Section 3406(b)(3)). USFWS, in consultation with other federal and state agencies, has
directed the use of this dedicated water yield since 1993.

5 The CALFED Program, which began in June 1995, was charged with the responsibility 6 of developing a long-term Bay-Delta solution. A major element of the CALFED 7 Program is the Ecosystem Restoration Program (ERP), which was intended to provide the 8 foundation for long-term ecosystem and water quality restoration and protection 9 throughout the region. Among the non-flow factors causing decline that have been 10 targeted by the program are unscreened diversions, waste discharges and water pollution, 11 impacts due to poaching, land derived salts, exotic species, fish barriers, channel 12 alterations, loss of riparian wetlands, and other causes of estuarine habitat degradation. 13 The level of risk faced by the Central Valley steelhead DPS may have diminished since 14 the 1996 listing proposal as a result of habitat restoration and other measures that have 15 recently been implemented through the CALFED and CVPIA programs. Although most 16 restoration measures designed to recover Chinook salmon stocks can benefit steelhead, 17 focusing restoration solely on Chinook salmon may lead to inadequate measures to 18 restore steelhead because of their different life histories and resource requirements, 19 particularly for rearing juveniles (McEwan 2001). Additional actions that benefit Central 20 Valley steelhead include efforts to enhance fisheries monitoring, such as the Central 21 Steelhead Monitoring Plan, actions Valley and conservation to address 22 artificial propagation.

23 In spite of the benefits derived from implementation of these two programs, NMFS 24 (2009) identified several major stressors presently applicable to the entire Central Valley 25 steelhead DPS. Many of the most important stressors specific to the steelhead DPS 26 correspond to the stressors described for the spring-run Chinook salmon ESU. As 27 previously stated, the 2009 NMFS OCAP BO (2009a) identified factors leading to the 28 current status of the spring-run Chinook salmon ESU, which also are applicable to the 29 steelhead DPS, including habitat blockage, water development and diversion dams, water 30 conveyance and flood control, land use activities, water quality, hatchery operations and 31 practices, over-utilization (e.g., ocean commercial and sport harvest, inland sport 1 harvest), disease and predation, environmental variation (e.g., natural environmental 2 cycles, ocean productivity, climate change), and non-native invasive species. The 3 previous discussions in this BA addressing limiting factors and threats for the spring-run 4 Chinook salmon ESU and their specific geographic influences, including the Sacramento 5 River and the Delta, are not repeated in this section of this BA. Stressors that are unique 6 to the steelhead DPS, or substantially differ in the severity from the stressor for the previously described spring-run Chinook salmon ESU, are described below. 7

8 Threats and stressors for the Central Valley steelhead DPS identified in Appendix B 9 (Threats Assessment) of the NMFS Draft Recovery Plan (NMFS 2009) include: (1) 10 destruction, modification, or curtailment of habitat or range; (2) overutilization for 11 commercial, recreational, scientific or education purposes; (3) disease or predation; (4) 12 inadequacy of existing regulatory mechanisms, including federal and non-federal efforts; 13 (5) other natural and man-made factors affecting its continued existence; and (6) non-14 lifestage specific threats and stressors including artificial propagation programs, small 15 population size, genetic integrity and long-term climate change. The following 16 summarization of threats and stressors for the Central Valley steelhead DPS is taken 17 directly from Appendix B (Threats Assessment) of the NMFS Draft Recovery Plan 18 (NMFS 2009).

19 DESTRUCTION, MODIFICATION, OR CURTAILMENT OF HABITAT OR RANGE

20 The spawning habitat for Central Valley steelhead has been greatly reduced from its 21 historical range (NMFS 2009). The vast majority of historical spawning habitat for 22 Central Valley steelhead has been eliminated by fish passage impediments associated 23 with water storage, withdrawal, conveyance, and diversions for agriculture, flood control, 24 and domestic and hydropower purposes (NMFS 2009). Modification of natural flow 25 regimes has resulted in increased water temperatures, changes in fish community 26 structures, depleted flow necessary for migration, spawning, rearing, and flushing of 27 sediments from spawning gravels. These changes in flow regimes may be driving a shift 28 in the frequencies of various life history strategies, especially a decline in the proportion 29 of the population migrating to the ocean. Land use activities, such as those associated 30 with agriculture and urban development, have altered steelhead habitat quantity and

quality. Although many historically harmful practices have been halted, much of the
 historical damage to habitats limiting steelhead remains to be addressed, and the
 necessary restoration activities will likely require decades.

4 OVERUTILIZATION FOR COMMERCIAL, RECREATIONAL, SCIENTIFIC OR EDUCATIONAL PURPOSES 5 (INLAND SPORT HARVEST)

6 Steelhead have been, and continue to be, an important recreational fishery throughout 7 their range. Although there are no commercial fisheries for steelhead in the ocean, inland 8 steelhead fisheries include tribal and recreational fisheries. In the Central Valley, 9 recreational fishing for steelhead is popular, yet harvest is restricted to only the visibly 10 marked hatchery-origin fish, which reduces the likelihood of retaining naturally spawned 11 The permits NMFS issues for scientific or educational purposes stipulate wild fish. 12 specific conditions to minimize take of steelhead individuals during permitted activities. 13 There are currently 11 active permits in the Central Valley that may affect steelhead. 14 These permitted studies provide information about Central Valley steelhead that is useful 15 to the management and conservation of the DPS. [Additional information regarding 16 inland sport harvest of steelhead in the Central Valley contained in Reclamation (2008) is 17 provided below.]

18 INLAND SPORT HARVEST

19 Historically in California, almost half of the river sport fishing effort has occurred in the 20 Sacramento-San Joaquin River system, particularly upstream from the city of Sacramento 21 (Emmett et al. 1991). There is little information on steelhead harvest rates in California. 22 Hallock et al. (1961) estimated that harvest rates for Sacramento River steelhead from the 1953/1954 through 1958/1959 seasons ranged from 25.1 to 45.6% assuming a 20% non-23 24 return rate of tags. The average annual harvest rate of adult steelhead above RBDD for 25 the 3-year period from 1991/1992 through 1993/1994 was 16% (McEwan and Jackson 26 1996). Since 1998, all hatchery steelhead have been marked with an adipose fin clip 27 allowing anglers to distinguish hatchery and wild steelhead. Current regulations restrict 28 anglers from keeping unmarked steelhead in Central Valley streams. Overall, this 29 regulation has greatly increased protection of naturally produced adult steelhead 30 (Reclamation 2008). However, the total number of steelhead contacted might be a

significant fraction of basin-wide escapement, and even low catch-and-release mortality
 may pose a problem for wild populations (Good et al. 2005).

3 DISEASE OR PREDATION

4 Steelhead are exposed to bacterial, protozoan, viral, and parasitic organisms in spawning 5 and rearing areas, hatcheries, migratory routes, and the marine environment. Very little 6 current or historical information exists to quantify changes in infection levels and 7 mortality rates attributable to these diseases for steelhead. Naturally spawned fish tend to 8 be less susceptible to pathogens than hatchery-reared fish. Introduction of non-native 9 species and modification of habitat have resulted in increased predatory populations and 10 salmonid predation in river systems. In general, predation rates on steelhead are 11 considered to be an insignificant contribution to the large declines observed in West 12 Coast steelhead populations. In some local populations, however, predation may 13 significantly influence salmonid abundance when other prey species are not present and 14 habitat conditions lead to the concentration of adults and/or juveniles.

15 INADEQUACY OF EXISTING REGULATORY MECHANISMS (FEDERAL EFFORTS, NON-FEDERAL EFFORTS)

16 FEDERAL EFFORTS

17 There have been several federal actions attempting to reduce threats to the Central Valley 18 steelhead DPS. The BOs for the CVP and SWP and other federal projects involving 19 irrigation and water diversion and fish passage, for example, have improved or 20 minimized adverse impacts to steelhead in the Central Valley. There have also been 21 several habitat restoration efforts implemented under CVPIA and CALFED programs 22 that have led to several projects involving fish passage improvements, fish screens, 23 floodplain management, habitat restoration, watershed planning, and other projects that 24 have contributed to improvement of steelhead habitat. However, despite federal actions 25 to reduce threats to the Central Valley steelhead DPS, the existing protective efforts are 26 inadequate to ensure the DPS is no longer in danger of extinction. There remain high 27 risks to the abundance, productivity, and spatial structure of the steelhead DPS.

1 Non-Federal Efforts

2 Measures to protect steelhead throughout the State of California have been in place since The State's Natural Communities Conservation Planning (NCCP) program 3 1998. involves long-term planning with several stakeholders. A wide range of measures have 4 5 been implemented, including 100% marking of all hatchery steelhead, zero bag limits for 6 unmarked steelhead, gear restrictions, closures, and size limits designed to protect smolts. 7 NMFS and CDFW are working to improve inland fishing regulations to better protect 8 both anadromous and resident forms of O. mykiss populations. A proposal to develop a 9 comprehensive status and trends monitoring plan for Central Valley steelhead was 10 submitted for funding consideration to the CALFED ERP in 2005. The proposal, drafted 11 by CDFW and the interagency Central Valley Steelhead Project Work Team, was 12 selected by the ERP Implementing Agency Managers, and is to receive funding as a 13 directed action. Long-term funding for implementation of the monitoring plan, once it is 14 developed, still needs to be secured. There are many sub-watershed groups, landowners, 15 environmental groups, and non-profit organizations that are conducting habitat 16 restoration and planning efforts that may contribute to the conservation of steelhead. 17 However, despite federal and non-federal efforts to promote the conservation of the 18 Central Valley steelhead DPS, few efforts address conservation needs at scales sufficient 19 to protect the entire steelhead DPS. The lack of status and trend monitoring and research 20 is one of the critical limiting factors to this DPS.

21 OTHER NATURAL AND MAN-MADE FACTORS AFFECTING THE CONTINUED EXISTENCE OF THE DPS

22 NMFS and the Biological Review Team (BRT) are concerned that the proportion of 23 naturally produced fish is declining. Two artificial propagation programs for steelhead in 24 the Central Valley - Coleman National Fish Hatchery and FRFH - may decrease risk to 25 the DPS to some degree by contributing increased abundance to the DPS. Potential 26 threats to natural steelhead posed by hatchery programs include: (1) mortality of natural 27 steelhead in fisheries targeting hatchery-origin steelhead; (2) competition for prey and 28 habitat; (3) predation by hatchery-origin fish on younger natural fish; (4) genetic 29 introgression by hatchery-origin fish that spawn naturally and interbreed with local 30 natural populations; and (5) disease transmission.

1 Changes in climatic events and global climate, such as El Niño ocean conditions and 2 prolonged drought conditions, can threaten the survival of steelhead populations already 3 reduced to low abundance levels as the result of the loss and degradation of freshwater 4 and estuarine habitats. Floods and persistent drought conditions have reduced already 5 limited spawning, rearing, and migration habitats. Unscreened water diversions and CVP 6 and SWP pumping plants entrain outmigrating juvenile steelhead and fry, leading to 7 fish mortality.

8 <u>NON-LIFESTAGE SPECIFIC THREATS AND STRESSORS FOR THE DPS (ARTIFICIAL PROPAGATION</u> 9 <u>PROGRAMS, SMALL POPULATION SIZE, GENETIC INTEGRITY AND LONG-TERM CLIMATE CHANGE</u>)

Potential threats to the Central Valley steelhead population that are not specific to a particular lifestage include the potential negative impacts of the current artificial propagation program utilizing several hatcheries in the Sacramento-San Joaquin drainage, the small wild population size, the genetic integrity of the population due to both hatchery influence and small population size, and the potential effects of long-term climate change. Each of these potential threats is discussed in the following sections.

16 Artificial Propagation Program

17 Recent research has indicated that approximately 63 to 92% of steelhead smolt 18 production is of hatchery-origin (NMFS 2003). These data suggest that the relative 19 proportion of wild to hatchery smolt production is decreasing (NMFS 2003). All 20 California hatchery steelhead programs began 100% adipose fin-clipping in 1998 to 21 differentiate between hatchery steelhead from natural steelhead.

22 Propagation of steelhead at the Coleman National Fish Hatchery has been occurring for 23 over 50 years. Hatchery-origin and natural-origin steelhead have been managed as a 24 single stock; mixing of hatchery and natural origin population components occurred 25 through spawning at the hatchery and intermingling with natural spawners in Battle 26 Niemela et al. (2008) used genetic pedigree analysis to evaluate relative Creek. 27 reproductive success and fitness among hatchery-origin and natural origin population 28 components based on multilocus DNA microsatellite genotypes. Preliminary results 29 suggest that hatchery origin spawners experienced low relative reproductive success, 30 producing significantly fewer adult offspring in comparison to natural origin spawners.

Additionally, repeat spawning was more prevalent in the natural origin component of
 the population.

3 POPULATION SIZE

4 In the technical memorandum titled Updated Status of Federally Listed ESUs of West 5 Coast Salmon and Steelhead (Good et al. 2005), NMFS estimated the abundance of 6 natural spawners for the steelhead DPS (then classified as an ESU), which was reported 7 as the geometric mean (and range) of the most recent data available at that time, 8 consistent with previous coast-wide status reviews of the species (Weitkamp et al. 1995; 9 Busby et al. 1996; Gustafson et al. 1997; Johnson et al. 1997; Myers et al. 1998). 10 Geometric means were calculated to represent the abundance of natural spawners for 11 each population or quasi-population. Geometric means were calculated for the most 12 recent 5 years of steelhead data, to correspond with modal age at maturity (Good et al. 13 2005). Where possible, the BRTs obtained population or ESU-level estimates of the 14 fraction of hatchery-origin spawners or calculated estimates from information using scale 15 analyses, fin clips, etc. (Good et al. 2005).

16 The Central Valley steelhead DPS mean annual escapement of natural spawners was 17 estimated at 1,952 based on a 5-year period ending in 1993 (Good et al. 2005). During 18 that time period a minimum escapement of 1,425 and a maximum escapement of 12,320 19 were observed (Good et al. 2005). A long-term trend analysis indicated that the 20 population was declining (Good et al. 2005). In the Updated Status of Federally Listed 21 ESUs of West Coast Salmon and Steelhead (Good et al. 2005), NMFS suggests that there 22 has been no significant status change since the 1993 data and the Central Valley steelhead 23 population continues to decline (Good et al. 2005). Good et al. (2005) also suggested that 24 hatchery production is large relative to natural production. As an example, the steelhead 25 run in the lower Feather River has been increasing over the past several years; however, 26 over 99% of the run is of direct hatchery-origin (DWR 2002).

27 GENETIC INTEGRITY

There is still significant local genetic structure to Central Valley steelhead populations,although fish from the San Joaquin and Sacramento basins cannot be distinguished

1 genetically (Nielsen et al. 2003). Hatchery effects appear to be localized – for example, 2 Feather River and FRFH steelhead are closely related as are American River and Nimbus 3 Hatchery fish (DWR 2002). Leary et al. (1995) report that hatchery straying has 4 increased gene flow among steelhead populations in the Central Valley and that a smaller 5 amount of genetic divergence is observed among Central Valley populations compared to 6 wild British Columbia populations largely uninfluenced by hatcheries. Natural annual production of steelhead smolts in the Central Valley is estimated at 181,000 and hatchery 7 8 production is 1,340,000 for a ratio of 0.148 (Good et al. 2005). Current monitoring by 9 hydroacoustic tracking has revealed that Mokelumne River/Hatchery steelhead (FRFH 10 source stock) are straying into the American River (J. Smith, EBMUD, pers. comm. as 11 cited in NMFS 2009).

12 There has also been significant transfer of genetic material among hatcheries within the 13 Central Valley as well as some transfer from systems outside the Central Valley. There 14 have also been transfers of steelhead from the FRFH to the Mokelumne Hatchery. For 15 example, eyed eggs from the Nimbus Hatchery were transferred to the FRFH several 16 times in the late 1960s and early 1970s (DWR 2002). Also, Nimbus Hatchery steelhead 17 eggs have often been transferred to the Mokelumne Hatchery. Additionally, an Eel River 18 strain of steelhead was used as the founding broodstock for the Nimbus Hatchery (CDFG 19 1991a). In the late 1970s, a strain of steelhead was brought in from Washington State for 20 the FRFH (DWR 2002).

21 Long-Term Climate Change

Because steelhead normally spend a longer time in freshwater as juveniles than other
anadromous salmonids, any negative effects of climate change may be more profound on
steelhead populations.

25 HATCHERY OPERATIONS AND PRACTICES

In addition to the immediately previous discussion taken from Appendix B (Threats Assessment) of the NMFS Draft Recovery Plan (NMFS 2009), an additional discussion regarding the impacts of hatcheries on the Central Valley steelhead DPS is provided below.

1 Hatcheries have come under scrutiny for their potential effects on wild salmonid 2 populations (Bisson et al. 2002; Araki et al. 2007). The concern with hatchery operations 3 is two-fold. First, they may result in unintentional, but maladaptive genetic changes in 4 wild steelhead stocks (McEwan and Jackson 1996). CDFW believes its hatcheries take 5 eggs and sperm from enough individuals to avoid loss of genetic diversity through 6 inbreeding depression and genetic drift. However, artificial selection for traits that 7 improve hatchery success (e.g., fast growth, tolerance of crowding) are not avoidable and 8 may reduce genetic diversity and population fitness (Araki et al. 2007). Past and present 9 hatchery practices represent the major threat to the genetic integrity of Central Valley 10 steelhead (NMFS 2009). Overlap of spawning hatchery and natural fish within the 11 steelhead DPS exists, resulting in genetic introgression. Also, a substantial problem with 12 straying of hatchery fish exists within this DPS (Hallock 1989). Habitat fragmentation 13 and population declines resulting in small, isolated populations also pose genetic risk 14 from inbreeding, loss of rare alleles, and genetic drift (NMFS 2009).

15 The second concern with hatchery operations revolves around the potential for 16 undesirable competitive interactions between hatchery and wild stocks. Intraspecific 17 competition between wild and artificially produced stocks can result in wild fish declines 18 (McMichael et al. 1997; 1999). Although wild fish are presumably more adept at 19 foraging for natural foods than hatchery-reared fish, this advantage can be negated by 20 density-dependent effects resulting from large numbers of hatchery fish released at a 21 specific locale, as well as the larger size and more aggressive behavior of the hatchery 22 fish (Reclamation 2008).

23 Currently, four hatcheries in the Central Valley produce steelhead to supplement the 24 Central Valley wild steelhead population. These four Central Valley steelhead hatcheries 25 (Mokelumne River, FRFH, Coleman, and Nimbus hatcheries) collectively produce 26 approximately 1.5 million steelhead yearlings annually when all four hatcheries reach 27 production goals (CMARP 1998). The hatchery steelhead programs originated as 28 mitigation for the habitat lost by construction of dams. Steelhead are released at 29 downstream locations in January and February at about four fish per pound, generally 30 corresponding to the initiation of the peak of outmigration (Reclamation 2008). In the 31 Central Valley, practices such as transferring eggs between hatcheries and trucking smolts to distant sites for release contribute to elevated straying levels (USDOI 1999, as
 cited in NMFS 2009a).

3 According to Reclamation (2008), the hatchery runs in the American and Mokelumne 4 rivers are probably highly introgressed mixtures of many exotic stocks introduced in the 5 early days of the hatcheries (McEwan and Jackson 1996; NMFS 1998b). Beginning in 6 1962, steelhead eggs were imported into Nimbus Hatchery from the Eel, Mad, upper 7 Sacramento, and Russian rivers and from the Washougal and Siletz Rivers in Washington 8 and Oregon, respectively (McEwan and Nelson 1991, as cited in McEwan and Jackson 9 1996). Egg importation has also occurred at other Central Valley hatcheries (McEwan 10 and Jackson 1996).

11 Reclamation (2008) further states that stock introductions began at the FRFH in 1967, 12 when steelhead eggs were imported from Nimbus Hatchery to be raised as broodstock. 13 In 1971, the first release of Nimbus origin fish occurred. From 1975 to 1982, steelhead 14 eggs or juveniles were imported from the American, Mad, and Klamath rivers and the 15 Washougal River in Washington. The last year that Nimbus-origin fish were released into 16 the Feather River was 1988. Based on preliminary genetic assessments of Central Valley 17 steelhead, NMFS (1998b) concluded the FRFH steelhead were part of the Central Valley 18 DPS despite an egg importation history similar to the Nimbus Hatchery stock, which 19 NMFS did not consider part of the Central Valley DPS.

20 The increase in Central Valley hatchery production has reversed the composition of the 21 steelhead population, from 88% naturally-produced fish in the 1950s (McEwan 2001) to 22 an estimated 23 to 37% naturally-produced fish (Nobriga and Cadrett 2003). The 23 increase in hatchery steelhead production proportionate to the wild population has 24 reduced the viability of the wild steelhead populations, increased the use of out-of-basin 25 stocks for hatchery production, and increased straying (NMFS and CDFG 2001). Thus, 26 the ability of natural populations to successfully reproduce and continue their genetic 27 integrity likely has been diminished (Reclamation 2008).

In addition, harvest impacts associated with hatchery-wild population interactions have been identified as a stressor to wild Central Valley steelhead stocks (NMFS 2009). The relatively low number of spawners needed to sustain a hatchery population can result in

1 high harvest-to-escapements ratios in waters where fishing regulations are set according 2 to hatchery population. This can lead to over-exploitation and reduction in the size of 3 wild populations existing in the same system as hatchery populations due to incidental 4 bycatch (McEwan 2001). According to CDFW creel census surveys, the majority (93%) 5 of steelhead catches occur on the American and Feather rivers, sites of steelhead 6 hatcheries (CDFG 2001d, as cited in NMFS 2009). Creel census surveys conducted 7 during 2000 indicated that 1,800 steelhead were retained, and 14,300 were caught and 8 released. The total number of steelhead contacted might be a significant fraction of 9 basin-wide escapement, so even low catch-and-release mortality may pose a problem for 10 wild populations. Additionally, NMFS (2005b) asserted that steelhead fisheries on some 11 tributaries and the mainstem Sacramento River may affect some steelhead juveniles.

12 **4.3.5.2** Lower Yuba River

13 The lower Yuba River steelhead population is exposed and subject to the myriad of 14 limiting factors, threats and stressors described above for the DPS. Concurrently with the 15 effort conducted for spring-run Chinook salmon, NMFS (2009) recently conducted a 16 comprehensive assessment of stressors affecting both steelhead within the lower Yuba 17 River, and lower Yuba River steelhead populations as they migrate downstream (as 18 juveniles) and upstream (as adults) through the lower Feather River, the lower 19 Sacramento River, and the Bay-Delta system. For the lower Yuba River population of 20 steelhead, the number of stressors according to the categories of "Very High", "High", 21 "Medium", and "Low" that occur in the lower Yuba River or occur out of basin are presented below by lifestage (Table 4-9). 22

23 As shown by the numbers in Table 4-9, of the total number of 94 stressors affecting all 24 identified lifestages of lower Yuba River populations or steelhead, 31 are within the 25 lower Yuba River and 63 are out-of-basin. Because spawning and incubation occurs only 26 in the lower Yuba River, all of the stressors associated with these lifestages occur in the 27 lower Yuba River. For the adult immigration and holding, and the juvenile rearing and outmigration lifestages combined, a total of 49 "Very High" and "High" stressors were 28 29 identified, with 15 of those occurring in the lower Yuba River and 34 occurring 30 out-of-basin.

1 Table 4-9. The number of stressors according to the categories of "Very High", "High", 2 "Medium", and "Low" that occur in the lower Yuba River, or occur out-of-basin, by 3 lifestage for the lower Yuba River population of steelhead (Source: NMFS 2009).

| | | Stressor Categories | | | | | | | | |
|-------------------------------|------------------|---------------------|------|--------|-----|--|--|--|--|--|
| Lifestage | Location | Very High | High | Medium | Low | | | | | |
| Adult Immigration and Holding | | | | | | | | | | |
| | Lower Yuba River | 2 | 1 | 3 | 1 | | | | | |
| | Out of Basin | 1 | 5 | 10 | 4 | | | | | |
| Spawning | | | | | | | | | | |
| | Lower Yuba River | 3 | 2 | 0 | 2 | | | | | |
| | Out of Basin | N/A* | N/A | N/A | N/A | | | | | |
| Embryo Incubation | | | | | | | | | | |
| | Lower Yuba River | 1 | 0 | 4 | 0 | | | | | |
| | Out of Basin | N/A | N/A | N/A | N/A | | | | | |
| Juvenile Rearing and Outmigr | | | | | | | | | | |
| | Lower Yuba River | 5 | 1 | 1 | 5 | | | | | |
| | Out of Basin | 12 | 16 | 6 | 9 | | | | | |
| * N/A – Not Applicable. | | | | | | | | | | |

4 The NMFS (2009) Draft Recovery Plan states that "The lower Yuba River, below 5 Englebright Dam, is characterized as having a high potential to support a viable 6 population of steelhead, primarily because: (1) the river supports a persistent population 7 of steelhead and historically supported the largest, naturally reproducing population of 8 steelhead in the Central Valley (McEwan and Jackson 1996); (2) flow and water 9 temperature conditions are generally suitable to support all life stage requirements; (3) 10 the river does not have a hatchery on it; (4) spawning habitat availability does not 11 appear to be limited; and (5) high habitat restoration potential".

12 Similar to the statement for spring-run Chinook salmon, the NMFS (2009) Draft 13 Recovery Plan further states that "For currently occupied habitats below Englebright 14 Dam, it is unlikely that habitats can be restored to pre-dam conditions, but many of the 15 processes and conditions that are necessary to support a population of steelhead can be 16 improved with improvements to instream flow regimes, water temperatures, and habitat 17 availability. Continued implementation of the Yuba Accord is expected to address these 18 factors and considerably improve conditions in the lower Yuba River." 1 Many of the most important stressors specific to steelhead in the lower Yuba River 2 correspond to the stressors described for spring-run Chinook salmon in the lower Yuba 3 River, which included passage impediments and barriers, harvest and angling impacts, 4 poaching, physical habitat alteration, loss of riparian habitat and instream cover (e.g., 5 riparian vegetation, instream woody material), loss of natural river morphology and 6 function, loss of floodplain habitat, entrainment, predation, and hatchery effects.

7 The previous discussions in this BA addressing limiting factors and threats for the spring-8 run Chinook salmon population in the lower Yuba River that are pertinent to the 9 steelhead population in the lower Yuba River are not repeated in this section of the BA. 10 Stressors that are unique to steelhead in the lower Yuba River, and stressors that 11 substantially differ in severity for steelhead, are described below.

12 HARVEST/ANGLING IMPACTS

13 Fishing for steelhead on the lower Yuba River is regulated by CDFW. Angling 14 regulations on the lower Yuba River are intended to protect sensitive species, including 15 wild steelhead. CDFW angling regulations (2013/2014) permit fishing for steelhead from 16 the mouth of the Yuba River to the Highway 20 Bridge with only artificial lures with 17 barbless hooks all year-round. The regulations include a daily bag limit of two hatchery 18 trout or hatchery steelhead (identified by an adipose fin clip), and a possession limit of 19 four hatchery trout or hatchery steelhead. From the Highway 20 Bridge to Englebright 20 Dam, fishing for steelhead is permitted from December 1 through August 31 only, with 21 only artificial lures with barbless hooks. For this time period, the regulations include a 22 daily bag limit of two hatchery trout or hatchery steelhead (identified by an adipose fin 23 clip), and a possession limit of four hatchery trout or hatchery steelhead.

24 **POACHING**

By contrast to the previous discussion regarding the potential for poaching to be a stressor to spring-run Chinook salmon, no references have been reported regarding the potential poaching of steelhead at the fish ladders, or at the base of Daguerre Point Dam. In addition, no reference has been located regarding the occurrence of steelhead jumping out of the fish ladders at Daguerre Point Dam.

1 HATCHERY EFFECTS

The previous discussion in this BA addressing limiting factors, threats and stressors resulting from straying and other hatchery effects on the steelhead DPS that are pertinent to steelhead in the lower Yuba River are not repeated in this section of the BA. Hatchery-related stressors that are unique to steelhead in the lower Yuba River, or substantially differ in severity for Yuba River steelhead, are described below.

7 Although it has been oft-repeated that hatcheries historically have not been located on the 8 Yuba River, that does not appear to be the case. According to a document titled "A 9 History of California's Fish Hatcheries 1870–1960" (Leitritz 1970), an experimental fish 10 hatchery station (i.e., the Yuba River Hatchery) was established in 1928 by the California 11 Department of Natural Resources, Division of Fish and Game. The site was on Fiddle 12 Creek, a tributary of the North Fork Yuba River about 34 miles north of Nevada City, 13 Fish rearing began at the station in 1929. Over the years, near Camptonville. 14 improvements were made to the hatchery. No reference could be found regarding 15 salmon, but the hatchery was reported to hatch and rear trout, including steelhead (CDNR 16 1931). The hatchery continued operations until storms during November 1950 caused 17 such extensive damage that repairs could not be made and it was permanently closed 18 (Leitritz 1970).

Since that time, no fish hatcheries have been located on the lower Yuba River, and the river continues to support a persistent population of steelhead. According to the NMFS Draft Recovery Plan (NMFS 2009), the major threat to the genetic integrity of Central Valley steelhead results from past and present hatchery practices. These practices include the planting of non-natal fish, overlap of spawning hatchery and natural fish, and straying of hatchery fish.

25 GENETIC CONSIDERATIONS

From 1970 to 1979, CDFW annually stocked 27,270–217,378 fingerlings, yearlings, and
sub-catchable steelhead from Coleman National Fish Hatchery into the lower Yuba River
(CDFG 1991a). CDFW stopped stocking steelhead into the lower Yuba River in 1979.
In addition, it is possible that some hatchery-reared juvenile steelhead from the FRFH

may move into the lower Yuba River in search of rearing habitat. Some competition for
 resources with naturally spawned steelhead could occur as a result.

3 Previous genetic work on population structure of steelhead in California has relied 4 primarily on analyses of mitochondrial DNA (e.g. Berg and Gall 1988; Nielsen et al. 5 1997), which is a single gene that is often not reflective of population history or true 6 relationships (Chan and Levin 2005). However, microsatellites, also known as simple 7 sequence repeat loci, have been used in numerous studies of salmonids and have proven 8 to be a valuable tool for elucidating population genetic structure. Work on O. mykiss in 9 California using microsatellite loci has demonstrated that genetic structure can be 10 identified with such data, both at larger scales (Aguilar and Garza 2006) and at relatively 11 fine ones (Deiner et al. 2007; Pearse et al. 2007). The following discussion was taken 12 from Garza and Pearse (2008).

13 Garza and Pearse (2008) studied populations of O. mykiss in the Central Valley using 14 molecular genetic techniques to provide insight into population structure in the region. 15 Data were collected from 18 nuclear microsatellite loci and variation analyzed to trace 16 ancestry and evaluate genetic distinction among populations. The goals of the study were 17 to use population genetic analyses of the data to assess origins and ancestry of O. mykiss 18 populations above and below dams in Central Valley tributary rivers, to better understand 19 the relationship of these populations to others in California, and to provide information 20 on genetic diversity and population structure of these populations. Genotypes were 21 collected from over 1,600 individual fish from 17 population samples and five hatchery 22 rainbow trout strains. Fish populations from rivers and creeks that flow to both the 23 Sacramento and San Joaquin Rivers were evaluated, including the McCloud River, Battle 24 Creek, Deer Creek, Butte Creek, Feather River, Yuba River, American River, Calaveras 25 River, Stanislaus River and Tuolumne River sub-basins. Analyses included fish collected 26 both above and below barriers to anadromy in some of the study basins (Garza and 27 Pearse 2008).

28 Phylogeographic trees were used to visually and quantitatively evaluate genetic 29 relationships of Central Valley *O. mykiss* populations both with each other and with other 30 California populations. Genetic diversity was relatively similar throughout the Central

1 Valley. Above-barrier populations clustered with one another and below-barrier 2 populations are most closely related to populations in far northern California, specifically 3 the genetic groups that include the Eel and Klamath Rivers. Since Eel River origin 4 broodstock were used for many years at Nimbus Hatchery on the American River, it is 5 likely that Eel River genes persist there and have also spread to other basins by migration, 6 and that this is responsible for the clustering of the below-barrier populations with 7 northern California ones. This suggests that the below-barrier populations in this region 8 appear to have been widely introgressed with hatchery fish from out-of-basin broodstock 9 sources. In phylogeographic analyses, above-barrier populations are more similar to San 10 Francisco Bay O. mykiss populations than the below-barrier populations in the Central 11 Valley. Because this relationship is expected for steelhead, given their extraordinary 12 historic dependence on short distance migration events (Pearse and Garza 2007), they 13 may represent relatively non-introgressed historic population genetic structure for the 14 region. Other possible explanations for this pattern that rely on complicated, widespread 15 patterns of introgression with hatchery fish are not entirely ruled out, but are highly 16 improbable given that the above-barrier populations also group with moderate 17 consistency into geographically-consistent clusters (e.g. Yuba-Upper and Feather-Upper) 18 in all analyses and also because of the low apparent reproductive success of hatchery 19 trout in streams throughout California (Garza and Pearse 2008).

The analyses also identified possible heterogeneity between samples from different tributaries of the upper Yuba and Feather Rivers, although linkage disequilibrium was lower in these populations. Linkage disequilibrium can be caused by physical linkage of loci, sampling of related individuals/family structure, and by the sampling of more than one genetically distinct group within a population sample (Garza and Pearse 2008).

In general, although structure was found, all naturally-spawned *O. mykiss* populations within the Central Valley Basin were closely related, regardless of whether they were sampled above or below a known barrier to anadromy (Garza and Pearse 2008). This is due to some combination of pre-impoundment historic shared ancestry, downstream migration and, possibly, limited anthropogenic upstream migration. However, lower genetic diversity in above-barrier populations indicates a lack of substantial genetic input upstream and highlights lower effective population sizes for above-barrier populations. 1 The consistent clustering of the above-barrier populations with one another, and their 2 position in the California-wide trees, indicate that they are likely to most accurately 3 represent the ancestral population genetic structure of steelhead in the Central Valley 4 (Garza and Pearse 2008).

5 STRAYING INTO THE LOWER YUBA RIVER

6 The observation of adipose fin clips on adult steelhead passing upstream through the 7 VAKI Riverwatcher system at Daguerre Point Dam demonstrates that hatchery straying 8 into the lower Yuba River has, and continues, to occur. Although no information is 9 presently available regarding the origin of adipose-clipped steelhead observed at the 10 VAKI Riverwatcher system at Daguerre Point Dam, it is reasonable to surmise that they 11 most likely originate from the FRFH. The remainder of this discussion pertains to 12 hatchery effects associated with the straying of adult steelhead into the lower Yuba River.

13 If hatchery-origin steelhead stray into the lower Yuba River and interbreed with 14 naturally-spawning Yuba River steelhead, then such interbreeding has been suggested to 15 represent a threat to the genetic diversity and integrity of the naturally-spawning 16 steelhead population in the lower Yuba River. No previously conducted quantitative 17 analyses or data addressing the extent of hatchery-origin steelhead straying into the lower 18 Yuba River is available for presentation in this BA. However, some information is 19 presently available to assess the amount of straying of hatchery-origin (adipose fin-20 clipped) steelhead into the lower Yuba River from VAKI Riverwatcher data.

21 In the lower Yuba River, attempts were made to differentiate adult steelhead from other 22 O. mykiss (i.e., juvenile steelhead and resident rainbow trout) recorded passing Daguerre 23 Point Dam utilizing daily VAKI Riverwatcher data. However, only two years of data 24 (2010/2011 and 2011/2012) are available identifying adipose fin-clipped O. mykiss 25 passing through the VAKI Riverwatcher system, during which extensive inoperable 26 periods did not occur during the adult steelhead upstream migration period. Data 27 reduction, limitations and applications are described in Section 4.2.6 (Viability) of this 28 BA, below.

Analysis of the VAKI Riverwatcher data indicates that the percent contribution of hatchery-origin adult upstream migrating fish (represented by the percentage of adipose fin-clipped adult steelhead relative to the total number of adult upstream migrating
steelhead, because 100% of FRFH-origin steelhead have been marked since 1996) was
approximately 43% for the 2010/2011 biological year, and about 63% for the 2011/2012
biological year (RMT 2013).

5 4.3.6 Viability of the Central Valley Steelhead DPS

6 The VSP concept (McElhany et al. 2000) previously described in Section 4.1.6 of this
7 BA for the spring-run Chinook salmon ESU also is used to address and describe the
8 viability of the Central Valley Steelhead DPS.

9 4.3.6.1 DPS

10 As described by NMFS (2009), there are few data with which to assess the status of 11 Central Valley steelhead populations. Lindley et al. (2007) stated that, with the few 12 exceptions of streams with long-running hatchery programs such as Battle Creek and the 13 Feather, American and Mokelumne rivers, Central Valley steelhead populations are 14 classified as data deficient. In all cases, hatchery-origin fish likely comprise the majority 15 of the natural spawning run, placing the natural populations at high risk of extinction 16 (Lindley et al. 2007). As of 2009, NMFS (2009) reinforced the conclusion that the 17 Central Valley steelhead DPS is data deficient, with the exception of these 18 hatchery programs.

From 1967-1993, steelhead run-size estimates were generated from fish counts in the fish ladder at RBDD (CDFG 2010a). From these counts, estimates of the natural spawner escapement upstream of RBDD were generated. Because RBDD impacted winter-run Chinook salmon by delaying their upstream migration, dam operations were changed in 1993 so that dam gates were raised earlier in the season, which eliminated the need for fish to navigate fish ladders, but also eliminated the ability to generate accurate run-size estimates for the upper Sacramento River Basin (CDFG 2010a).

Presently, little information is available regarding the abundance of steelhead in the
Central Valley (CDFG 2010a). Currently there is virtually no coordinated,
comprehensive, or consistent monitoring of steelhead in the Central Valley. In 2004, the

1 Interagency Ecological Program Steelhead Project Work Team developed a proposal to 2 develop a comprehensive monitoring plan for Central Valley steelhead. In 2007, 3 development of this steelhead monitoring plan was funded by the CALFED Ecosystem 4 Restoration Program. In 2010, a document titled "A Comprehensive Monitoring Plan for 5 Steelhead in the California Central Valley" was completed by CDFG (2010a), which recommended steelhead monitoring activities in the Central Valley. The objectives of the 6 7 plan include: (1) estimate steelhead population abundance with levels of precision; (2) 8 examine trends in steelhead abundance; and (3) identify the spatial distribution of 9 steelhead in the Central Valley to assess their current range and observe changes in their 10 range that may occur over time. However, for the most part, recommendations in the 11 plan remain to be implemented.

12 According to NMFS (2009), data are lacking to suggest that the Central Valley steelhead 13 DPS is at low risk of extinction, or that there are viable populations of steelhead 14 anywhere in the DPS. Conversely, there is evidence to suggest that the Central Valley 15 steelhead DPS is at moderate or high risk of extinction (McEwan 2001; Good et al. 16 2005). Most of the historical habitat once available to steelhead has been lost (Yoshiyama 17 et al. 1996; McEwan 2001; Lindley et al. 2006). Furthermore, the observation that 18 anadromous O. mykiss are becoming rare in areas where they were probably once 19 abundant indicates that an important component of life history diversity is being 20 suppressed or lost (NMFS 2009). Habitat fragmentation, degradation, and loss are likely 21 having a strong negative impact on many resident as well as anadromous O. mykiss 22 populations (Hopelain 2003 as cited in NMFS 2009).

23 VIABLE SALMONID POPULATION (VSP) PARAMETERS AND APPLICATION

24 ABUNDANCE AND PRODUCTIVITY

According to NMFS (2009a) and CDFG (2010a), there is still a paucity of steelhead monitoring in the Central Valley. Therefore, data are lacking regarding abundance estimates for the steelhead DPS, or for specific steelhead populations in the Central Valley (NMFS 2009a). Recognizing these data limitations, NMFS (2009a) suggested that natural steelhead escapement in the upper Sacramento River declined substantially from 1967 through 1993, and that the little data that do exist indicate that the steelhead population continues to decline. Also, according to Lindley et al. (2007), even if there were adequate data on the distribution and abundance of steelhead in the Central Valley, their approaches for assessing steelhead population and DPS viability might be problematical because the effect of resident *O. mykiss* on the viability of steelhead populations and the DPS is unknown.

6 SPATIAL STRUCTURE

For the Central Valley steelhead DPS, Lindley et al. (2006) identified historical independent populations based on a model that identifies discrete habitat and interconnected habitat patches isolated from one another by downstream regions of thermally unsuitable habitat. They hypothesized that historically 81 independent populations of steelhead were dispersed throughout the Central Valley domain.

About 80% of the habitat that was historically available to steelhead is now behind impassable dams, and 38% of the populations have lost all of their habitats (NMFS 2009a). Although much of the habitat has been blocked, or degraded, by impassable dams, small populations of steelhead are still found throughout habitat available in the Sacramento River and many of the tributaries, and some of the tributaries to the San Joaquin River. The current distribution of steelhead is less well understood, but the DPS is composed of at least four diversity groups and at least 26 populations (NMFS 2009).

19 Remnant steelhead populations are presently distributed through the mainstem of the 20 Sacramento and San Joaquin rivers, as well as many of the major tributaries of these 21 rivers (NMFS 2009). Steelhead presence in highly variable "flashy" streams and creeks 22 in the Central Valley depend primarily on flow and water temperature, which can change 23 drastically from year to year (McEwan and Jackson 1996). As stated in NMFS (2009), 24 spawner surveys of small Sacramento River tributaries (Mill, Deer, Antelope, Clear, and 25 Beegum creeks) and incidental captures of juvenile steelhead during Chinook salmon 26 monitoring (Calaveras, Cosumnes, Stanislaus, Tuolumne, and Merced rivers) confirmed 27 that steelhead are widespread, if not abundant, throughout accessible streams and rivers 28 (Good et al. 2005).

1 Diversity

Steelhead naturally experience the most diverse life history strategies of the listed Central
Valley anadromous salmonid species (NMFS 2009a). However, steelhead has less
flexibility to track changes in the environment as the species' abundance decreases and
spatial structure of the DPS is reduced (NMFS 2009a).

6 The posited historical existence of 81 independent steelhead populations is likely to be an 7 underestimate because large watersheds that span a variety of hydrological and 8 environmental conditions, such as the Pit River, probably contained multiple populations 9 (Lindley et al. 2006). Regardless, the distribution of many discrete populations across a 10 wide variety of environmental conditions implies that the Central Valley steelhead DPS 11 contained biologically significant amounts of spatially structured genetic diversity 12 (Lindley et al. 2006). However, it appears that much of the historical diversity within 13 Central Valley O. mykiss has been lost or is threatened by dams, which have heavily 14 altered the distribution and population structure of steelhead in the Central Valley 15 (Lindley et al. 2006).

16 Although historically two different runs of steelhead (summer-run and winter-run) 17 occurred in the Central Valley (McEwan and Jackson 1996), the summer run has been 18 largely extirpated due to a lack of suitable holding and staging habitat, such as coldwater 19 pools in the headwaters of Central Valley streams, presently located above impassible 20 dams (Lindley et al. 2006).

Throughout the Central Valley (and in particular the Merced River, Tuolumne River, and upper Sacramento River) it is difficult to discriminate between adult anadromous and resident forms of *O. mykiss*, as well as their progeny (McEwan 2001), further complicating resource management agencies' understanding of steelhead distribution in the Central Valley (CDFG 2008).

The genetic diversity of steelhead also is compromised by hatchery-origin fish. According to Reclamation (2008), estimates of straying rates only exist for Chinook salmon produced at the FRFH. However, general principles and the potential effects of straying are also applicable for steelhead. Based on available genetic data, the effects of hatcheries that rear steelhead appear to be restricted to the populations on hatchery streams (DWR 2004c). These findings suggest that, although ongoing operations may
impact the genetic composition of the naturally spawning steelhead population in these
rivers, hatchery effects appear to be localized, although it should be noted that genetic
data for steelhead are limited (DWR 2004c).

5 SUMMARY OF THE VIABILITY OF THE CENTRAL VALLEY STEELHEAD DPS

6 Although data are lacking to quantitatively evaluate extinction risk for the Central Valley 7 steelhead DPS, NMFS (2009) states that there is evidence to suggest that the Central 8 Valley steelhead DPS is at moderate or high risk of extinction. Steelhead have been 9 extirpated from most of their historical range throughout the Central Valley domain, and 10 most of the historical habitat once available to steelhead is largely inaccessible. 11 Anadromous forms of *O. mykiss* are becoming less abundant or rare in areas where they 12 were probably once abundant, and habitat fragmentation, degradation, and loss are likely 13 having a strong negative impact on many resident as well as anadromous O. mykiss 14 populations. In addition, widespread hatchery steelhead production within this DPS also 15 raises concerns about the potential ecological interactions between introduced stocks and 16 native stocks (Corps 2007).

17 As previously discussed, NMFS completed a 5-year status review of the Central Valley 18 steelhead DPS during August 2011. Good et al. (2005) previously found that Central 19 Valley steelhead were in danger of extinction, with a minority of the NMFS BRT 20 viewing the DPS as likely to become endangered. The NMFS BRT's primary concerns 21 for the DPS included the low abundance of naturally-produced anadromous fish at the 22 DPS level, the lack of population-level abundance data, and the lack of information to 23 suggest that the monotonic decline in steelhead abundance evident from 1967-1993 dam 24 counts has stopped (NMFS 2011c).

Steelhead population trend data remain extremely limited (Williams et al. 2011). The Chipps Island midwater trawl dataset of USFWS provides information on the trend in abundance for the Central Valley steelhead DPS as a whole. Updated through 2010, the trawl data indicate that the decline in natural production of steelhead has continued unabated since the 2005 status review (NMFS 2011c). Catch-per-unit-effort has fluctuated but remained level over the past decade, but the proportion of the catch that is ad-clipped (100% of hatchery steelhead production have been ad-clipped starting in
 1998) has risen steadily, exceeding 90% in recent years and reaching 95% in 2010
 (NMFS 2011c). Because hatchery releases have been fairly constant, this implies that
 natural production of juvenile steelhead has been declining (NMFS 2011c).

5 According to NMFS (2011c), steelhead returns to the FRFH have decreased substantially 6 in the last several years with only 679, 312 and 86 fish returning in 2008, 2009 and 2010, 7 respectively. Because almost all of the returning fish are of hatchery origin and stocking 8 levels have remained fairly constant over the years, data suggest that adverse freshwater 9 and/or ocean survival conditions have caused or at least contribute to these declining 10 hatchery returns (NMFS 2011c). The Central Valley experienced three consecutive years 11 of drought (2007-2009), which NMFS (2011c) states would likely have impacted parr 12 and smolt growth and survival. Additionally, poor ocean conditions have occurred in at 13 least 2005 and 2006, which have affected Chinook populations in the Central Valley and 14 also may have affected steelhead populations (NMFS 2011c). Preliminary return data 15 for 2011 from CDFW suggest a strong rebound in return numbers during 2011, with 712 adults returning to the FRFH through April 5th (NMFS 2011c). Based on steelhead 16 17 returns to Central Valley hatcheries and the redd counts on Clear Creek, the American 18 River, and the Mokelumne River, it appears that naturally-produced steelhead may not 19 have been impacted by poor freshwater and marine rearing conditions as much as 20 hatchery-origin fish during the last several years (NMFS 2011c). However, NMFS 21 (2011c) suggests that this observation may reflect greater fitness of naturally-produced 22 steelhead relative to hatchery fish, and merits further study.

23 The steelhead DPS includes two hatchery populations — the FRFH and Coleman 24 National Fish Hatchery. Two additional hatchery populations (i.e., Nimbus and 25 Mokelumne River hatcheries) also are present in the Central Valley, but they were 26 founded from out-of-DPS broodstock and are not considered part of the DPS (NMFS 27 2011c). Recent genetic information suggests that below dam populations of O. mykiss 28 are similar genetically throughout the Central Valley and that genetic diversity and 29 population structure may have been lost over time. Garza and Pearse (2008) analyzed the 30 genetic relationships among Central Valley O. mykiss populations and found that all 31 below-barrier populations were generally closely related, and that there was a high level

of genetic similarity to Eel River and Klamath River steelhead in all below-barrier 1 2 population samples. These findings raises an issue about whether or not the steelhead 3 stocks propagated at the Nimbus and Mokelumne River hatcheries should be excluded 4 from the Central Valley steelhead DPS. These two stocks were excluded from the DPS 5 in 2006 because they originated from the Eel River which is not from within the DPS. 6 Because the Eel River strain appears to be widely introgressed in many Central Valley 7 steelhead populations, NMFS (2011c) states that it may be appropriate to re-evaluate whether or not these stocks should be in the DPS based upon the new 8 9 genetic information.

Using data through 2005, Lindley et al. (2007) found the data were insufficient to determine the status of any of the naturally-spawning populations of Central Valley steelhead, except for those spawning in rivers adjacent to hatcheries. These hatchery influenced populations were likely to be at high risk of extinction due to extensive spawning of hatchery-origin fish in natural areas (NMFS 2011c).

15 Overall, the status of the Central Valley steelhead DPS appears to have worsened since 16 the 2005 status review when the DPS was considered to be in danger of extinction (Good 17 et al. 2005). Analysis of catch data from the Chipps Island monitoring program suggests 18 that natural steelhead production has continued to decline and that hatchery origin fish 19 represent an increasing proportion of the juvenile production in the Central Valley. Data 20 from the Delta fish salvage facilities also suggests a general decline in the natural 21 production of steelhead (NMFS 2011c). Data on Coleman and FRFH hatchery 22 populations suggest they have declined in the last several years perhaps in response to 23 poor freshwater and ocean habitat conditions. Limited information suggest some 24 individual steelhead populations in the Central Valley are declining in abundance, but 25 more complete data for the Battle Creek population indicate the declines there have been 26 relatively moderate since 2005 and that the population in Clear Creek is increasing 27 (NMFS 2011c).

One continuing area of strength for the Central Valley steelhead DPS is its widespread spatial distribution throughout most watersheds in the Central Valley. All of the factors originally identified as being responsible for the decline of this DPS are still present,

1 although in some cases they have been reduced by regulatory actions (e.g., NMFS) 2 CVP/SWP OCAP Biological Opinion in 2009, actions required by CVPIA). Good et al. 3 (2005) described the threats to Central Valley salmon and steelhead as falling into three 4 broad categories, including: (1) loss of historical spawning habitat; (2) degradation of 5 remaining habitat; and (3) genetic threats from the stocking programs. Cummins et al. 6 (2008) attributed the much reduced biological status of anadromous salmonid stocks in 7 the Central Valley, including steelhead, to the construction and operation of the CVP and 8 SWP. Important conservation efforts have been implemented including the 2009 9 CVP/SWP biological opinion, CVPIA restoration efforts, and continued efforts to implement the Battle Creek Restoration Project that will eventually open up 42 miles of 10 11 high quality habitat to steelhead (NMFS 2011c). Although these efforts have provided 12 benefits to steelhead and its habitat in the Central Valley, threats from lost habitat and 13 degraded habitat continue to be important factors affecting the status of this DPS. Impacts to steelhead from harvest, research activities, disease and predation were considered 14 15 relatively minor factors in previous reviews, and there is little or no evidence indicating 16 impacts from these factors have changed (NMFS 2011c). In contrast, threats from other factors such as hatcheries, drought, poor ocean survival conditions, and climate change 17 18 have not been addressed and/or they have increased since the 2005 status review and 19 some are likely responsible for the recent declining abundance of the DPS 20 (NMFS 2011c).

21 In summary, the most recent biological information suggests that the extinction risk of 22 this DPS has increased since the last status review and that several of the listing factors 23 have contributed to the decline, including recent years of drought and poor ocean 24 conditions (NMFS 2011c). According to NMFS (2011c), there continue to be ongoing 25 threats to the genetic integrity of naturally-spawning steelhead from Central Valley 26 steelhead hatchery programs, but it is unclear if or how this factor has influenced the 27 overall viability of the DPS. The best available information on the biological status of 28 the DPS and continuing and new threats to the DPS indicate that its ESA status as a 29 threatened species is appropriate (NMFS 2011c).

1 4.3.6.2 Lower Yuba River

2 As with all naturally-spawning populations of steelhead in the Central Valley, Lindley et 3 al. (2007) characterized the steelhead population in the lower Yuba River as data 4 deficient, and therefore did not characterize its viability. Data limitations, particularly 5 regarding abundance and productivity, continue to render problematic quantitative 6 estimation procedures to assess the viability of the steelhead population in the lower 7 Yuba River. Continued monitoring of adult steelhead in the lower Yuba River is 8 providing additional information that is needed to assess extinction risk based on Lindley 9 et al. (2007) criteria regarding population size, recent population decline, occurrences of 10 catastrophes within the last 10 years that could cause sudden shifts from a low risk state 11 to a higher one, and the impacts of hatchery influence. The VSP parameters of 12 abundance, productivity, spatial structure and diversity for the steelhead population in the 13 lower Yuba River are discussed below.

14 ABUNDANCE AND PRODUCTIVITY

15 VAKI RIVERWATCHER DATA

16 Ongoing monitoring of the adult steelhead population in the lower Yuba River has been 17 conducted since 2003 with VAKI Riverwatcher systems at Daguerre Point Dam. By 18 contrast to Chinook salmon, escapement surveys involving carcass mark-recovery 19 experiments are not performed on steelhead/*O. mykiss*.

In the lower Yuba River, silhouettes and corresponding photographs were examined for species identification and categorization using methodology similar to that which is described for spring-run Chinook salmon. However, the accurate identification of *O*. *mykiss* in the VAKI Riverwatcher is more difficult than it is for Chinook salmon.

By contrast to the identification of Chinook salmon which may be conducted with a single attribute, the identification of steelhead becomes more problematic with the absence of a defining silhouette or a clear digital photograph. Additionally, the silhouettes of steelhead cannot reliably be differentiated from resident rainbow trout, and photo documentation of an individual is problematic because adult steelhead typically immigrate during periods of high flow and associated high turbidity and low visibility. The VAKI Riverwatcher systems cannot differentiate an individual as a resident form of
 the species (i.e., rainbow trout) or as anadromous (i.e., steelhead). Additionally, the
 VAKI Riverwatcher systems cannot directly distinguish between an adult or juvenile *O*.
 mykiss (RMT 2013).

5 DIFFERENTIATION OF ADULT STEELHEAD VAKI RIVERWATCHER COUNTS

6 The silhouettes and/or electronic images of each fish passage event that was identified as 7 an *O. mykiss* fish passage event allow the VAKI Riverwatcher systems to calculate an 8 approximate length (in centimeters) for the observed fish.

9 As reported by the RMT (2013), as an initial step in the differentiation of adult steelhead 10 passing upstream of Daguerre Point Dam, the length distribution of all fish identified as 11 O. mykiss passing through both the north and south ladders at Daguerre Point Dam over 12 the entire data availability period (January 1, 2004 through February 29, 2012) was 13 plotted and visually examined (Figure 4-10). This figure indicates the possible presence 14 of at least six length groups. These groups represent the potential combination of 15 juvenile and adult anadromous O. mykiss (steelhead), as well as juvenile and adult 16 resident O. mykiss (rainbow trout). However, this length-frequency distribution does not provide information necessary to differentiate between steelhead and rainbow trout. 17

Beginning March 1, 2009, VAKI Riverwatcher fish identified as *O. mykiss* also were classified as fish with or without clipped adipose fins, based on the inspection of the fish silhouette and photogrammetric representation (digital photographs and/or video imagery). The analysis of the length-frequency distribution of all adipose fin-clipped *O. mykiss* provides a means of differentiating adult steelhead passing upstream of Daguerre Point Dam from all other *O. mykiss*, because all adipose fin-clipped *O. mykiss* are steelhead that were released by a Central Valley hatchery.

The lengths of all fish passing upstream at Daguerre Point Dam that were identified as *O. mykiss* with clipped adipose fins (i.e., all hatchery steelhead) between March 1, 2009 through February 29, 2012 are presented in **Figure 4-11**. Visual examination of the observed length distribution in Figure 4-11 indicates the possible presence of up to five groups of fish. Two of the length categories demarcating the first two possible groups of fish occur at 20 cm (7.9 inches) and 29 cm (11.4 inches).



1

Figure 4-10. Length distribution of all fish identified by the VAKI Riverwatcher systems as
 O. mykiss passing upstream through the north and south ladders of Daguerre Point Dam
 from January 1, 2004 through February 29, 2012 (Source: RMT 2013).



5

6 Figure 4-11. Length distribution of all fish identified by the VAKI Riverwatcher systems as 7 adipose clipped *O. mykiss* passing upstream through the north and south ladders of 8 Daguerre Point Dam from March 1, 2009 through February 29, 2012 (Source: RMT 2013).

1 According to CDFG and USFWS (2010), the normal FRFH release schedule includes the 2 release of steelhead yearlings, from January to February, released in the Feather River 3 near Gridley at four fish per pound. Although not readily available from CDFW, other 4 sources indicate that steelhead smolts averaging 4 to 5 fish per pound range in length 5 from approximately 8-9 inches (20-23 cm) (IDFG 1992). The presence of small, adipose 6 fin-clipped steelhead in the lower Yuba River as displayed in Figure 4-11 may be related 7 to releases of yearling FRFH-produced steelhead on the Feather River. Since 2007, the FRFH has been releasing only steelhead yearlings at various sites along 8 9 the Feather River, as well as in the Sacramento River at Sutter Slough, and in Butte Creek 10 (Table 4-10). To determine whether fish planted in the lower Feather River may have

been detected in the lower Yuba River, an examination of the VAKI Riverwatcher data was conducted for adipose fin-clipped steelhead consistent with the observed potential

13 length-mode demarcation length of 29 cm (11.4 in) (RMT 2013).

Table 4-10. Recent releases of hatchery steelhead by the Feather River Fish Hatchery (Source: Regional Mark Information System (RMIS) of the Regional Mark Processing Center; RMT 2013).

| Release Dates | | Brood | Numbers | Released | Release | Study | | Agency | | | | | |
|---------------------|---|-------|----------------------------------|-----------------------|--------------------|-------------------|------------------------------------|-----------|---------|--|--|--|--|
| Start | End | Year | Tagged ¹ Adclipped | Untagged Adclipped | Stage ² | Type ³ | Release Location | Reporting | Release | | | | |
| 01/08/07 | 02/05/07 | 2006 | 0 | 10,036 | Y | E | Feather River Thermalito Bypass | CDFG | CDWR | | | | |
| 02/05/07 | 02/21/07 | 2006 | 0 | 488,043 | Y | Е | Feather River | CDFG | CDWR | | | | |
| 05/29/07 | 05/29/07 | 2006 | 0 | 1,643 | Y | E | Feather River | CDFG | CDWR | | | | |
| 05/30/08 | 05/30/08 | 2007 | 0 | 1,109 | Y | E | Feather River | CDFG | CDWR | | | | |
| 02/01/08 | 02/14/08 | 2007 | 0 | 307,986 | Y | Р | Feather River Boyds Pump Ramp | CDFG | CDWR | | | | |
| 02/03/09 | 02/03/09 | 2008 | 0 | 2,750 | Y | Р | Feather River at Live Oak | CDFG | CDFG | | | | |
| 02/03/09 | 02/17/09 | 2008 | 0 | 398,148 | Y | Р | Feather River Boyds Pump Ramp | CDFG | CDFG | | | | |
| 02/01/10 | 02/11/10 | 2009 | 0 | 272,798 | Y | Р | Feather River Boyds Pump Ramp | CDFG | CDFG | | | | |
| 02/02/11 | 02/15/11 | 2010 | 0 | 49,800 | Y | Ρ | Feather River Boyds Pump Ramp | CDFG | CDFG | | | | |
| ¹ Tagged | ¹ Tagged releases refer to releases with coded wire tags | | | | | | | | | | | | |

² Release stage Y indicates yearling releases.

³ Study type E stands for experimental releases, and study type P indicates a production releases.

1 From February 1, 2010 to February 2, 2011 (i.e., the starting date for the last reported 2 release of adipose fin-clipped juvenile steelhead from the FRFH), 104 adipose fin-clipped 3 juvenile steelhead with lengths less than or equal to 29 cm (11.4 in) were recorded 4 passing upstream of Daguerre Point Dam. Most of these individuals were observed in the 5 VAKI Riverwatcher system during February through April of 2010. Additionally, from 6 February 2, 2011 through January 31, 2012, a total of 1,702 adipose fin-clipped steelhead 7 with lengths less than or equal to 29 cm (11.4 in) were recorded passing upstream of 8 Daguerre Point Dam. While these individuals were observed in the VAKI Riverwatcher 9 system throughout calendar year 2011, they were most frequently observed during April 10 and May of 2011. In other words, most of the observed adipose fin-clipped juvenile 11 steelhead less than or equal to 29 cm (11.4 in) passing upstream of Daguerre Point Dam occurred within a few months after plantings of juvenile steelhead in the Feather River 12 13 from the FRFH. Additionally, between February 2011 and January 2012, approximately 14 676 adipose fin-clipped steelhead with lengths less than or equal to 29 cm were recorded 15 passing downstream of Daguerre Point Dam, with the majority of these individuals 16 passing downstream during April through June. Therefore, approximately one-third of the presumed FRFH steelhead that migrated upstream of Daguerre Point Dam during 17 18 2011 apparently turned around and migrated back downstream of Daguerre Point Dam 19 shortly after passing upstream of Daguerre Point Dam (RMT 2013).

20 If the observation of adipose fin-clipped juvenile steelhead passing upstream at Daguerre 21 Point Dam is associated with the release of yearling steelhead from the FRFH into the 22 lower Feather River, then it logically follows that the planted FRFH yearling steelhead 23 would have had to swim 6 miles upstream from the planting location at Boyds Pump 24 Ramp to the mouth of the lower Yuba River, and then an additional nearly 12 miles 25 upstream to reach Daguerre Point Dam. Although this phenomenon may seem somewhat 26 illogical, it has been reported elsewhere (Steiner Environmental Consulting 1987, as cited 27 in RMT 2013) and is an explanation for the observation of adipose fin-clipped juvenile 28 steelhead passing upstream at Daguerre Point Dam, because no marked juvenile steelhead 29 have been reported to be released over this time frame into the lower Yuba River.

30 The length-frequency distribution of all adipose fin-clipped steelhead observed at 31 Daguerre Point Dam from March 1, 2009 through February 29, 2012 was used to differentiate between "juvenile" and "adult" steelhead. The second step in the separation
of "juvenile" and "adult" steelhead was to fit modeled length-frequency distributions to
the observed data to determine a threshold length to separate both fish groups. A detailed
description of the analytical processes is provided in RMT (2013).

5 Unlike the methodology employed for Chinook salmon, the daily counts of adult 6 steelhead passing upstream of Daguerre Point Dam were not corrected for days when the 7 VAKI Riverwatcher systems were not fully operational. The RMT determined it would 8 be inappropriate to attempt to correct the adult steelhead counts due to: (1) the relatively 9 low numbers of adult steelhead recorded during most of the steelhead biological years; 10 and (2) the frequently extended durations when the VAKI Riverwatcher systems were not 11 fully operational during the steelhead immigration season. Instead, the daily counts of 12 adult steelhead passing upstream at Daguerre Point Dam were used to represent the 13 abundance of steelhead, with the understanding that the resultant estimates are minimum 14 numbers, and most of the survey years considerably underestimate the potential number 15 of steelhead because the annual estimates do not include periods of VAKI Riverwatcher 16 system non-operation, and do not consider the fact that not all steelhead migrate past 17 Daguerre Point Dam, due to some spawning occurring downstream Daguerre Point Dam.

18 ASSESSMENT OF AVAILABLE VAKI RIVERWATCHER DATA

For assessment purposes, a "steelhead biological year" was identified as extending from August 1 through July 31 each year, because: (1) preliminary review of the VAKI Riverwatcher data indicated a general paucity of upstream migrant *O. mykiss* during early summer; (2) the immigration of adult steelhead in the lower Yuba River has been reported to occur beginning during August (CALFED and YCWA 2005; McEwan and Jackson 1996); and (3) the RMT (2010b) identified the steelhead upstream migration period as beginning during August in the lower Yuba River (RMT 2013).

26 ANNUAL TIME SERIES OF STEELHEAD PASSING UPSTREAM OF DAGUERRE POINT DAM

27 **Figures 4-12 through 4-16** illustrate the daily counts of adult steelhead passing upstream

at Daguerre Point Dam through both the North and South ladders combined, and the

percentage of the daily number of hours when the VAKI Riverwatcher systems were
 operational at both ladders, during the eight steelhead biological years.

Examination of Figures 4-12 through 4-16 demonstrates that although the VAKI
Riverwatcher systems have been in place since June of 2003, reliable estimates of the
number of adult steelhead passing upstream at Daguerre Point Dam are essentially
restricted to the last two years of available data (2010/2011 and 2011/2012).

7 Due to system failures, including equipment malfunctions and operationally detrimental 8 environmental conditions (heavy overcast and foggy conditions resulting in lack of 9 photovoltaic charging of the system), the VAKI Riverwatcher systems were partially 10 operational or completely non-operational during several months each year of sampling. 11 Additionally, high flows and turbidities reduced the ability of the system to identify, or 12 prevented the system from identifying, adult steelhead oftentimes when the systems were 13 operational. Although improvements to the system have been made over time, it was not 14 until the most recent system improvements were implemented during the 2010/2011 15 sampling season that the system began demonstrating sustained reliability in the 16 documentation of steelhead passing upstream of Daguerre Point Dam, over a range of 17 environmental conditions.

18 Since June 2003, numerous improvements have been implemented to improve the 19 reliability of the VAKI Riverwatcher systems, and particularly their ability to document 20 passage during the steelhead upstream migration season. A chronology of the VAKI 21 Riverwatcher system improvements that have occurred over time are described in 22 RMT (2013).

This suite of improvements to the VAKI Riverwatcher systems at Daguerre Point Dam have resulted in much more reliable estimates of steelhead passing the dam. Correspondingly, the largest number of steelhead recorded immigrating past Daguerre Point Dam occurred during the 2010/2011 sampling season. As a result, it is not reasonable to consider data gathered prior to 2010/2011 to be reliable estimates of the annual number of adult steelhead passing upstream of Daguerre Point Dam (RMT 2013).




(dark green bars), and daily number of hours when the VAKI Riverwatcher systems were operational (light green bars), during the 2003/2004 and 2004/2005 steelhead biological

years (August 1 through July 31) (Source: RMT 2013).

7

2



3

Figure 4-13. Daily counts of adult steelhead passing upstream Daguerre Point Dam (dark green bars), and daily number of hours when the VAKI Riverwatcher systems were operational (light green bars), during the 2005/2006 and 2006/2007 steelhead biological years (August 1 through July 31) (Source: RMT 2013).

2



(dark green bars), and daily number of hours when the VAKI Riverwatcher systems were operational (light green bars), during the 2007/2008 and 2008/2009 steelhead biological

years (August 1 through July 31) (Source: RMT 2013).

2



3

Figure 4-15. Daily counts of adult steelhead passing upstream of Daguerre Point Dam (dark green bars), and daily number of hours when the VAKI Riverwatcher systems were operational (light green bars), during the 2009/2010 and 2010/2011 steelhead biological years (August 1 through July 31) (Source: RMT 2013).



1 2 3 4

Figure 4-16. Daily counts of adult steelhead passing upstream of Daguerre Point Dam (dark green bars), and daily number of hours when the VAKI Riverwatcher systems were operational (light green bars), during the 2011/2012 steelhead biological year (August 1 5 through July 31) (Source: RMT 2013).

6 As stated approximately six years ago by Lindley et al. (2006), there are almost no data 7 with which to assess the status of any of the Central Valley steelhead populations, with 8 the exceptions of the hatchery programs on Battle Creek and the Feather, American and 9 Mokelumne rivers. Therefore, they classified Central Valley steelhead populations as 10 data deficient. As of 2010, CDFG (2010a) stated that steelhead monitoring programs in 11 the Central Valley lack statistical power, are not standardized and in many cases lack 12 dedicated funding.

13 The relatively short time period encompassed by the reporting of reliable abundance 14 estimates, and in consideration that steelhead may have returned to the lower Yuba River 15 but remained and spawned in the river downstream of Daguerre Point Dam, currently 16 render problematic the determination of abundance or trends in the productivity of the steelhead over recent years (RMT 2013). Continued implementation of the improved 17 18 VAKI Riverwatcher systems at Daguerre Point Dam is likely to obtain some of the data 19 necessary to allow abundance estimation and productivity evaluation of steelhead in the 20 lower Yuba River. However, presently the lack of multi-year abundance data precludes the provision of quantitative values associated with extinction risk assessment, addressing
 abundance and productivity (RMT 2013).

3 SPATIAL STRUCTURE

4 Spatial structure and considerations regarding anadromous salmonid viability was 5 presented for spring-run Chinook salmon previously in this BA. The spatial structure 6 considerations, as one of the four VSP parameters, for steelhead are analogous to those 7 for spring-run Chinook salmon previously presented. Namely, spatial structure of 8 morphological units in the lower Yuba River is complex, diverse, and persistent.

9 **Diversity**

10 Phenotypic Considerations

11 O. mykiss in the lower Yuba River exhibit a high amount of diversity in phenotypic 12 expression and life history strategy. As demonstrated in Figures 4-12 through 4-16, O. 13 mykiss categorized as adult steelhead exhibit a broad temporal distribution in passing 14 upstream of Daguerre Point Dam. O. mykiss (including steelhead) exhibit highly diverse 15 spatial and temporal distributions in patterns of spawning, and juvenile outmigration 16 (RMT 2013). Moreover, O. mykiss in the lower Yuba River exhibit polyphenism, or the 17 occurrence of several phenotypes in a population which may not be due to different 18 genetic types, including expressions of anadromy or residency. A thorough discussion of 19 anadromy vs. residency of O. mykiss in the lower Yuba River is provided in RMT (2013). 20 A polymorphic O. mykiss population structure may be necessary for the long-term 21 persistence in highly variable environments such as the Central Valley (McEwan 2001). 22 Resident fish may reduce extinction risk through the production of anadromous 23 individuals that can enhance weak steelhead populations (Lindley et al. 2007). Such 24 considerations may be applicable to the O. mykiss populations in the lower Yuba River.

25 Genetic Considerations

Although no fish hatcheries have been located on the Yuba River since 1950, and the lower Yuba River continues to support a persistent population of steelhead, the genetic integrity of these fish is presently uncertain. According to the NMFS Draft Recovery Plan (NMFS 2009a), the major threat to the genetic integrity of Central Valley steelhead
 results from past and present hatchery practices. These practices include the planting of
 non-natal fish, overlap of spawning hatchery and natural fish, and straying of hatchery
 fish.

5 The observation of adipose fin clips on adult steelhead passing upstream through the 6 VAKI Riverwatcher system at Daguerre Point Dam demonstrates that hatchery straying 7 into the lower Yuba River occurs. Although no information is presently available 8 regarding the origin of adipose-clipped steelhead observed at the VAKI Riverwatcher 9 system at Daguerre Point Dam, it is reasonable to surmise that they most likely originate 10 from the FRFH.

11 As previously stated, analysis of the VAKI Riverwatcher data indicates that the percent 12 contribution of hatchery-origin adult upstream migrating fish (represented by the 13 percentage of adipose fin-clipped adult steelhead relative to the total number of adult 14 upstream migrating steelhead, because 100% of FRFH-origin steelhead have been 15 marked since 1996) was approximately 43% for the 2010/2011 biological year, and about 16 63% for the 2011/2012 biological year (RMT 2013). If hatchery-origin steelhead stray 17 into the lower Yuba River and interbreed with naturally-spawning Yuba River steelhead, 18 then such interbreeding has been suggested to represent a threat to the genetic diversity 19 and integrity of the naturally-spawning steelhead population in the lower Yuba River. 20 Nonetheless, the question remains regarding the implication of straying of hatchery-21 origin adult steelhead into the lower Yuba River, given past management practices. From 22 1970 to 1979, CDFW annually stocked 27,270-217,378 fingerlings, yearlings, and sub-23 catchable steelhead from Coleman National Fish Hatchery into the lower Yuba River 24 (CDFG 1991a). CDFW stopped stocking steelhead into the lower Yuba River in 1979. 25 In addition, as previously discussed, it is possible that some hatchery-reared juvenile 26 steelhead from the FRFH may move into the lower Yuba River in search of rearing 27 habitat. Some competition for resources with naturally spawned steelhead could occur 28 as a result.

Garza and Pearse (2008) studied populations of *O. mykiss* in the Central Valley using
 molecular genetic techniques to provide insight into population structure in the region.

Genotypes were collected from over 1,600 individual fish from 17 population samples
 and five hatchery rainbow trout strains. Evaluated fish populations included those from
 the McCloud River, Battle Creek, Deer Creek, Butte Creek, Feather River, Yuba River,
 American River, Calaveras River, Stanislaus River and Tuolumne River sub-basins.
 Analyses included fish collected both above and below barriers to anadromy in some of
 the study basins (Garza and Pearse 2008).

7 Phylogeographic trees were used to visually and quantitatively evaluate genetic 8 relationships of Central Valley O. mykiss populations both with each other and with other 9 California populations. Genetic diversity was relatively similar throughout the Central 10 Valley. Above-barrier populations clustered with one another and below-barrier 11 populations are most closely related to populations in far northern California, specifically 12 the genetic groups that include the Eel and Klamath Rivers. Since Eel River origin 13 broodstock were used for many years at Nimbus Hatchery on the American River, it is 14 likely that Eel River genes persist there and have also spread to other basins by migration, 15 and that this is responsible for the clustering of the below-barrier populations with 16 northern California ones. This suggests that the below-barrier populations in this region 17 appear to have been widely introgressed with hatchery fish from out-of-basin broodstock 18 sources. In phylogeographic analyses, above-barrier populations are more similar to San 19 Francisco Bay O. mykiss populations than the below-barrier populations in the Central 20 Valley. Because this relationship is expected for steelhead, given their extraordinary 21 historic dependence on short distance migration events (Pearse and Garza 2007), they 22 may represent relatively non-introgressed historic population genetic structure for the 23 region. Other possible explanations for this pattern that rely on complicated, widespread 24 patterns of introgression with hatchery fish are not entirely ruled out, but are highly 25 improbable given that the above-barrier populations also group with moderate 26 consistency into geographically-consistent clusters (e.g. Yuba-Upper and Feather-Upper) 27 in all analyses and also because of the low apparent reproductive success of hatchery 28 trout in streams throughout California (Garza and Pearse 2008).

The analyses also identified possible heterogeneity between samples from different tributaries of the upper Yuba and Feather Rivers, although linkage disequilibrium was lower in these populations. Linkage disequilibrium can be caused by physical linkage of loci, sampling of related individuals/family structure, and by the sampling of more than
 one genetically distinct group within a population sample (Garza and Pearse 2008).

3 In general, although structure was found, all naturally-spawned O. mykiss populations 4 within the Central Valley Basin were closely related, regardless of whether they were 5 sampled above or below a known barrier to anadromy (Garza and Pearse 2008). This is 6 due to some combination of pre-impoundment historic shared ancestry, downstream 7 migration and, possibly, limited anthropogenic upstream migration. However, lower 8 genetic diversity in above-barrier populations indicates a lack of substantial genetic input 9 upstream and highlights lower effective population sizes for above-barrier populations. 10 The consistent clustering of the above-barrier populations with one another, and their 11 position in the California-wide trees, indicate that they are likely to most accurately 12 represent the ancestral population genetic structure of steelhead in the Central Valley 13 (Garza and Pearse 2008).

The above discussions indicating that below-barrier populations of steelhead in the Central Valley, including the lower Yuba River (particularly in consideration of historic plantings and documented straying) likely do not accurately represent the ancestral population genetic structure. In other words, the current steelhead population in the lower Yuba River likely does not represent a "pure" ancestral genome (RMT 2013).

19 **<u>EXTINCTION RISK</u>**

As stated approximately six years ago by Lindley et al. (2006), there are almost no data with which to assess the status of any of the Central Valley steelhead populations, with the exceptions of the hatchery programs on Battle Creek and the Feather, American and Mokelumne rivers. Therefore, they classified Central Valley steelhead populations, including the lower Yuba River, as data deficient.

According to NMFS (2009a), data are lacking to suggest that the Central Valley steelhead DPS is at low risk of extinction, or that there are viable populations of steelhead anywhere in the DPS. Lindley et al. (2007) stated that even if there were adequate data on the distribution and abundance of steelhead in the Central Valley, approaches for assessing steelhead population and DPS viability might be problematic because the effect of resident *O. mykiss* on the viability of steelhead populations and the DPS is unknown. 1 For the lower Yuba River, the data limitations previously discussed preclude multi-year 2 abundance and trend analyses (RMT 2013). However, continued implementation of the 3 improved VAKI Riverwatcher systems at Daguerre Point Dam is likely to obtain some of 4 the data necessary to allow abundance estimation and productivity evaluation of 5 steelhead in the lower Yuba River (RMT 2013). Moreover, the previous discussion 6 regarding the limited applicability of VSP parameters and extinction risk criteria for spring-run Chinook salmon also pertain to steelhead in the lower Yuba River, in 7 8 consideration of non-independent populations. For additional discussion, see 9 RMT (2013).

10 4.3.7 Public Review Draft Recovery Plan Considerations

The discussion regarding recovery plan implementation provided for spring-run Chinook salmon in Section 4.2.8 of this BA also directly pertains to steelhead in the Yuba River Basin. Therefore, it is not repeated in this section of this BA.

14 4.4 Southern DPS of North American Green Sturgeon

15 The green sturgeon is the most widely distributed member of the sturgeon family 16 Acipenseridae (70 FR 17386). North American green sturgeon are found in rivers from 17 British Columbia south to the Sacramento River, California, and their ocean range is 18 from the Bering Sea to Ensenada, Mexico. In assessing North American green sturgeon 19 status, NMFS determined that two DPSs exist. The northern DPS is made up of known 20 North American green sturgeon spawning (or single stock populations) in the Rogue, 21 Klamath and Eel rivers. In 2005, the southern DPS was believed to contain only a single 22 spawning population in the Sacramento River (70 FR 17386). However, four fertilized 23 green sturgeon eggs collected in 2011 near the Thermalito Afterbay Outlet provide the 24 first documentation of at least some successful spawning in the Feather River (A. 25 Seesholtz, DWR, pers. comm., June 16, 2011).

The Southern DPS of North American green sturgeon (*Acipenser medirostrus*) was listed as a federally threatened species on April 7, 2006 (71 FR 17757) and includes the green sturgeon population spawning in the Sacramento River and utilizing the Sacramento-San Joaquin River Delta, and San Francisco Estuary. NMFS (2009b) *Draft Environmental Assessment for the Proposed Application of Protective Regulations Under Section 4(D)* of the Endangered Species Act for the Threatened Southern Distinct Population Segment of North American Green Sturgeon indicated that the Southern DPS of North American green sturgeon faces several threats to its survival, including the loss of spawning habitat in the upper Sacramento River, and potentially in the Feather and Yuba rivers, due to migration barriers and instream alterations.

8 4.4.1 ESA Listing Status

9 On October 9, 2009, NMFS (74 FR 52300) designated critical habitat for the Southern 10 DPS of North American green sturgeon. This designated critical habitat includes most of 11 the DPS's occupied range, including: (1) coastal marine waters from Monterey Bay to the 12 Washington/Canada border; (2) coastal bays and estuaries in California, Oregon, and 13 Washington; and (3) fresh water rivers in the Central Valley, California. In the Central 14 Valley, critical habitat for green sturgeon includes the Sacramento River, lower Feather 15 River, lower Yuba River, the Sacramento-San Joaquin River Delta, and San Francisco 16 Estuary. NMFS (74 FR 52300) defined specific habitat areas in the Sacramento, Feather, 17 and Yuba rivers in California to include riverine habitat from each river mouth upstream 18 to and including the furthest known site of historic and/or current sighting or capture of 19 North American green sturgeon, as long as the site is still accessible. Critical habitat in 20 the lower Yuba River includes the stream channels to the ordinary high water line 21 extending from the confluence with the mainstem Feather River upstream to Daguerre 22 Point Dam.

Section 4(c)(2) of the ESA requires that NMFS review the status of listed species under its authority at least every five years and determine whether any species should be removed from the list or have its listing status changed. In October 2012, NMFS noticed the initiation of the 5-year status review of the Southern DPS of North American green sturgeon (77 FR 64959).

The purpose of the 5-year review is to ensure the accuracy of the listing classification for the Southern DPS of North American green sturgeon. A 5-year review is based on the best scientific and commercial data available; therefore, NMFS is requesting submission of any such information on the Southern DPS that has become available since the listing determination in 2006. To ensure that the 5-year review is complete and based on the best available scientific and commercial information, NMFS is soliciting new information from the public, governmental agencies, Tribes, the scientific community, industry, environmental entities, and any other interested parties concerning the status of the Southern DPS since the listing determination in 2006 (77 FR 64959).

8 4.4.2 Critical Habitat Designation

9 The essential physical and biological habitat features identified for the Southern DPS of 10 North American green sturgeon include food resources (e.g., benthic invertebrates and 11 small fish), substrate types (i.e., appropriate spawning substrates within freshwater 12 rivers), water flow (particularly in freshwater rivers), water quality, water depth, 13 migratory corridors, and sediment quality. The following summary descriptions of the 14 current conditions of the freshwater PCEs for the Central Valley steelhead DPS were 15 taken from the 2009 NMFS OCAP BO (NMFS 2009a) and the 2009 NMFS Draft 16 Biological and Conference Opinion for the Federal Energy Regulatory Commission's 17 (FERC) Relicensing of the California Department of Water Resources Oroville Facilities 18 (FERC Project No. 2100-134) (NMFS 2009d).

19 4.4.2.1 Primary Constituent Elements

20 FRESHWATER RIVERINE SYSTEMS

21 Food Resources

Abundant food items for larval, juvenile, sub-adult, and adult lifestages should be present in sufficient amounts to sustain growth (larvae, juveniles, and sub-adults) or support basic metabolism (adults). Although specific data is lacking on food resources for green sturgeon within freshwater riverine systems, nutritional studies on white sturgeon suggest that juvenile green sturgeon most likely feed on macro benthic invertebrates, which can include plecoptera (stoneflies), ephemeroptera (mayflies), trichoptera (caddis flies), chironomid (dipteran fly larvae), oligochaetes (tubifex worms) or decapods (crayfish).

1 These food resources are important for juvenile foraging, growth, and development 2 during their downstream migration to the Delta and bays. In addition, sub-adult and adult 3 green sturgeon may forage during their downstream post-spawning migration or on non-4 spawning migrations within freshwater rivers. Sub-adult and adult green sturgeon in 5 freshwater rivers most likely feed on benthic invertebrates similar to those fed on in bays 6 and estuaries, including freshwater shrimp and amphipods. Many of these different 7 invertebrate groups are endemic to and readily available in the Sacramento River from 8 Keswick Dam downstream to the Delta. Heavy hatches of mayflies, caddis flies, and 9 chironomids occur in the upper Sacramento River, indicating that these groups of 10 invertebrates are present in the river system. NMFS anticipates that the aquatic lifestages 11 of these insects (nymphs, larvae) would provide adequate nutritional resources for green 12 sturgeon rearing in the river.

13 SUBSTRATE TYPE OR SIZE

14 Suitable freshwater riverine system habitat includes substrates suitable for egg deposition 15 and development (e.g., cobble, gravel, or bedrock sills and shelves with interstices or 16 irregular surfaces to "collect" eggs and provide protection from predators, and free of 17 excessive silt and debris that could smother eggs during incubation), larval development 18 (e.g., substrates with interstices or voids providing refuge from predators and from high 19 flow conditions), and sub-adults and adult lifestages (e.g., substrates for holding and 20 Stream surveys by USFWS and Reclamation biologists have identified spawning). 21 approximately 54 suitable holes and pools between Keswick Dam and the GCID 22 diversion that would support spawning or holding activities for green sturgeon, based on 23 identified physical criteria. Many of these locations are at the confluences of tributaries 24 with the mainstem Sacramento River or at bend pools. Observations of channel type and 25 substrate compositions during these surveys indicate that appropriate substrate is 26 available in the Sacramento River between Keswick Dam and the GCID diversion. 27 Ongoing surveys are anticipated to further identify river reaches in the upper river with 28 suitable substrate characteristics and their utilization by green sturgeon.

1 WATER FLOW

2 An adequate flow regime (i.e., magnitude, frequency, duration, seasonality, and rate-of-3 change of fresh water discharge over time) is necessary for normal behavior, growth, and 4 survival of all lifestages in the upper Sacramento River. Such a flow regime should 5 include stable and sufficient water flow rates in spawning and rearing reaches to maintain water temperatures within the optimal range for egg, larval, and juvenile survival and 6 7 development (11-19°C) (Cech et al. 2000; Mayfield and Cech 2004; Van Eenennaam et al. 2005; Allen et al. 2006). Sufficient flow is also needed to reduce the incidence of 8 9 fungal infestations of the eggs, and to flush silt and debris from cobble, gravel, and other 10 substrate surfaces to prevent crevices from being filled in and to maintain surfaces for 11 feeding. Successful migration of adult green sturgeon to and from spawning grounds is 12 also dependent on sufficient water flow. Spawning success is more associated with water 13 flow and water temperature than compared with other variables. Spawning in the 14 Sacramento River is believed to be triggered by increases in water flow to about 14,000 15 cfs (Brown 2007). Post-spawning downstream migrations are triggered by increased 16 flows, ranging from 6,150-14,725 cfs in the late summer (Vogel 2005) and greater than 17 3,550 cfs in the winter (Erickson et al. 2002; Benson et al. 2007). The current suitability of these flow requirements is almost entirely dependent on releases from Shasta Dam. 18 19 High winter flows associated with the natural hydrograph do not occur within the section 20 of the river utilized by green sturgeon with the frequency and duration that occurred 21 during pre-dam conditions.

22 WATER QUALITY

23 Adequate water quality, including temperature, salinity, oxygen content, and other 24 chemical characteristics necessary for normal behavior, growth, and viability of all green 25 sturgeon lifestages, is required for the proper functioning of the freshwater habitat. 26 Suitable water temperatures include: (1) stable water temperatures within spawning 27 reaches (wide fluctuations could increase egg mortality or deformities in developing 28 embryos); (2) water temperatures within $51.8-62.6^{\circ}F$ (optimal range = $57.2-60.8^{\circ}F$) in 29 spawning reaches for egg incubation (March-August) (Van Eenennaam et al. 2005); (3) 30 water temperatures below 68°F for larval development (Werner et al. 2007 as cited in

1 NMFS 2009a); and (4) water temperatures below 75.2°F for juveniles (Mayfield and 2 Cech 2004; Allen et al. 2006). Due to the temperature management of the releases from 3 Keswick Dam for winter-run Chinook salmon in the upper Sacramento River, water 4 temperatures in the river reaches utilized currently by green sturgeon appear to be 5 suitable for proper egg development and larval and juvenile rearing. Suitable salinity 6 levels range from fresh water [<3 parts per thousand (ppt)] for larvae and early juveniles 7 [to about 100 days post hatch (dph)] to brackish water (10 ppt) for juveniles prior to their 8 transition to salt water. Prolonged exposure to higher salinities may result in decreased 9 growth and activity levels and even mortality (Allen and Cech 2007). Salinity levels are 10 suitable for green sturgeon in the Sacramento River and freshwater portions of the Delta 11 for early lifestages. Adequate levels of DO are needed to support oxygen consumption 12 by early lifestages (Allen and Cech 2007). Current DO levels in the mainstem 13 Sacramento River are suitable to support the growth and migration of green sturgeon. 14 Suitable water quality also includes water free of contaminants (i.e., pesticides, 15 organochlorines, elevated levels of heavy metals, etc.) that may disrupt normal 16 development of embryonic, larval, and juvenile lifestages of green sturgeon. Legacy 17 contaminants such as mercury still persist in the watershed and pulses of pesticides have 18 been identified in winter storm discharges throughout the Sacramento River Basin.

19 WATER DEPTH

20 Pools of \geq 5 m depth are critical for adult green sturgeon spawning and for summer 21 holding within the Sacramento River. Summer aggregations of green sturgeon are 22 observed in these pools in the upper Sacramento River upstream of the GCID diversion. 23 The significance and purpose of these aggregations are unknown at the present time, 24 although it is likely that they are the result of an intrinsic behavioral characteristic of 25 green sturgeon. Adult green sturgeon in the Klamath and Rogue rivers also occupy deep 26 holding pools for extended periods of time, presumably for feeding, energy conservation, 27 and/or refuge from high water temperatures (Erickson et al. 2002; Benson et al. 2007). 28 As described above, approximately 54 pools with adequate depth have been identified in 29 the Sacramento River upstream of the GCID diversion.

1 MIGRATION CORRIDOR

2 Unobstructed migratory pathways are necessary for passage within riverine habitats and 3 between riverine and estuarine habitats (e.g., an unobstructed river or dammed river that 4 still allows for passage). Unobstructed migratory pathways are necessary for adult green 5 sturgeon to migrate to and from spawning habitats, and for larval and juvenile green 6 sturgeon to migrate downstream from spawning/rearing habitats within freshwater rivers 7 to rearing habitats within the estuaries. Unobstructed passage throughout the Sacramento 8 River up to Keswick Dam (RM 302) is important, because optimal spawning habitats for 9 green sturgeon are believed to be located upstream of the RBDD (RM 242).

10 Green sturgeon adults that migrate upstream during April, May, and June are completely 11 blocked by the ACID diversion dam. Therefore, five miles of spawning habitat are 12 inaccessible upstream of the diversion dam. It is unknown if spawning is occurring in this 13 area. Adults that pass upstream of ACID dam before April are forced to wait six months 14 until the stop logs are pulled before returning downstream to the ocean. Upstream 15 blockage at the ACID diversion dam forces sturgeon to spawn in approximately 12% less 16 habitat between Keswick Dam and RBDD. Newly emerged green sturgeon larvae that 17 hatch upstream of the ACID diversion dam are forced to hold for six months upstream of 18 the dam or pass over it and be subjected to higher velocities and turbulent flow below the 19 dam, thus rendering the larvae and juvenile green sturgeon more susceptible to predation.

Closure of the gates at RBDD from May 15 through September 15 previously precluded
all access to spawning grounds above the dam during that time period. However, as
previously discussed, the RBDD gates were permanently raised in September 2011.

Juvenile green sturgeon first appear in USFWS sampling efforts at RBDD during May, June, and July. Juvenile green sturgeon are likely subjected to the same predation and turbulence stressors caused by RBDD as the juvenile anadromous salmonids, leading to diminished survival through the structure and waters immediately downstream.

27 SEDIMENT QUALITY

28 Sediment should be of the appropriate quality and characteristics necessary for normal 29 behavior, growth, and viability of all lifestages. This includes sediments free of

1 contaminants (e.g., elevated levels of heavy metals such as mercury, copper, zinc, 2 cadmium, and chromium), polycyclic aromatic hydrocarbons, and organochlorine 3 pesticides) that can result in negative effects on any lifestages of green sturgeon. Based 4 on studies of white sturgeon, bioaccumulation of contaminants from feeding on benthic 5 species may negatively affect the growth, reproductive development, and reproductive 6 success of green sturgeon. The Sacramento River and its tributaries have a long history of 7 contaminant exposure from abandoned mines, separation of gold ore from mine tailings 8 using mercury, and agricultural practices with pesticides and fertilizers which result in 9 deposition of these materials in the sediment horizons in the river channel. Disturbance of 10 these sediment horizons by natural or anthropogenic actions can liberate the sequestered 11 contaminants into the river. This is a continuing concern throughout the watershed.

12 ESTUARINE HABITAT AREAS

13 FOOD RESOURCES

14 Abundant food items within estuarine habitats and substrates for adult, sub-adult and 15 juvenile lifestages are required for the proper functioning of this PCE for green sturgeon. 16 Prey species for green sturgeon within bays and estuaries primarily consist of benthic 17 invertebrates and fish, including crangonid shrimp, callianassid shrimp, burrowing 18 thalassinidean shrimp, amphipods, isopods, clams, annelid worms, crabs, sand lances, 19 and anchovies. These prey species are critical for the rearing, foraging, growth, and 20 development of juvenile, sub-adult, and adult green sturgeon within the bays and 21 estuaries. Currently, the estuary provides these food resources, although annual 22 fluctuations in the population levels of these food resources may diminish the 23 contribution of one group to the diet of green sturgeon relative to another food source. 24 The recent spread of the Asian overbite clam has shifted the diet profile of white sturgeon 25 to this invasive species. The overbite clam now makes up a substantial proportion of the 26 white sturgeon's diet in the estuary. NMFS assumes that green sturgeon have also altered 27 their diet to include this new food source, because of its increased prevalence in the 28 benthic invertebrate community.

1 WATER FLOW

2 Within bays and estuaries adjacent to the Sacramento River (i.e., the Sacramento-San 3 Joaquin Delta and the Suisun, San Pablo, and San Francisco bays), sufficient inflow to 4 allow adults to successfully orient to the incoming flow and migrate upstream to 5 spawning grounds is required. Sufficient flows are needed to attract adult green sturgeon 6 to the Sacramento River from the bay and to initiate the upstream spawning migration 7 into the upper river. Currently, flows provide the necessary attraction to green sturgeon to 8 enter the Sacramento River. Nevertheless, these flows are substantially less than those 9 that historically occurred and stimulated the spawning migration.

10 WATER QUALITY

11 Adequate water quality, including temperature, salinity, oxygen content, and other 12 chemical characteristics, is necessary for normal behavior, growth, and viability of all 13 lifestages. Suitable water temperatures for juvenile green sturgeon should be below 75°F. 14 At temperatures above 75.2°F, juvenile green sturgeon exhibit decreased swimming 15 performance (Mayfield and Cech 2004) and increased cellular stress (Allen et al. 2006). 16 Suitable salinities in the estuary range from brackish water (10 ppt) to salt water (33 ppt). 17 Juveniles transitioning from brackish to salt water can tolerate prolonged exposure to salt 18 water salinities, but may exhibit decreased growth and activity levels (Allen and Cech 19 2007), whereas sub-adults and adults tolerate a wide range of salinities (Kelly et al. 2007) 20 as cited in Reclamation 2008). Sub-adult and adult green sturgeon occupy a wide range 21 of DO levels, but may need a minimum DO level of at least 6.54 mg O2/l (Kelly et al. 22 2007 as cited in Reclamation 2008; Moser and Lindley 2007 as cited in Reclamation 23 2008). Suitable water quality also includes water free of contaminants, as described 24 above. In general, water quality in the Delta and estuary meets these criteria, but local 25 areas of the Delta and downstream bays have been identified as having deficiencies. 26 Water quality in the areas such as the Stockton turning basin and Port of Stockton 27 routinely have depletions of DO and episodes of first flush contaminants from the 28 surrounding industrial and urban watershed. Discharges of agricultural drain water have 29 also been implicated in local elevations of pesticides and other related agricultural 30 compounds within the Delta and the tributaries and sloughs feeding into the Delta.

Discharges from petroleum refineries in Suisun and San Pablo Bay have been identified
 as sources of selenium to the local aquatic ecosystem (Linville et al. 2002).

3 WATER DEPTH

4 A diversity of depths is necessary for shelter, foraging, and migration of juvenile, sub-5 adult, and adult lifestages. Sub-adult and adult green sturgeon occupy deep (≥ 5 m) 6 holding pools within bays and estuaries as well as within freshwater rivers. These deep 7 holding pools may be important for feeding and energy conservation, and may serve as 8 thermal refugia for sub-adult and adult green sturgeon (Benson et al. 2007). Tagged 9 adults and sub-adults within the San Francisco Bay estuary primarily occupied waters 10 with depths of less than 10 m, either swimming near the surface or foraging along the 11 bottom (Kelly et al. 2007 as cited in Reclamation 2008). In a study of juvenile green 12 sturgeon in the Delta, relatively large numbers of juveniles were captured primarily in 13 shallow waters from 3 to 8 feet deep, indicating juveniles may require shallower depths 14 for rearing and foraging (Radtke 1966). Thus, a diversity of depths is important to 15 support different lifestages and habitat uses for green sturgeon within estuarine areas.

16 Currently, there is a diversity of water depths found throughout the San Francisco Bay 17 estuary and Delta waterways. Most of the deeper waters, however, are comprised of 18 artificially maintained shipping channels, which do not migrate or fluctuate in response to 19 the hydrology in the estuary in a natural manner. The channels are simplified trapezoidal 20 shapes with little topographical variation along the channel alignment. Shallow waters 21 occur throughout the Delta and San Francisco Bay. Extensive "flats" occur in the lower 22 reaches of the Sacramento and San Joaquin River systems as they leave the Delta region 23 and are even more extensive in Suisun and San Pablo bays. In most of the region, 24 variations in water depth in these shallow water areas occur due to natural processes, with 25 only localized navigation channels being dredged (e.g., the Napa River and Petaluma 26 River channels in San Pablo Bay).

27 MIGRATION CORRIDOR

Within the waterways comprising the Delta and bays downstream of the SacramentoRiver, unobstructed passage is needed for juvenile green sturgeon during the rearing

1 phase of their life cycle. Rearing fish need the ability to freely migrate from the river 2 through the estuarine waterways of the Delta and bays and eventually out into the ocean. 3 Passage within the bays and the Delta is also critical for adults and sub-adults for feeding 4 and summer holding, as well as to access the Sacramento River for their upstream 5 spawning migrations and to make their outmigration back into the ocean. Within bays 6 and estuaries outside of the Delta and the areas comprised by Suisun, San Pablo, and San Francisco bays, unobstructed passage is necessary for adult and sub-adult green sturgeon 7 8 to access feeding areas, holding areas, and thermal refugia, and to ensure passage back 9 out into the ocean. Currently, unobstructed passage has been diminished by human 10 actions in the Delta and bays. The CVP and SWP water projects alter flow patterns in the 11 Delta due to export pumping and create entrainment issues in the Delta at the pumping and fish facilities. 12

13 Power generation facilities in Suisun Bay create risks of entrainment and thermal barriers 14 through their cooling water diversions and discharges. Installation of seasonal barriers in 15 the South Delta and operations of the radial gates in the Delta Cross Channel facilities 16 alter migration corridors available to green sturgeon. Actions such as the hydraulic 17 dredging of ship channels and operations of large ocean going vessels create additional 18 sources of risk to green sturgeon within the estuary. Hydraulic dredging can result in the 19 entrainment of fish into the dredger's hydraulic cutterhead intake. Commercial shipping 20 traffic can result in the loss of fish, particularly adult fish, through ship and propeller 21 strikes.

22 SEDIMENT QUALITY

Sediment quality (i.e., chemical characteristics) is necessary for normal behavior, growth, and viability of all lifestages. This includes sediments free of contaminants (e.g., elevated levels of selenium, polycyclic aromatic hydrocarbons [PAHs], and organochlorine pesticides) that can cause negative effects on all lifestages of green sturgeon (see description of sediment quality for riverine habitats above).

1 4.4.3 Historical Distribution and Abundance

Green sturgeon are widely distributed along the Pacific Coast, have been documented offshore from Ensenada, Mexico, to the Bering Sea, and are found in rivers from British Columbia to the Sacramento River (Moyle 2002). As is the case for most sturgeon, the Southern DPS of North American green sturgeon are anadromous; however, they are the most marine-oriented of the sturgeon species (Moyle 2002).

7 The historical distribution of green sturgeon in the Sacramento-San Joaquin river basins 8 is poorly documented, but Adams et al. (2007) summarizes information that suggests that 9 green sturgeon may have been distributed above the locations of present-day dams on the 10 Sacramento and Feather rivers (Mora et al. 2009). Historical records from the 1930s 11 indicate that green sturgeon were not listed as either "known to occur" or "presumed to 12 occur" in the Yuba or American Rivers (Sumner and Smith 1939; Evermann and 13 Clark 1931).

14 According to NMFS (2009a), spawning populations of green sturgeon in North America 15 are currently found in only three river systems: the Sacramento and Klamath rivers in 16 California and the Rogue River in southern Oregon. Data from commercial trawl 17 fisheries and tagging studies indicate that the green sturgeon occupy ocean waters down 18 to the 110 meter contour (Erickson and Hightower 2007). During the late summer and 19 early fall, sub-adults and non-spawning adult green sturgeon frequently can be found aggregating in estuaries along the Pacific coast (Emmett et al. 1991; Moser and Lindley 20 21 2007 as cited in Reclamation 2008). Particularly large concentrations of green sturgeon 22 from both the northern and southern populations occur in the Columbia River estuary, 23 Willapa Bay, Grays Harbor and Winchester Bay, with smaller aggregations in Humboldt 24 Bay, Tillamook Bay, Nehalem Bay, and San Francisco and San Pablo bays (Emmett et al 25 1991; Moyle et al. 1992 as cited in Reclamation 2008; Beamesderfer et al. 2007). Lindley 26 et al. (2008) reported that green sturgeon make seasonal migratory movements along the 27 west coast of North America, overwintering north of Vancouver Island and south of Cape 28 Spencer, Alaska. Individual fish from the Southern DPS of green sturgeon have been 29 detected in these seasonal aggregations. Information regarding the migration and habitat 30 use of green sturgeon has recently emerged. Lindley (2006 as cited in NMFS 2009a)

1 presented preliminary results of large-scale green sturgeon migration studies, and verified 2 past population structure delineations based on genetic work and found frequent large-3 scale migrations of green sturgeon along the Pacific Coast. This work was further 4 expanded by recent tagging studies of green sturgeon conducted by Erickson and 5 Hightower (2007) and Lindley et al. (2008). To date, the data indicate that green 6 sturgeon are migrating considerable distances up the Pacific Coast into other estuaries, 7 particularly the Columbia River estuary. This information also agrees with the results of 8 previous green sturgeon tagging studies (CDFG 2002), where CDFW tagged a total of 9 233 green sturgeon in the San Pablo Bay estuary between 1954 and 2001. A total of 17 10 tagged fish were recovered: 3 in the Sacramento-San Joaquin Estuary, 2 in the Pacific 11 Ocean off of California, and 12 from commercial fisheries off of the Oregon and 12 Washington coasts. Eight of the 12 commercial fisheries recoveries were in the Columbia 13 River estuary (CDFG 2002).

In the lower Feather River, green sturgeon have intermittently been observed (Beamesderfer et al. 2007). NMFS (2008b) states that the presence of adult, and possibly sub-adult, green sturgeon within the lower Feather River has been confirmed by photographs, anglers' descriptions of fish catches (P. Foley, pers. comm. cited in CDFG 2002), incidental sightings (DWR 2005), and occasional catches of green sturgeon reported by fishing guides (Beamesderfer et al. 2004).

20 In the mid-1970s, green sturgeon were caught each year on the Feather River, with the 21 majority of catches occurring from March to May and a few additional catches occurring 22 in July and August (USFWS 1995). In 1993, seven adult green sturgeon were captured at 23 the Thermalito Afterbay Outlet, ranging in size from 60.9 to more than 73.2 inches 24 (USFWS 1995). In a broad scale survey from 1999 to 2001, green sturgeon were 25 infrequently observed within the area downstream of the Thermalito Afterbay Outlet and 26 none observed upstream (DWR 2003a). In 2006, four green sturgeon were positively 27 identified by DWR biologist near the Thermalito Afterbay Outlet. Eight additional 28 sturgeon were also observed in the same area but could not be positively identified as 29 green sturgeon (DWR 2007a as cited in Reclamation 2008).

1 Although adult green sturgeon occurrence in the Feather River has been previously 2 documented, larval and juvenile green sturgeon have not been collected despite attempts 3 to collect larval and juvenile sturgeon during early spring through summer using rotary 4 screw traps, artificial substrates, and larval nets deployed at multiple locations (Seesholtz 5 et al. 2003). Moreover, unspecific past reports of green sturgeon spawning (Wang, 1986; USFWS 1995; CDFG 2002) have not been corroborated by observations of young fish or 6 7 significant numbers of adults in focused sampling efforts (Niggemeyer and Duster 2003; 8 Seesholz et al. 2003; Beamesderfer et al. 2004). Based on these results, in 2006, NMFS 9 concluded that an effective population of spawning green sturgeon did not exist in the 10 lower Feather River (71 FR 17757). However, four fertilized green sturgeon eggs were 11 collected near the Thermalito Afterbay Outlet on June 14, 2011, thus providing the first 12 documentation of at least some successful spawning in the Feather River (A. Seesholtz, 13 DWR, pers. comm., June 16, 2011).

14 Historical accounts of sturgeon in the Yuba River have been reported by anglers, but 15 these accounts do not specify whether the fish were white or green sturgeon 16 (Beamesderfer et al. 2004). Since the 1970s, numerous surveys of the lower Yuba River 17 downstream of Englebright Dam have been conducted, including annual salmon carcass 18 surveys, snorkel surveys, beach seining, electrofishing, rotary screw trapping, redd 19 surveys, and other monitoring and evaluation activities. Over the many years of these 20 surveys and monitoring of the lower Yuba River, only one confirmed observation of an 21 adult green sturgeon has occurred prior to 2011. The NMFS September 2008 Draft 22 Biological Report, Proposed Designation of Critical Habitat for the Southern Distinct 23 Population Segment of North American Green Sturgeon (NMFS 2008a) states that of the 24 three adult or sub-adult sturgeon observed in the Yuba River below Daguerre Point Dam 25 during 2006, only one was confirmed to be a green sturgeon, and that "Spawning is 26 possible in the river, but has not been confirmed and is less likely to occur in the Yuba 27 River than in the Feather River. No green sturgeon juveniles, larvae, or eggs have been 28 observed in the lower Yuba River to date."

As part of ongoing sturgeon monitoring efforts in the Feather River Basin under the
 AFRP, Cramer Fish Sciences conducted roving underwater video surveys in the lower
 Feather and lower Yuba rivers using a drop-down camera suspended from a motorized

1 boat. On May 24, 25 and 26, 2011, underwater videographic monitoring was conducted 2 in the lower Yuba River downstream of Daguerre Point Dam. Although results are 3 preliminary, a memorandum dated June 7, 2011 Cramer Fish Sciences (2011) stated that 4 they observed what they believed were 4-5 green sturgeon near the center of the channel 5 at the edge of the bubble curtain below Daguerre Point Dam. The sturgeon were 6 observed either on a gravel bar approximately 1.5 m deep, or in a pool approximately 4 m deep immediately adjacent to the gravel bar. Photographs taken by Cramer Fish Sciences 7 8 (2011) were forwarded to green sturgeon experts. Olaf P. Langness, Sturgeon and Smelt 9 Projects, Washington Department of Fish and Wildlife Region 5, expressed the opinion 10 that the photographs were of green (rather than white) sturgeon. Also, David Woodbury, 11 NMFS Sturgeon Recovery Coordinator, expressed his opinion that the fish in the 12 photographs were green sturgeon.

During 2012, underwater videography also was used in an attempt to document the
 presence of green sturgeon downstream of Daguerre Point Dam, but no observations of
 green sturgeon were made.

16 YCWA (2013) examined the potential occurrence of green sturgeon in the lowermost 24 17 miles of the Yuba River based on detections of acoustically-tagged green sturgeon in the 18 Yuba River. The examination included coordination with agencies and organizations 19 involved with green sturgeon research in the Central Valley, and collection of available 20 information and data regarding the presence and use of the Yuba River by green 21 sturgeon. YCWA collaborated with DWR's Feather River Program, the California Fish 22 Tracking Consortium (CFTC), and CDFW's Heritage and Wild Trout and Steelhead 23 Management and Recovery Programs to examine whether any of the acoustically-tagged 24 green sturgeon were found in the lower Yuba River. The CFTC is tracking 217 green 25 sturgeon acoustically tagged in the Central Valley, and DWR's Feather River Program 26 has acoustically tagged 2 green sturgeon in the lower Feather River.

None of the 217 green sturgeon acoustically-tagged in the Central Valley were detected
in the Yuba River, with the exception of one fish tagged by DWR in the Feather River.
This individual fish was detected once on September 6, 2011 in the Yuba River by the
CDFW's lowermost acoustic receiver located at the confluence of the Yuba and Feather

rivers. That fish also was detected upstream in the Feather River earlier on the same day
and downstream in the Sacramento River on the evening of September 6, 2011.
Therefore, the fish apparently only entered the mouth of the lower Yuba River for a very
brief period of time before continuing its downstream migration in the Feather and
Sacramento rivers.

6 4.4.4 General Life History and Habitat Requirements

Limited information regarding green sturgeon distribution, movement and behavioral
patterns, as well as lifestage-specific habitat utilization preferences, is available for the
Sacramento and Feather rivers.

10 **4.4.4.1** Adult Immigration, Holding and Emigration

11 Green sturgeon in the Sacramento River have been documented and studied more widely 12 than they have in either the Feather or the Yuba rivers. Green sturgeon adults in the 13 Sacramento River are reported to begin their upstream spawning migrations into 14 freshwater during late February, before spawning between March and July, with peak 15 spawning believed to occur between April and June (Adams et al. 2002). NMFS (2009) 16 reports that, based on recent data gathered from acoustically tagged adult green sturgeon, 17 these fish migrate upstream during May as far as the mouth of Cow Creek, near Bend 18 Bridge on the Sacramento River.

19 For the Sacramento River, NMFS (2009) reports that adult green sturgeon prefer deep 20 holes (\geq 5 m depth) at the mouths of tributary streams, where they spawn and rest on the 21 bottom. After spawning, the adults hold over in the upper Sacramento River between 22 RBDD and the GCID diversion until November (Klimley 2007). Heublein et al. (2006, 23 2009) reported the presence of adults in the Sacramento River during the spring through 24 the fall into the early winter months, holding in upstream locations before their 25 emigration from the system later in the year. Green sturgeon downstream migration 26 appears to be triggered by increased flows and decreasing water temperatures, and occurs 27 rapidly once initiated (NMFS 2009). Some adult green sturgeon rapidly leave the system 28 following their suspected spawning activity and re-enter the ocean in early summer (Heublein 2006). NMFS (2009) states that green sturgeon larvae and juveniles are
 routinely observed in rotary screw traps at RBDD and the GCID diversion, indicating that
 spawning occurs upstream of both these sites.

Before the studies conducted by UC Davis, there were few empirical observations of green sturgeon movement in the Sacramento River (Heublein et al. 2009). The study by Heublein et al. (2009) is reportedly the first to describe the characteristics of the adult green sturgeon migration in the Sacramento River, and to identify putative regions of spawning habitat, based on the recorded movements of free-swimming adults.

9 The Sacramento River adjacent to the GCID diversion routinely contains a large 10 aggregation of green sturgeon during summer and fall months, although the GCID 11 aggregation site is atypical of over-summering habitats in other systems, being an area of 12 high water velocity (Heublein et al. 2009). The GCID site is over five meters deep, with 13 structural current refuges and eddy formations. It is possible that green sturgeon occupy 14 lower-velocity subsections of the site, although observations of green sturgeon capture, 15 and manual tracking estimates, indicate that green sturgeon are found in, or in very close 16 proximity to, high velocity areas (Heublein et al. 2009).

17 4.4.4.2 Adult Spawning

18 Adult green sturgeon are believed to spawn every two to five years (Beamesderfer et al. 19 2007). Upon maturation of their gonadal tissue, but prior to ovulation or spermiation, the 20 adult fish enter freshwater and migrate upriver to their spawning grounds (NMFS 2009a). 21 Heublein et al. (2009) observed that green sturgeon enter San Francisco Bay in March 22 and April and migrate rapidly up the Sacramento River to the region between GCID and 23 Cow Creek. The fish lingered at these regions at the apex of their migration for 14 to 51 24 days, presumably engaged in spawning behavior, before moving back downriver 25 (Heublein et al. 2009).

To investigate adult immigration, spawning or juvenile nursery habits of green sturgeon in the upper Sacramento River, Brown (2007) developed a study to identify green sturgeon spawning locations and dates in the upper Sacramento River. Using a depth finder, study sites were selected at locations upstream of deeper holes in higher velocity water in the Sacramento River (Brown 2007). The study was originally designed in 1997 using the prevalent methodology at the time (e.g., artificial substrate mats) for the capture of eggs and larvae of white sturgeon. Brown (2007) reports that later findings from artificial spawning and larval rearing of green sturgeon (Van Eenennaam et al. 2001) indicate that green sturgeon eggs may be less adhesive than eggs from other acipenserids, possibly reducing the effectiveness of artificial substrate sampling.

Brown (2007) suggested that spawning in the Sacramento River may occur from April to
June, and that the potential spawning period may extend from late April through July, as
indicated by the rotary screw trap data at the RBDD from 1994 to 2000.

10 Heublein et al. (2009) stated that, in contrast to the behavior of green sturgeon observed 11 during 2004–2005, the majority of out-migrants detected in 2006 displayed an entirely 12 different movement strategy. Nine of the ten tagged fish detected that year exited the 13 system with no extended hold-over period and with no apparent relation to flow increases, eight leaving before July 4th and the last on August 22nd. Heublein et al. (2009) 14 15 suggested that the rapid out-migration of green sturgeon in 2006, and the reduced 16 aggregation period at the GCID site could be a result of consistently higher flows and 17 lower temperatures than in previous study years. Alternatively, this could be an unusual 18 behavior, related to unknown cues, that has not been documented in green sturgeon 19 before this study (Heublein et al. 2009).

The apex detections of individual fish indicate reaches and dates when spawning might have occurred during the study conducted by Heublein et al. (2009). They reported that spawning may have occurred between May and July, and that high water velocities and extensive bedrock habitat were found in all of the apex detection reaches. Furthermore, water temperatures did not exceed 62.6°F in these reaches during this study, which would have permitted normal green sturgeon larval development (Van Eenennaam et al. 2005 as cited in Heublein et al. 2009).

The Sacramento River currently hosts the only known spawning population of green sturgeon (Poytress et al. 2010). During 2009, four spawning sites of green sturgeon were confirmed in the upper Sacramento River (Poytress et al. 2010). Three confirmed sites from 2008 surveys were reconfirmed and one of three newly sampled sites in 2009 was
 confirmed by the presence of green sturgeon eggs on artificial substrate mats.

3 During 2010, five spawning sites of green sturgeon were confirmed within a 60 river 4 kilometer reach of the upper Sacramento River, California (Poytress et al. 2011). As 5 stated by Poytress et al. (2010), spawning events occurred several river kilometers upstream and downstream of the RBDD before and after the June 15th seasonal dam gate 6 7 closure. Spawning occurred directly below RBDD within two weeks after the gate 8 closure. The temporal distribution pattern suggested by 2009 sampling results indicates 9 spawning of Sacramento River green sturgeon occurs from early April through late June (Poytress et al. 2010). Sampling conducted during 2010 suggested that spawning of 10 11 Sacramento River green sturgeon occurs from early April through mid-June (Poytress et 12 al. 2011). During 2010 sampling, depths for eggs collected from all of the sites combined 13 ranged from 2.4 to 10.9 m (7.9 to 35.8 ft) with an average of 6.9 m (22.6 ft). Sacramento 14 River flows and water temperatures at sites located above RBDD during the estimated 15 spawning period ranged from 166 to 459 m3s-1 (5,862 cfs to 16,209 cfs), with an average 16 of 293 m3s-1 (10,347 cfs), and 52.0°F to 57.9°F during the estimated spawning period. 17 Sacramento River flows and temperatures at sites located below RBDD during the 18 estimated spawning period ranged from 268 to 509 m3s-1 (9,464 cfs to 17,975 cfs), with 19 an average of 349 m3s-1 (12,324 cfs), and 52.9°F to 60.1°F during the estimated 20 spawning period (Poytress et al. 2011).

21 The habitat requirements of green sturgeon are not well known. Eggs are likely 22 broadcast and externally fertilized in relatively fast water and probably in depths greater 23 than three meters (Moyle 2002). Preferred spawning substrate is likely large cobble 24 where eggs settle into cracks, but spawning substrate can range from clean sand to 25 bedrock (Moyle 2002). Spawning is believed to occur over substrates ranging from clean 26 sand to bedrock, with preferences for cobble (Emmett et al. 1991; Moyle et al. 1995). 27 Eggs likely adhere to substrates, or settle into crevices between substrates (Van 28 Eenennaam et al. 2001; Deng et al. 2002). Both embryos and larvae exhibited a strong 29 affinity for benthic structure during laboratory studies (Van Eenennaam et al. 2001; Deng 30 et al. 2002; Kynard et al. 2005), and may seek refuge within crevices, but use flatsurfaced substrates for foraging (Nguyen and Crocker 2007 as cited in
 NMFS 2009a).

3 4.4.4.3 Embryo Incubation

4 Green sturgeon larvae hatch from fertilized eggs after approximately 169 hours of 5 incubation at a water temperature of 59°F (Van Eenennaam et al. 2001; Deng et al. 2002), 6 which is similar to the sympatric white sturgeon development rate (176 hours). Van 7 Eenennaam et al. (2005) indicated that an optimum range of water temperatures for egg 8 development was between 57.2°F and 62.6°F. Water temperatures over 73.4°F resulted in 9 100% mortality of fertilized eggs before hatching. Water temperatures above 68°F are 10 reportedly lethal to green sturgeon embryos (Cech et al. 2000; Beamesderfer and Webb 11 2002).

12 Newly hatched green sturgeon are approximately 12.5 to 14.5 mm long. After 13 approximately 10 days, larvae begin feeding and growing rapidly. Green sturgeon larvae 14 do not exhibit the initial pelagic swim-up behavior characteristic of other Acipenseridae. 15 They are strongly oriented to the bottom and exhibit nocturnal activity patterns. Under 16 laboratory conditions, green sturgeon larvae cling to the bottom during the day, and move 17 into the water column at night (Van Eenennaam et al. 2001). After six days, the larvae 18 exhibit nocturnal swim-up activity (Deng et al. 2002) and nocturnal downstream 19 migrational movements (Kynard et al. 2005). Exogenous feeding starts at approximately 20 14 days (23 to 25 mm) (Van Eenennaam et al. 2001).

21 4.4.4.4 Juvenile Rearing

22 Green sturgeon larvae do not exhibit the initial pelagic swim-up behavior characteristic 23 of other *acipenseridae*. They are strongly oriented to the bottom and exhibit nocturnal 24 activity patterns (NMFS 2009a). After 6 days, the larvae exhibit nocturnal swim-up 25 activity (Deng et al. 2002) and nocturnal downstream migrational movements (Kynard et 26 al. 2005). Juvenile fish continue to exhibit nocturnal behavior beyond the metamorphosis 27 from larvae to juvenile stages (NMFS 2009a). Kynard et al. (2005) laboratory studies 28 indicated that juvenile fish continued to migrate downstream at night for the first six 29 months of life. Observations made during nocturnal sampling in the Sacramento River indicate a possible preference of larvae for mid-channel environments or swift water
velocity areas (Poytress et al. 2010). When ambient water temperatures reached 8°C
(46.4°F), downstream migrational behavior diminished and holding behavior increased
(Kynard et al. 2005). This data suggests that 9 to 10 month old fish would hold over in
their natal rivers during the ensuing winter following hatching, but at a location
downstream of their spawning grounds (NMFS 2009a).

Post-migrant larvae are benthic, foraging up- and downstream diurnally with a nocturnal
activity peak (NMFS 2009a). Foraging larvae select open habitat, not structure habitat,
but continue to use cover during the day (NMFS 2009a).

As reported in Corps (2007a), metamorphosis to the juvenile stage is complete at 45 days, and juveniles continue to grow rapidly, reaching 300 mm in one year. Juveniles spend from one to four years in fresh and estuarine waters and disperse into salt water at lengths of 300 to 750 mm (Corps 2007a).

The primary diet for juvenile green sturgeon reportedly consists of small crustaceans, such as amphipods and opossum shrimp (CDFG 2001). As juvenile green sturgeon develop, they reportedly eat a wider variety of benthic invertebrates, including clams, crabs, and shrimp (CDFG 2001).

Green sturgeon juveniles tested under laboratory conditions had optimal bioenergetic
 performance (i.e., growth, food conversion, swimming ability) between 59°F and 66.2°F
 under either full or reduced rations (Mayfield and Cech 2004).

Larvae and juvenile green sturgeon appear to be nocturnal (Cech et al. 2000), which may protect them from downstream displacement (LCFRB 2004). Green sturgeon larvae and juveniles (up to day 84) forage day and night, but activity is reported to peak at night. At day 110 to 118, juvenile green sturgeon move downstream at night, and habitat preference suggests that juveniles prefer deep pools with low light and some rock structure (Kynard et. al. 2005).

Wintering juveniles forage actively at night between dusk and dawn and are inactiveduring the day, seeking the darkest available habitat (Kynard et al. 2005).

Rearing habitat preferences of green sturgeon larvae and juveniles in the Sacramento
 River are poorly understood (Stillwater Sciences 2007). However, additional information
 about habitat use is available for white sturgeon populations, which has been used as a
 proxy for green sturgeon.

5 The seemingly random foraging patterns used by young sturgeon are probably a result of 6 their poor ability to use visual cues to locate and capture food. Juveniles of other species 7 of sturgeon have been shown to be non-visual feeders (Sbikin 1974), and it is generally 8 assumed that most sturgeon use other senses than vision when feeding (Buddington and 9 Christofferson 1985). This means that the success sturgeon have with mobile prey could 10 be dependent on the amount of light available for prey to detect their approach (Utter et 11 al. 1985). A non-visual predatory strategy would be an advantage to sturgeon when 12 feeding on large populations of visually oriented prey species in habitats that are often 13 turbid (Miller 1978, as cited in Utter et al. 1985). A dependence on sensory systems 14 other than vision would also be advantageous when foraging at night or in areas too deep 15 for light penetration. A random searching pattern is characteristic of all ages of juvenile 16 sturgeon that were observed in laboratory and hatchery settings (Utter et al. 1985).

17 Olfactory cues are important for sturgeon when feeding on odorous food types. Sturgeon 18 have large olfactory rosettes with both ciliated and microvillus receptors (Hara 1972, as 19 cited in Utter et al. 1985), and Utter et al. (1985) observed that sturgeon behavior 20 is instantaneously affected by contact with food odors. Sturgeon will often stop after 21 detecting an odor and begin circling the general area in an attempt to contact the food 22 item (Utter et al. 1985).

Tagged adult and subadult green sturgon in the San Francisco Bay estuary primarily occupied waters over shallow depths of less than 10 m, either swimming near the surface or foraging along the bottom (Kelly et al. 2007 as cited in Reclamation 2008). In a study of juvenile green sturgeon in the Delta, relatively large numbers of juveniles were captured primarily in shallow waters from 1–3 m deep, indicating juveniles may require shallower depths for rearing and foraging (Radtke 1966).